# A Survey on *Aspergillus chevalieri* Infection Isolated from Poultry Feed

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Abstract: Population and characteristics Aspergillus chevalieri in finished poultry feed were investigated. The aim was

to isolate and enumerate *Aspergillus chevalieri* from finished poultry feed. Thirteen samples poultry feed of chick starter, broiler and layer, sold by retailers in traditional markets were collected. Moisture content of the feeds was determined by oven dried method. Fungal population was analyzed using dichloran 18% glycerol agar (DG18) medium. All fungal colonies were isolated and incubated in potato dextrose agar slants (7 days, 29 °C). Morphological characteristics were observed. Results showed that chick starter has the highest moisture content (10%) and the most infected by *Aspergillus chevalieri* (31.33×10<sup>4</sup> cfu g<sup>-1</sup>) followed by

broiler (0.33×10<sup>3</sup> cfu g<sup>-1</sup>) and none in layer.

### 1 INTRODUCTION

The occurence of fungal infection on poultry feed in tropical countries become the primary cause deterioration during storage. High rainfall and relative humidity promote improper stored feed absorb water favour from the environment. This process leads to an increase moisture content exceeds critical value for fungal growth and it may contaminated by mycotoxins (JECFA, 1998). Maize, soybean, rice bran, peanut and by product that major of the component are susceptible infected by molds (Okoly et al. 2007; Okun et al. 2015). Among fungal genera, Aspergillus, Fusarium, Rhizopus, Penicillium and Mucor were the most common found contaminating in poultry feed (Krnjaja et al. 2008; Kana et al. 2013; Nemati et al. 2014; Kehinde et al. 2014). Infection of the molds deteriorate feeds and reduce its nutritional componds. As xerophilic fungi, A. chevaliery is able to grow on low water activity. The presence of A.chevalieri on poultry feed might occur when the fungus colonize the raw materials during harvesting, grow in it during postharvest handlings such as drying, transportation and storage. The objective of this research was designed to survey Aspergillus chevalieri infection and isolated from

finished poultry feed (chick stater feed, broiler and layer feed) sold by retailers in traditional markets.

#### 2 MATERIALS AND METHODS

## 2.1 Sample Collection

A total of 13 composite samples of finished poultry feed: chick starter, broiler and layer, were collected (500 g each sample) during the dry season (months of May and June 2018) from different retailers in Medan, North Sumatera, Indonesia. Each sample was packed separately in sterilized polythylene bag and stored (-4 °C) in refrigerator for further use.

# 2.2 Fungal Population and Identification

Population of *A. chevalieri* was determined by dilution method in dichloran 18% glycerol agar (DG18 medium) according to Pitt and Hocking (2009). Twenty five gram of each sample in 500 mL erlenmeyer was diluted in 250 mL sterilized distilled water. The suspension was homogenized using shaker (Gallenkamp SG92-02-311, England) 100 rpm for 2 minutes. Four dilutions, 10<sup>-1</sup>, 10<sup>-2</sup>, 10<sup>-3</sup> and 10<sup>-4</sup> were

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made. From the 10<sup>-3</sup> or 10<sup>-4</sup> dilution, 1 mL was transfered to petridish (diameter 9 cm) and pour plated in DG18 medium. Each sample was cultured triplicates. The plates were incubated at at 29±2°C for 6 days. All colonies were counted as colony forming unit (cfu g<sup>-1</sup>) of the sample and identified according to Pitt and Hocking (2009). Macroscopic and microscopic identification were conducted under Zeiss Prima Star 37081 light microscope, Gottingen, Germany.

#### 2.3 Determination of Moisture Content

For determination of moisture content, every 50 g of sample was put on an aluminum foil dish and dried in an oven at 130 °C for 2 hours and reweighed (BSI, 1980). Three replicates were made for each sample.

#### 3 RESULTS AND DISCUSSION

All finished poultry feed sold by retailers commonly packaged in polypropylene bag and stored in open air. Moisture content of the poultry feed are presented in Figure 1.

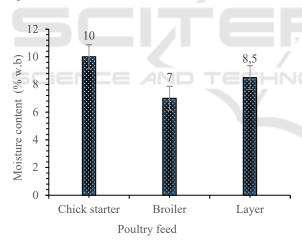


Figure 1: Moisture content (% wet basis) of finished poultry feed (chick starter, broiler and layer feed) collected from retailers at traditional markets in Medan, North Sumatera.

Chick starter has the highest moisture content (10%) followed by layer (8.5%) and broiler (7%). Feed moisture content determine *A. chevalieri* population. Most of the colony was found in chick starter (5.49 cfu g<sup>-1</sup>) followed by broiler (2.52 cfu g<sup>-1</sup>). None *A. chevalieri* was found on layer (Figure 2).

Previous study by Saleemi et al. (2010) and Sivakumar et al. (2014) reported that *Aspergillus* was the most common on poultry feed. The most

prevalence of *A. chevalieri* on poultry feed was reported by Greco et al. (2014).

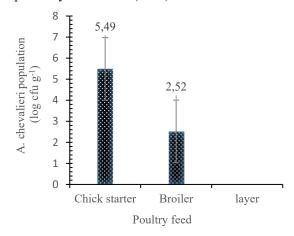


Figure 2: Fungal population (cfu g<sup>-1</sup>) isolated from finished poultry feed (chick starter, broiler and layer) after 6 days incubation (29±2 °C) on dichloran 18% glycerol agar (DG18) medium.

Poultry feed raw materials such as corn, rice bran, peanut and soybean are vulnerable infected by *Aspergillus* [Okun et al. 2015; Krnjaja et al. 2008). We assumed the infection of the molds occurred on poultry raw materials during harvest and their population increase during storage.

#### 4 CONCLUSION

All finished poultry feed sold by retailers in traditional markets were infected by *A. chevalieri*. The presence of the xerophilic mold potential deterioration on poultry feed during storage. Prevention fungal growth and deterioration were required during storage of the feed.

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#### REFERENCES

JECFA, (1998). Aflatoxins, Safety Evaluation of Certain Food Additives and Contaminants. WHOOkoly, IC. Ogbuewu, PI. Uchegbu, MC. Opara, MN. Okorie, JO. Omede AA. Okoli, GC. Ibekwe, PI.

- (2007). Assessment of mycoflora of poutry feed raw materials in humid tropical environment. *J. American Sci.* 3: 5-9.
- BSI, (1980). Methods of test for cereals and pulses. Part 3. Determination of moisture content of cereal and cereal products (routine methods). British Standard Institution. ISBN 0580114333.
- Greco, MV, Franci, ML, Golba, SLR. Pardo, AG. Pose, GM. (2014). The Scientific World Journal, vol.4, Article ID 968215, http://dx.doi.org/10.1155/2014/968215.
- Krnjaja, V, Stojanović, LJ. Cmiljanić, R. Trenkovski, S. Tomašević, D. (2008). Fungal contamination and natural occurrence of ochratoxin A (OTA) in poultry feed. *Biotechnol. in Animal Husbandry* 24: 87-93.
- Kana, JR. Gnonlonfin, BGJ. Harvey, J. Wainaina J. Wanjuki, I., Skilton, RA. Teguia, A. (2013). Assessment of aflatoxin contamination of maize, peanut meal, and poltry feed mixtures from different agroecological zones in Cameroon. *J Animal and Poultry Sci.* 2: 98-107.
- Kehinde, MT. Oluwafemi, F. Itoandon, EE. Orji, FA. Ajayi, OI. (2014). Fungal profile and aflatoxin contamination in poultry feeds sold in Abeokuta, Ogun State, Nigeria. *Nigerian Food Journal* 32: 73-79.
- Nemati, Z. Janmohammadi, H. Taghijadeh, A. Nejad, HM. Mogaddam, GH. Arzanlou, M. (2014). European J. Zoological Res. 3: 56-60.
- Okun, DO. Khamis, FM. Muluvi, GM. Ngeranwa, JJ. Ombura, FO. Yongo, MO. Kenya, EU. (2015). Distribution of indigenous strains of atoxigenic and toxigenic *Aspergillus flavus* and *Aspergillus parasiticus* in maize and peanuts agro-ecological zones of Kenya. *Agriculture and Food Security* 14 2-10.
- Pitt, JI. Hocking, AD. (2009). Fungi and Food Spoilage. Springer, New York, 3rd edition.
- Saleemi, MK. Khan, MZ. Khan, A. Javed, I. (2010). Mycoflora of poultry feeds and mycotoxins producing potential of Aspergillus species. Pak J Bot. 42: 427-434.
- Sivakumar, VK. Singaravelu, G. Sivamani, V. (2014).
  Isolation, characterization and growth optimization of toxigenic moulds from different animal feeds in Tamilnadu. *Int. J Curr. Microbiol. Appl. Sci.* 3: 430-445.