Privacy-preserving Copy Number Variation Analysis with Homomorphic Encryption

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- Keywords: Privacy-preserving Genomic Data Processing, Copy Number Variation, Homomorphic Encryption, Applied Cryptography.
- Abstract: Innovative pharma-genomics and personalized medicine services are now possible thanks to the availability for processing and analysis of a large amount of genomic data. Operating on such databases, is possible to test for predisposition to diseases by searching for genomic variants on whole genomes as well as on exomes, which are collections of protein coding regions called exons. Genomic data are therefore shared amongst research institutes, public/private operators, and third parties, creating issues of privacy, ethics, and data protection because genome data are strictly personal and identifying. To prevent damages that could follow a data breach—a likely threat nowadays—and to be compliant with current data protection regulations, genomic data files should be encrypted, and the data processing algorithms should be privacy-preserving. Such a migration is not always feasible: not all operations can be implemented straightforwardly to be privacypreserving; a privacy-preserving version of an algorithm may not be as accurate for the purpose of biomedical analysis as the original; or the privacy-preserving version may not scale up when applied to genomic data processing because of inefficiency in computation time. In this work, we demonstrate that at least for a wellknown genomic data procedure for the analysis of copy number variants called copy number variations (CNV) a privacy-preserving analysis is possible and feasible. Our algorithm relies on Homomorphic Encryption, a cryptographic technique to perform calculations directly on the encrypted data. We test our implementation for performance and reliability, giving evidence that it is practical to study copy number variations and preserve genomic data privacy. Our proof-of-concept application successfully and efficiently searches for a patient's somatic copy number variation changes by comparing the patient gene coverage in the whole exome with a healthy control exome coverage. Since all the genomics data are securely encrypted, the data remain protected even if they are transmitted or shared via an insecure environment like a public cloud. Being this the first study for privacy-preserving copy number variation analysis, we demonstrate the potential of recent Homomorphic Encryption tools in genomic applications.

1 INTRODUCTION

Thanks to the technological revolution brought up by Next Generation Sequencing (NGS), which makes possible to sequence a human genome in hours and for a few hundred USD, national large-scale projects (*e.g.*, the 100,000 Genomes $Project^1$) and private initiatives can now afford sequencing the genomes of hundreds of thousand of individuals. Large digital genomic databases of sequenced raw data and other genomic data (*i.e.*, .bam files) are being created for present and future data analysis and research.

The availability of genomic databases is already paving the way for innovative *genomic medicine*, for instance personalized immunotherapeutic methods for cancer treatment (Kakimi et al., 2017), with unquestionable benefits in terms of quality of future treatments and healthcare. But, storing and processing genomic data also raises serious concerns about privacy rights: genomic data contains extremely sensitive and personal information, for instance about the owner's, his/her relative's, and future kin's susceptibility and predisposition to specific diseases such as Alzheimer's, schizophrenia, and cancer. This is not an hypothetical threat: cases of misuse of genomic data for individual discrimination have already reported

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(Naveed et al., 2015), showing that an extensive availability of genomic databases can have serious ethical and legal concerns.

Several methods have been proposed and studied for genomic data protection. The most common and simplistic is pseudonymization *i.e.*, removing from a genome data file any reference that could link it back to its owner. For genomic data protection, pseudonymization has been proved to be ineffective (Gymrek et al., 2013). Genomic data are rich in information, and retrieve to whom a piece of genomic data belong is easily done by triangulating data.

Another way to protect genomic data is to obfuscate (*i.e.*, filter out) private sensitive sequences of nucleotids from the raw data (Decouchant et al., 2018). The idea is worth, but filtering information affects the quality of processing, even if the filtering is done selectively and in small amount. Besides, changing well-established NGS protocols and machines require time and effort, for instance due to the definition of new standards, which is delaying the availability of this technology.

Yet another option is to encrypt the genomic data before they are stored. This strategy should at least protect the data in case data leakage and as such, promises to be compliant with legal requirements in directives such as the Europe's General Data Protection Regulation (GDPR) and the California Consumer Privacy Act (CCPA). However, data base encryption is insufficient to ensure a long-lasting protection if, for the purpose of processing, encrypted data decrypted before use. Decrypted data are unprotected. It is hard to control that they are not copied and stored unencrypted, or re-transmitted in clear. The risks of privacy violation remains high.

A better solution would be to run the analysis directly on the encrypted data without ever decrypting them. The practices is, at least in theory, possible thanks to secure data processing algorithms. Researchers have explored this possibility and for specific procedures of genomic analysis, mostly about small variants, namely Single-Nucleotide Polymorphisms and indels (i.e., small insertion - deletions). The challenge is that producing a secure (i.e., privacypreserving) version of a specific data processing procedure requires redesigning the algorithm of analysis by using only specific secure functionalities. A privacy-preserving version of an algorithm with comparative and acceptable quality of analysis and performance of the original version is not always possible. And even if it were possible a privacy-preserving variant may remain inefficient, unable to scale up to process large size data as those required for medical purposes.

We study how to realize secure data processing for an important class of genomic data procedures, those aiming at the analysis of copy number variations (see Section 2.1). We assess the performances of this new privacy-preserving processing, and we give evident that a professional implementation can be efficient. To calculate copy number variation values on encrypted genomic data, we use Homomorphic Encryption (HE) (see Section 2.4), a cryptographic technique that enables to operate directly and exclusively on encrypted data and that produce results that are still encrypted. By using HE, data processors never get to know the genomic data they process, nor they can make sense of the results of the processing unless authorized to do so: both genomic data and the copy number variation analysis can be safely outsourced to third parties e.g., research laboratories specialized in that type of analysis or a cloud service with sufficient power of computation for the data processing.

Contribution. We design a proof-of-concept algorithm for the detection of copy number variations that uses HE operations, thus providing the first *privacy-preserving* procedure for CNV analysis which relies on this cryptographic technique. Our algorithm uses basic homomorphic operations (addition, subtraction, multiplication), for which an efficient implementation exists in standard cryptographic software libraries, making it possible to implement a prototype of the procedure.

We test the quality of our privacy-preserving version by looking for somatic copy number variation changes in an exome sample² in comparison to reference exome samples used as control group. These samples come from an existing Melanoma whole exome study (Magi et al., 2013).

The results of our privacy-preserving procedure (once decrypted) are proven compatible with those reported in the study. We also show that we can extend our privacy-preserving procedure to calculate copy number variation values for the whole genome and not only for exomes. Our work reveals the potential of using HE in the analysis of copy number variation over an exome and potentially on the whole genome, and that it is practical if we assume a convenient sampling strategy.

Our proof-of-concept software and the data sets we have used are on git^3 .

²Exomes are collections of protein coding regions of the genome, regions that are singularly called exons.

³github.com/huseyindemirci44/privateCNVanalysis

2 BACKGROUND

Before describing the core of our privacy-preserving procedure, we remind a few basic facts on copy number variation, variant data sets and analysis, and Homomorphic Encryption (HE). We mention relevant works on private genomic data processing that use cryptographic techniques, and we comment their use in copy number variation.

2.1 Copy Number Variation (CNV)

The Human Genome Project, the international scientific research project that had been running from 1990 to 2003 and whose goal was determining the base pairs of the human genome ⁴, has revealed absence and replications of genetic material inside the genome. These structural variants in the number of copies of specific regions are the result of genomic mutations, called deletions and duplications. They cover large regions of the genome and involve several genes.

Copy number variations are structural variants in a person's genome involving more than one thousand bases. They differ from small variants such as indels and repetitions that involves only a few bases. A non-mutated genome region is expected to occour in 2 copies, respectively coming from the chromosomes of the mother and of the father, but because of deletion or duplication, the number of copy of the region may respectively fall down to 1 or 0, or raise up to 3, 4, or more copies. These copy number variations also change the number of genes and gene expression levels inside copy number variation regions. Recent studies claim that around 4.8% - 9.7% of the human genome is copy number variation (Zarrei et al., 2015).

Generally copy number variations do not have a direct phenotypic effect, but elevated copy number variations of particular genes have been shown be associated with specific cancer types (Shlien and Malkin, 2009; Shao et al., 2019). Changes in copy number variation are correlated with increased risk of developing psychiatric disorders such as schizophrenia (Joober and Boksa, 2009; Marshall et al., 2017); with major disease classes including autoimmune, cardio-metabolic, oncologic, and neuropsychiatric diseases (Li et al., 2020); and with Mendelian and complex diseases such as Alzheimer and autism (Zhang et al., 2009).

Needless to say, the interest to find copy number variations and to understand how copy number variation affect to the phenotype is increasing.

2.2 CNV Analysis

It is a common practice to keep and maintain data bases of genomic variants for research. For instance, DECIPHER (Bragin et al., 2014) and "Database of Genomic Variants" (MacDonald et al., 2014) stores copy number variation data for both patient and healthy individuals. Data were usually calculated using traditional methods such as fluorescence *in situ* hybridization (FISH) and array comparative genomic hybridization (aCGH), but nowadays computational methods working on digital genomic data are preferred.

The detection of copy number variation is done using specif algorithms and procedures, and we refer to (Zhang et al., 2019) and (Moreno-Cabrera et al., 2020) for a comprehensive discussion on them. In this work, we refer to one of such methods, the Excavator tool (Magi et al., 2013), whose output values are reported Table 1. They tell about the presence of variations in a genome. Copy number variants can be calculated using read depth of coverage information (Yoon et al., 2009). For instance, in the Excavator tool, the mean read count (*EMRC*) for each exon e in a genomic region is calculated by the following formula:

$$EMRC = \frac{RC}{I}$$

where RC is the number of reads aligned in the region and L is its size in base pairs. *EMRC* calculated from the genome of an individual are then compared with reference values calculated from control genomes, after a normalization step.

Comparing with control genomes is a common method in structural variant analysis. It serves to filter polymorphic variants and to increase statistical significance. Statistical methods, such as Hidden Markov Models, are used to estimate the locations and probability of genomic events. Such a justified need to analyze this class of genomics variants as well as other variants would surely benefit from methods of processing that are privacy-preserving and capable to protect the personal sensitive health related information contained in the genome of patients without compromising the quality of the analysis.

2.3 Private Genomic Data Processing

There has been several studies about using cryptographic technique in ensuring privacy in genomic data processing. *Differential Privacy*, tailored to genomic databases, has been tested to enable privacypreserving Genome Wide Association Study (GWAS) applications (Simmons et al., 2016). Another technique, *Secure Multiparty Computation*, has been

⁴An organism's genome is its complete set of DNA.

Copy Number	Explanation	Notation
Double Gain	\geq 2 extra copies are duplicated	2
One copy gain	Extra 1 copy is obtained	1
Normal case	2 copies exist	0
Heterozygous deletion	One of the copies is lost	-1
Homozygous deletion	There is no DNA sequence	-2

Table 1: Copy Number Variation.

tested in "trio exome analysis" for small cohorts of patients, and demonstrated to be executable in seconds whereas the remaining variant set of any patient remained privately protected (Jagadeesh et al., 2017; Akgün et al., 2020). These methods are not directly applicable to copy number variation analysis, because the detection of variants is generally out of their working pipeline, which focus on Single-Nucleotide Polymorphisms and short indels. Instead, Homomorphic Encryption (HE) has been applied to realize a secure exact logistic regression model for rare disease GWAS studies (Wang et al., 2016), but to our best knowledge at the moment of writing⁵, there is no specific work that studies how to protect genomic privacy in copy number variation detection, nor one that proposes using HE for this task. Once copy number variation values are calculated in a private manner and stored (even in encrypted form) in a .vcf file, all the cryptographic methods previously mentioned can be employed to ensure privacy protection in further data analysis, tests, and processing.

2.4 Homomorphic Encryption

This cryptographic technique can be considered as a bridge (*i.e.*, an homomorphism) between operations on a plaintext (unencrypted) domain and operations on an encrypted domain. This means that HE makes it possible to perform computations directly on the encrypted. The output of the computation, which is encrypted, once decrypted, is the same as the output that would have been produced if the computation were performed on the unencrypted data.

For example, an additive HE scheme would ensure that $Enc_k(d + d') = Enc_k(d) \oplus Enc_k(d')$, where Encis the encryption function, k encryption key, \oplus the homomorphic addition, and d and d' pieces of data.

HE enables outsourcing the computation of personal data while ensuring privacy. A subject that has encrypted some personal data by using its public key can send it safely to an insecure party to get them processed. The party processes the data without ever getting access to the plain data and returns the encrypted results back to the subject who is the only one who can decrypt them by using the private key. Even if the data breach were breached on the party side, confidentiality is ensured because the data cannot be decrypted without the private key.

For reasons of space, we have to limit our account of HE schemes. The reader can refer to (Acar et al., 2018) for a comprehensive survey on their theory and implementations, but for the scope of this work we point out that there are different types of HE schemes. The most common ones are the following:

- *Partially Homomorphic Encryption*. They preserve the homomorphism only for computations using one operation, for example only addition, or only multiplication.
- Somewhat Homomorphic Encryption. They preserve homomorphism for bounded computations, for instance computation using a specific number of additions and multiplications.
- *Fully Homomorphic Encryption (FHE)*. They are schemes that preserve the homomorphism for arbitrary computations, for instance, computations with an arbitrary number of additions and multiplications.

The latter is the most general, and a proof for its existence for arbitrary computation using addition and multiplication has been presented in 2009 by Gentry who used mathematical lattices for the purpose (Gentry, 2009). Most of the HE schemes of today also rely on solutions for hard-to-solve computational algebraic problems over lattices, known as Ring Learning with Errors (Regev, 2009). Recently, libraries such as Microsoft's SEAL⁶ and IBM's Fully Homomorphic Encryption Toolkit For Linux⁷ have been made available for developing HE applications.

3 REFERENCE SCENARIO

To contextualize the copy number variation (CNV) analysis with HE, let us refer to the following situation. A *patient*, who had his genome sequenced and

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⁶https://github.com/microsoft/SEAL

⁷https://github.com/IBM/fhe-toolkit-linux



genomic data coverage files stored in a biobank, consults a doctor. The doctor suggests a test which requires identifying copy number variants because he intends to investigate changes in the tumor suppressor genes like TP53 for a possible drug resistance analysis. This could ideally be carried out by the same biobank by comparing the patient's tumor sequence coverage data with those other patients used as a control set.

Since both the patient's and other patients' genomic data, in this example the sequence coverage files, are personal data, such data are stored encrypted by the biobank.

Figure 1 shows a possible secure processing protocol. The message exchange involves the following agents: a *Trusted Party* (TP), usually an authority such as the Public Health System Institute or a national Certified Institution, which is in charge of creating a pair of encryption-decryption keys (pk, sk), respectively, public and private *meant to be used in support to secure genomic processing*; the *Medical Unit* (MU), usually a doctor or a researcher in the need of processing genomic data; the *Storage and Processing Unit* (SPU), which is generally represented by a public cloud service for instance run by a biobank; the *patients*, which are the data subjects of the genomic data, which herein we assume to be a particular patient's genomic data (P_0 and g_0), and the genomic data files of a cohort of reference patients (\vec{P} and \vec{g}).

Figure 1 protocol exchange is as follows: Before storing their genomic data on the clouds, the patients $(P_0, \text{ as well as } \vec{P})$, request (message 1) and obtain the public key *pk* from TP, using which they encrypt their genomic data, respectively g_0 and \vec{g} . The encrypted data, resp. $Enc_{pk}(g_0)$, $Enc_{pk}(\vec{g})$, are send to the SPU (message 2), which stores them for future uses.

When a MU, having agreed with patient P_0 to make a CNV analysis for which it needs the patient's genome g_0 and/or those of the cohort (\vec{g}), makes a request for the analysis to the SPU (message 3). The SPU performs on behalf of MU the calculation by running the secure homomorphic encryption version of copy number variation algorithm (see later, Algorithm 1) on the encrypted genomic data previously stored. The SPU sends back to the MU the encrypted outcome of the analysis, $Enc_{pk}(CNV(g_0, \vec{g}))$, which cannot yet be decrypted. MU requests for and obtains the decryption key from the TP (message 6), after which MU gets to know the result of the analysis.

None of the agents but the data subjects even know the patients' personal genomic data in cleartext. It is important to stress that the protocol in Figure 1 does not protect against active intruders, such as an SPU that tries to fool TP into getting the private key. Additional measures need to be in place for reaching higher levels of security, which is a goal out of the scope of this work. The protocol is meant to ensure data privacy against a passive but curious SPU, for which the use of privacy-preserving copy number variation is an adequate measure.

4 A PRIVACY-PRESERVING CNV

We now present our implementation of the secure CNV detection that relies on HE. The pseudocode is reported in Algorithm 1.

For simplicity of exposition, we assume that all the computations are performed locally in one machine instead of being distributed as they should be according to Figure 1. Consequence of this simplification is that we do distinguish between different roles, not we show the interroles communication. The feasibility and performance benchmarks that we obtain from the experiment remain valid because we are not interested in measuring the overhead due to the communication but only the impact of the HE computation for a CNV analysis on genomic data.

The procedure PPCNV is inspired by the procedure we described in Section 2. It takes in input the *coverage files*⁸ for patient and control cases, respectively, *CovPatient* and *CovControl*, and the list of genes *Genelist* to point out the regions to be analyzed. It returns the list *CNV* of copy number predictions for the genes in *Genelist*. The procedure refers to certain threshold parameters (th_i in the pseudocode), which represent the boundary conditions to have 0, 1, 2, 3, 4 or more copies of DNA parts.

The rationale of procedure PPCNV is as follows (we assume basic knowledge about coverage files, exomes, and genes): the patient's gene coverage file, which contains the number of occurrences of each gene on the different reads (i.e., overlapping fragments) of the patient's genome after sequencing, is compared with a control coverage file of an healthy individual. The number of copy variants for a gene g is determined looking at the difference between the coverage files. For instance if the difference is zero, then there is no variants; if it is bigger than 2 there are more than 2 copy variants. Intermediate values are determined reasoning on the value of the difference: if it is bigger then 1.5, there are quite likely 2 copy variants, whereas if it is bigger then 0.5, there is quite likely 1 copy variation. We use a window of five

⁸Coverage files contain information regarding the base/read coverage of each sequence in an assembly file.

overlapping consecutive gene regions, and we take the value that occurs in the majority of the cases as the final copy variant value. The key point of PPCNV is to implement this ideas using homomorphic operations, namely \otimes and \oplus (and therefore \ominus). There is no homomorphic operator that can be used to determine whether *e.g.*, (x-y) > 1.5. The idea is then to rewrite the expression as $x - (1.5 \times y)$, which can be calculated homomorphically. That done, we can encrypt the coverage files and calculate $x \ominus (1.5 \otimes y)$ for all genes, keeping the sensitive coverage files protected.

Algorithm 1: Privacy-preserving CNV Procedure.

PP	CNV(<i>CovPatient</i> , <i>CovControl</i> , <i>Genelist</i>)
1	
2	// SPU calculation
3	forall $g \in Genelist$
4	for $i = 0$ to 4 do
5	$c_{g_i} := Enc_{pk}(th_i \otimes CovControl_g);$
6	for $i = 0$ to 3 do
7	$d_{g_i} := c_{g_0} \ominus c_{g_{i+1}}$
8	return $(d_{g_0}, d_{g_1}, d_{g_2}, d_{g_3})$
9	
10	// MU calculation
11	forall $g \in Genelist$
12	for $i = 0$ to 3 do
13	$d_{g_i} := Dec_{sk}(d_{g_i})$
14	$n_g := 0;$
15	if $d_{g_0} > 0$ then $n_g := 2$
16	else if $d_{g_1} > 0$ then $n_g := 1$
17	else if $d_{g_2}^{"} < 0$ then $n_g^" := -1$
18	else if $d_{g_3} < 0$ then $n_g := -2$;
19	$n_x :=$
20	$maxCount(\{n_g, n_{g+1}, n_{g+2}, n_{g+3}, n_{g+4}\})$
21	$CNV_g := n_x$
22	return CNV _g

In Algorithm 1 *Enc* and *Dec* represent HE encryption and decryption operations with the public and private keys. Values th_1, th_2, th_3, th_4 are the HE threshold parameters where $th_1 \approx 2.$, $th_2 \approx 1.5$, $th_3 \approx 0.5$ and $th_4 \approx 0$. The threshold numbers represent the boundary conditions to have 0, 1, 2, 3, 4 or more copies of DNA parts. After obtaining the initial copy number variant predictions, we select the prediction value that occurs with maximal frequency (*i.e.*, function *maxCount*) over a non-overlapping window of length 5 to filter out instant fluctuations since the copy number variation values do not change frequently. The most frequent element in the window is estimated as the copy number variation value for that window.

In Algorithm 1, we made another simplification, this time with consequences. The final calculation of copy number variation values is estimated after certain intermediate values, d_g s in the code, have been decrypted (step 11–13). This means that the SPU has to send the d_g values back to the MU, and it will MU that finalizes the calculation.

This simplification is motivated because implementing homorphically the test '> 0' (lines 16–18) is not straightforward and it would introduce a computational burden that seems unnecessary. The genomic data to protect are the coverage files, while the encrypted values d_g , once sent back to the MU, who decrypts them, do not disclose, to MU, more information then the values CNV_g , which MU gets to know eventually.

The current implementation runs efficiently because we implemented HE encryption and decryption using natif3 operations for vectors. The comparison for the list of whole exome genes is calculated using 4 vector encryption-decryption pairs and the required intersections conveniently as described in Algorithm 1. In this way, we could label all the gene coverages to a CNV value using the threshold parameters. The actual values of the parameters can be adjusted according to the coverage settings, tumor/normal sample ratio, including the normalization step for the samples.

5 EXPERIMENTAL SETTING, DATA SETS, AND LIBRARIES

To test the feasibility of this CNV query using HE we have set up the following experiment setting. We considered a publicly available Melanoma tumor sample data and compared it with a healthy control individual. We have calculated the mean read count (*GMRC*) per gene as follows:

$$GMRC = \frac{RC}{L}$$

where *RC* is the number of reads aligned in the gene, L is the size of the gene in base pairs. We use GMRC values to make a comparison for case and control cases. First, we compare GMRC values for a single gene such as TP53 or PTEN. Later we have used the same comparison query for a list consisting of 100 genes, representing a gene panel. Then we have carried out analysis over the chromosome level. We have used the whole list of 1228 genes in Chromosome 9. This chromosome is chosen since it contains CDKN2A gene which is highly damaged in Melanoma cancer (Pfarr et al., 2016). Finally, we have conducted this experiment on the whole set of Refseq curated list of genes, consisting of 23.882 genes out of 31.848 locations where both the samples have coverage greater than 5 (Supplementary Files:

geneslist.txt, allgeneshealthy.csv, allgenespatient.csv, genes_Ch9_healthy.csv genes_Ch9_patient.csv). We have measured the performance of the encryption and decryption processes for these queries. We have compared our CNV results for Chromosome 9 with the original study (Magi et al., 2013) to analyze the accuracy of the secure CNV detection algorithm.

The data we have used in this paper has been taken from the samples that has been used in the Excavator CNV detection tool (Magi et al., 2013). The data consists of a tumor whole exome sequence from a Melanoma patient (Melanoma₀₁) and a control healthy individual $(Sample_{01})$. We have uploaded the .bam files of the samples with accession number ERR174231 and ERR174237 which were aligned according to Genome Reference Consortium Human Build 37 (GRCh37, hg19) from the Sequence Read Archive (SRA) database (Leinonen et al., 2010) to the Galaxy web platform. We have used the public server usegalaxy.org (Afgan et al., 2018) to calculate the coverage values with .bed tools. Then using these base read coverage values, we calculate the read counts per gene (GMRC) as described above, which provide the input of Algorithm 1.

The HE operations throughout this work are executed with the help of Pyfhel: PYthon For HE Libraries⁹ which uses the SEAL (Chen et al., 2017) library as a backend. Pyfhel provides vector and fraction encryption/decryption capabilities which were especially helpful while performing operations on the list of genes. We note that the available implementations of HE only allow simple operations such as addition, subtraction, and multiplications.

6 **RESULTS**

Using Algorithm 1, we have compared the coverage values gathered from two .bam files. Figure 2 depicts the copy number variant estimations for Chromosome 9 obtained by comparing the Melanoma tumor file with the healthy sample file which indicates double copy gains at the beginning and end of Chromosome 9, and two double deletion regions. We also detect one copy gains at certain points. We check these results against the CNV values reported in the original study (Magi et al., 2013). Among these variants, 29 of the 30 double deletion genes are the same with (Magi et al., 2013). Our algorithm has predicted the CNV region a little larger so that we have the *MTAP* gene in the boundary. In particular we are able to detect the double loss of CNV in the gene *CDKN2A* which is as-

⁹https://github.com/ibarrond/Pyfhel

sociated with Melanoma cancer. For double copy gain genes, 22 of the 65 do not appear in the Excavator list. This comes from the fact that Excavator algorithm did not report the variations at the end of Chromosome 9. Both works predicted no one copy losses wherever our algorithm detected 50 one copy gains where 19 of them were reported as double gain in Excavator list. Excavator did not identify any one copy gains or losses. Table 2 presents the ratio of genes that are consistent with the results of the (Magi et al., 2013) tool.

The whole list of genes and their estimated CNVs for Chromosome 9 are presented in the Supplementary File (Chromosome9_Estimated_CNV.csv). It is an expected phenomena that two CNV detection algorithms may slightly differ since they have different sensitivity to parameters such as depth of the sequencing and they use different statistical models (Zhang et al., 2019). Nevertheless, the comparison results demonstrate the usability of a HE based CNV prediction. We emphasize that our aim is not to improve the performance of CNV detection but to demonstrate the feasibility of a secure version of CNV analysis using existing HE framework.

Computational Performances. The time performance of the HE experiment is provided in Table 3. These experiments are carried out with a laptop with i-7 1.8- 2.3 GHz. CPU and 16 GB RAM with 64 bit Windows Enterprise operating system. We used Python version 3.7.4 on the Spyder Environment. We note that time performance depends on the HE parameters. For a single gene value the calculations can be executed using Pyfhels's homomorphic operations for fraction. For larger gene lists, vector operations can be used. To have a decryption without noise, the vector parameter defined by *m* must be larger than our query list. Figure 3 represents the performance of the encryption, decryption and total time processes



Figure 2: Copy Number Estimations for Chromosome 9.

which are collected while encrypting a list of 1000 genes with respect to the HE parameter m. The graph demonstrates a linear increase for encryption and decryption times according to the parameter m. For a fixed vector size *m*, the execution times do not change with the length of the gene list since actually the algorithm implements encryption and decryption operations along the vector size. In the worst case where we choose the vector size m = 32.768 to ensure the inclusion of the whole exome gene list in the encryption array, the results are still in the order of deciseconds. In Table 3 only the time performance of the HE operations are demonstrated. The preprocess required to obtain the read count tables have not been considered since, this is a standard step in any CNV analysis without an encryption. Our aim is to measure the workload of the computations executed in the encrypted domain.

Table 2: Comparison of CNVs for Chromosome 9 with (Magi et al., 2013).

CNV Detected	Number of genes	Overlap ratio*
2	65	0.66
1	50	0.38**
0	841	0.90
-1	0	1
-2	30	0.96

* The ratio of genes that exist in (Magi et al., 2013) ** 19 of the 50 Double Gain genes are found in the in One Copy



Figure 3: Time performance according to the encryption parameter m.

7 DISCUSSION

What we have exposed so far shows that is feasible to code a CNV analysis algorithm using HE, that represents a privacy-preserving version of CNV detection. This task is greatly simplified thanks to software libraries that offer reliable and optimized implementations of HE operations such as vector encryption

Analysis Region	Number	of	HE parameters	Execution
	Locations			Time (Sec.)
Single Gene	1		p = 65.537, m = 1.024	0.0005
Gene Panel	100		p = 65.537, m = 1.024	0.0035
Chromosome 9	1.289		p = 65.537, m = 2.048	0.0088
Whole Exome	23.882		p = 65.537, m = 32.768	0.3448
Whole Genome**	320.00		p = 65.537, m = 32.768	4.7972

Table 3: Time performance of the HE experiments. The Execution Time includes the Encryption + Decryption Time

** Estimated Time from whole exome study

and decryption, and that they already solve otherwise time-consuming and tricky technical issues *e.g.*, bootstrapping strategies to control noise in the scheme.

Our experiments show that the execution is efficient, that is within seconds. We estimate that it remains efficient when extended to processing a whole genome.

We also demonstrated that this privacy-preserving version of the algorithm performs a meaningful analysis of copy number variants. This has been demonstrated initially by comparing two coverage files for certain genomic positions or a gene panel, and then extending the approach to process the whole exome level. We speculate that it can be further extended to a whole genome analysis, if we apply a coverage comparison strategy. For instance, 10% of the whole genome can be covered with a window size of 1000 bases, which will result around 320.000 coverage regions to compare which is of order 10-15 times larger magnitude compared to the whole exome study. Since copy number variation values do not change frequently through the genome, we may identify the values with good precision by sampling 10% of the genome. If this speculation is correct, the whole genome CNV analysis can still be conducted in seconds.

In our use case, we have taken a healthy exome sequence as control to simulate somatic CNV detection. For a standard CNV analysis, the number of samples can be increased to improve statistical significance. In this case, the execution time will increase linearly with the number of samples. Notably, this step is completely parallelizable where we can use the computational power of the public cloud environment. Since the encryption-decryption processes are fast, the total time performance still remains feasible for real time applications.

An alternative strategy to direct comparison of genomes is calculating the mean of the copy number variation values over the control samples as a base line and use this base line as a reference for CNV detection. The required averaging computation can be done in the encrypted domain and needs to be executed only once in the offline mode. Then comparing each genome coverage with this average control baseline will have the similar performance with this work.

The results demonstrate that the existing libraries of HE enable real time analysis for small files like coverage or .vcf files. However, a complete genomic analysis pipeline requires to perform operations on raw file types such as .sam, .bam or .fastq. At the moment performing tasks like genome alignment on encrypted data is not feasible since there are yet no efficient algorithms for operations like comparison of two strings in the encrypted domain. Moreover, whenever there are conditional statements, the HE implementation slows down substantially and the data size expands as a result of re-linearization process in encryption. Therefore, it is crucial to define an algorithm with the most basic operations.

Security Assumptions and Parameters. In the proposed protocol we have deployed a Trusted Party for the generation of encryption and decryption keys. TP as a certified and controlled institute, could access genomic data but has not storage and processing capabilities. These tasks are executed at the storage and processing unit, SPU. The SPU is not reliable. A passive malicious adversary who has somehow access to the SPU (the admin of SPU, for instance), can not obtain sensitive data since the security of the HE is guaranteed by the Public Key encryption system. The implemented HE algorithm in Pyfhel library is the BGV somewhat homomorphic encryption scheme without any bootstrapping¹⁰. This method also depends on the Learning with Errors over Rings (RLWE) problem and we refer the interested reader to the paper (Brakerski et al., 2014) for mathematical details of the algorithm. We have chosen a security level equivalent to 128-bit symmetric (AES) encryption according to the suggestions in (Albrecht et al., 2018). The parameter p is the integer modulus and m is the vector size where the Ring operations are defined. The private and public keys of the Homomorphic Encryption system are generated according to the specifications of the scheme, where public key is a secret Ring el-

¹⁰Personal communication with the developers.

ement and private key is a pair of two Ring elements which satisfy certain criteria, respectively.

We note that the requirement of a Trusted Party is inevitable for this kind of study, since for the derivation of the genome sequence at least, there needs to be a certified trusted authority. We note that previous related work also followed this assumption (see (Wang et al., 2016; Ayday et al., 2013)).

Limitations. We limited our HE to simple arithmetic operations in our redesigned CNV algorithm, leaving to the MU (*i.e.*, the doctor) the task to run inequality tests on the plain data. We argued that this choice does not affect security, but it would be interesting to measure the performance of a full HE version of CNV. We leave it for future work.

An obvious limitation of this work is the number of cases analyzed. As we intended to prove that a copy number variation analysis using HE is feasible, we chose as example the analysis of a single exome. We plan to extend this work to several exoms and whole genome samples to have a better evaluation of accuracy and fine tuning of the threshold parameters in future work.

Another issue is the normalization of sequencing data coming from various runs and technologies. Comparison of data obtained from different sequencing technologies or parameters is a challenging process. Tumor-normal sampling ratio is also of critical importance while working with tumor data. A careful normalization is required before data analysis for accurate CNV results.

8 CONCLUSIONS

In this work we have presented a proof-of-concept study for the estimation of copy number variation values by implementing a secure coverage comparison using Homomorphic Encryption (HE).

We are able to calculate the copy number values for a predefined list of genes privately without revealing personal genomic information. We have worked on a somatic diagnosis use case with a Melanoma whole exome data and compared the copy number variation results with the original study (Magi et al., 2013). Our results demonstrate the practicality and accuracy of a privacy-preserving copy number variation (CNV) analysis at the whole exome and whole genome levels.

As CNV analysis takes a broader deployment in genomic pipelines, making it possible to perform in a privacy preserving manner in compliance with data protection regulations is becoming important. This study is the first work focusing on this important class of variants which have been neglected up to now.

As a future work, we plan to apply the proposed CNV method to a set of whole genome sequences. This will satisfy a better evaluation for the validity of the sampling strategy and its precision on large whole genome cohorts and more accurate determination of the algorithm parameters.

Another upcoming line of research is the use of HE for privacy-preserving .vcf queries to execute applications such as private rare mutation discovery, variant prioritization for causes of disease, and disease predisposition calculation. Although there exist individual examples, there is still work to do for developing more efficient and unified approaches. Recent HE libraries enable various types of genomic applications which can now be executed in real time hindering mathematical and implementation difficulties and opening the door for the age of privacy-preserving personalized medicine.

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