

# STATISTICAL MECHANICS OF PROTEINS IN THE RANDOM COIL STATE

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**Abstract:** Denatured proteins are mostly partially folded and compact proteins. A statistical analysis on thermodynamic properties is presented to describe and characterize denatured proteins. Conformational free energy, energy, entropy and heat capacity expressions are derived using the Rotational Isomeric States model of polymer theory. The state space and the probabilities of each state are comprised from a coil database. Properties for the denatured state are obtained for a sample set of proteins taken from the Protein Data Bank. Thermodynamic expressions of denatured state are derived.

## 1 INTRODUCTION

Random configurations of protein chains are obtained under the constraints imposed by chain connectivity and the torsion states of the backbone torsion angles  $\phi$  and  $\psi$  in the absence of sequence-distant long-range interactions. The term ‘randomly coiled proteins’ describing this state have been studied in detail by Flory and collaborators, based on the Rotational Isomeric States (RIS) Model of polymer theory (Flory, 1969); (Brant and Flory, 1965); (Brant et al., 1969); (Conrad and Flory, 1976); (Flory and Jernigan, 1965); (Rehahn et al., 1997); (Engin et al., 2009). The RIS model for a protein chain consists of two major components: (1) The statistical weights of the torsion states of the  $\phi$  and  $\psi$  angles, and (2) The proper matrix multiplication operations leading to the partition function of the chain. Thermodynamics of the single chain then follows upon proper matrix operations based on the partition function and its derivatives (Callen, 1985); (Flory, 1974). Understanding the random configurations of proteins is important due to several reasons: Firstly, the set of random configurations covers all possible initial conformations of proteins. Depending on the primary sequence, some conformations emerge as highly probable due to the amino acid specific regions of the ( $\phi$ ,  $\psi$ ) angles. Secondly, under strongly denaturing conditions, a wide range of values become available to  $\phi$  and  $\psi$ , and

conformations are close to those of the random coil (Dill and Shortle, 1991); (Tanford, 1968). These conformations are many in number, and therefore a statistical characterization is required to understand the thermodynamics of the denatured state. Thirdly, the functionally important ‘intrinsically disordered protein’ concept where the primary sequence prohibits the folded state, may suitably be analyzed by the tools used to understand the random conformations (Tompa, 2011); (Orosz and Ovádi, 2011).

Thus, a better statistical understanding of denatured proteins is required for answering questions referring to functional properties of proteins. The number of states available to the denatured chain may vary from an enormous set to only a few in numbers as observed in switches. The general statistical mechanical model that we adopt is not restricted with this variation. The size of available states is determined by the probabilities of the latter, and several sources for such probabilities are either available and may be extracted from various databases, or may be generated by suitable training techniques of bioinformatics, depending on the constraints and requirements of the problem at hand. In the present study, we extract the probabilities from the Ramachandran plots obtained from the coil library (Fitzkee et al., 2005) which is accepted to be representative of the random coiled state of proteins (Ormeçi et al., 2007); (Engin et al., 2009); (Unal et al., 2010). Having characterized the probabilities from the knowledge data base, we

apply the matrix multiplication technique to obtain the partition function, and the thermodynamic functions such as energy, entropy and heat capacity for the denatured state. Finally we present random coil results for thermodynamic functions for several proteins whose primary sequences are chosen from the Protein Data Bank.

## 2 STATISTICAL EVALUATION

A denatured protein assumes a multitude of conformations, each subject to a certain probability determined by the configurational features of the residues which are either of local or nonlocal nature. Local effects result from interactions among neighboring amino acids along the chain. We refer to this state the random coiled state of the protein. Determination of the conformation of a chain using near neighbor interactions only reduces the problem to a Markov process. Nonlocal effects are those among residues separated by more than two residues along the chain. Having adopted the probabilities from the coil library, where the sequence-distant long-range interaction are absent because secondary or tertiary structures are lacking, is a good approximation to the Markov nature of the coiled state.

Markov statistics of denatured proteins have an important place in protein statistics in general, because: (i) This is the first approximation to the difficult problem of non-Markov behavior, (ii) Markov behavior is responsible for a large body of observed phenomena, (iii) There is already a powerful and successful Markov model of characterizing the conformations of polymers, i.e., the Rotational Isomeric States (RIS) model that has been studied in some detail. The specific aim of the present paper is to extend the RIS model to calculate the thermodynamic properties of denatured chains using data generated from the denatured components of chains from the PDB.

Rotational Isomeric State (RIS) formalism (Flory, 1969) replaces the continuous distribution of backbone torsion angles by a distribution over several discrete states, and integrals over the energy surface are approximated by summations over these states. The native state of a protein is obtained when each torsion angle selects a single unique value. Two torsion angles around the alpha carbon,  $C^\alpha$ , describe the local conformation of a residue. The Flory isolated pair hypothesis suggests that each pair of torsion angles is independent of the angles

occupied by neighboring pairs (Brant and Flory, 1965); (Flory, 1969). Rose and coworkers. Zaman et al., (2003), Jha et al., (2005) and Keskin et al., (2004), Esposito et al., (2005) and Colubri et al., (2006) showed the existence of significant correlations between neighboring torsion angle pairs. In a recent work it has been shown that the usage of  $(\psi_i, \phi_{i+1})$  provides more information on backbone behavior as opposed to independent usage of residues (Lennox et al., 2009).

Some values of torsion angles are more favorable than others, and different amino acid types have different propensities to occur in different angles (Karplus, 1996). The dependence between the torsion states of two neighboring residues is a function of the type of the residues (Keskin et al., 2004). We elaborate further on this point in discussing the construction of energy maps below.

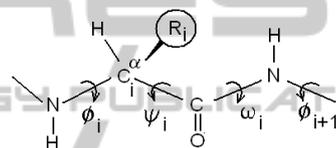


Figure 1: Torsion angles of the  $i^{\text{th}}$  amino acid.

The frequency of occurrence of a given amino acid at a given torsion state leads to the probabilities. For calculations of the random denatured conformations of proteins, a coil library serves as the source of information where torsion angle data is taken from the set of amino acids those are not in helical or beta structures. In this paper, we use the Rose Protein Coil Library (Fitzkee et al., 2005).

### 2.1 States

The backbone torsion angles for the  $i^{\text{th}}$  amino acid are shown in Figure 1. Each bond can assume different angles, with different preferences. Each residue has three torsion angles,  $\phi$ ,  $\psi$ , and  $\omega$ . The occurrence of a residue in a given  $\phi$  and  $\psi$  state, irrespective of its type is presented in Figure 1. An examination of this figure shows that the choice of isomeric states for the  $\phi$  and  $\psi$  angles is more complicated than the choice in synthetic polymer applications. In the latter, usually there are a few states like trans, gauche+ and gauche-, and their combinations for two successive bonds along the chain. In the protein case, there are several discrete states centered on different regions for the successive  $\phi$  and  $\psi$  angles, and for different amino acids.

We construct state probabilities over the Ramachandran map for each residue. 13 states are identified for the following  $\phi$  axis intervals: (-180,-150), (-150,-120), (-120,-105), (-105,-75), (-75,-40), (-40,-20), (-20,-10), (-10,30), (30,70), (70,105), (105,130), (130,155), (155,180). The corresponding intervals over  $\psi$  axis are: (-180,-160), (-160,-135), (-135,-105), (-105,-75), (-75,-40), (-40,-15), (-15,20), (20,60), (60,90), (90,110), (110,130), (130,160). For the  $\omega$  angle, there are two states, one is either (-180,-160) or (160,180), and the other is (-20,20). The states chosen in this manner are representative of the regions given by Karplus (1996) and also in (Unal et al., 2009). Thus, we identified 13 states for the angle  $\phi$ , 13 states for  $\psi$ , and 2 states for  $\omega$  as rotational isomeric states.

## 2.2 State Probabilities

The pair wise dependent probabilities of observed states of angles are defined as

$$\begin{aligned} P_X(\phi_i, \psi_i) &= N_X(\phi_i, \psi_i) / \sum N_X \\ P_X(\psi_i, \omega_i) &= N_X(\psi_i, \omega_i) / \sum N_X \\ P_{XY}(\omega_i, \phi_{i+1}) &= N_{XY}(\omega_i, \phi_{i+1}) / \sum N_{XY} \end{aligned} \quad (1)$$

where  $N_X(\phi_i, \psi_i)$  is the number of residue type X observed in the indicated states, and  $\sum N_X$  is the total number of conformations (Keskin et al., 2004); (Unal et al., 2009). Similarly,  $N_{XY}(\omega_i, \phi_{i+1})$  is the number of dipeptides of XY in the given conformations. Here,  $P_X(\phi_i, \psi_i)$  and  $P_X(\psi_i, \omega_i)$  are the probabilities of observing residue X to be in state  $(\phi_i, \psi_i)$ , and in state  $(\psi_i, \omega_i)$  respectively.  $P_{XY}(\omega_i, \phi_{i+1})$  is the joint probability of observing residue X in state  $(\omega_i)$  and Y in state  $(\phi_{i+1})$ . The neighbor-dependence introduced in the third of (1) is a dependence that originates from the residue type differences. Otherwise, (1) acknowledge the Flory isolated pair hypothesis. The conformational energies are defined as

$$\begin{aligned} E_X(\phi_i, \psi_i) &= -RT \ln \left( \frac{P_X(\phi_i, \psi_i)}{P_X^0(\phi_i) P_X^0(\psi_i)} \right) \\ E_X(\psi_i, \omega_i) &= -RT \ln \left( \frac{P_X(\psi_i, \omega_i)}{P_X^0(\psi_i) P_X^0(\omega_i)} \right) \\ E_{XY}(\omega_i, \phi_{i+1}) &= -RT \ln \left( \frac{P_{XY}(\omega_i, \phi_{i+1})}{P_{XY}^0(\omega_i) P_{XY}^0(\phi_{i+1})} \right) \end{aligned} \quad (2)$$

where the superscript 0 indicates the uniform distribution probabilities. Hence, they are directly proportional to the size of the angular intervals;  $P_X^0(\phi_i) = P_X^0(\psi_i) = 1/13$  and  $P_X^0(\omega_i) = 1/2$ . Statistical weights  $u_{\phi_i, \psi_i}$ ,  $u_{\psi_i, \omega_i}$ , and  $u_{\omega_i, \phi_{i+1}}$  corresponding to the energies may be defined by

$$\begin{aligned} u_{\phi_i, \psi_i; X} &= \exp(-E_X(\phi_i, \psi_i) / RT) \\ u_{\psi_i, \omega_i; X} &= \exp(-E_X(\psi_i, \omega_i) / RT) \\ u_{\omega_i, \phi_{i+1}; XY} &= \exp(-E_{XY}(\omega_i, \phi_{i+1}) / RT) \end{aligned} \quad (3)$$

where R is the gas constant, T is the temperature.

The statistical weight matrix for a configuration can be written as a product of statistical weights of each bond pair,  $(\phi, \psi)$ ,  $(\psi, \omega)$ , and  $(\omega, \phi)$ . For this purpose, the statistical weight matrix for a given residue X is defined as  $U_X^{\phi\psi} = [u_{\phi\psi}]_X$ ,  $U_X^{\psi\omega} = [u_{\psi\omega}]_X$ , and  $U_{XY}^{\omega\phi} = [u_{\omega\phi}]_{XY}$ . Depending on the number of states of each angle, dimensions of the statistical weight matrices  $U_X^{\phi\psi}$ ,  $U_X^{\psi\omega}$ , and  $U_{XY}^{\omega\phi}$ , are  $13 \times 13$ ,  $13 \times 2$ , and  $2 \times 13$ , respectively. The superscripts  $(\phi, \psi)$ ,  $(\psi, \omega)$ , and  $(\omega, \phi)$  identify the bond pairs over which statistical weights are calculated.

## 2.3 Calculation of the Thermodynamic Quantities

The partition sum of statistical weights for all configurations of the chain is given by (Flory, 1974)

$$Z = J^* U_1^{\phi\psi} U_1^{\psi\omega} U_1^{\omega\phi} U_2^{\phi\psi} U_2^{\psi\omega} \dots U_n^{\phi\psi} U_n^{\psi\omega} J \quad (4)$$

where  $J^* = [1 \ 0 \ \dots \ 0]$ ,  $J = \text{column}[1 \ 1 \dots 1]$ .

The thermodynamic properties, and the coefficients derived from them depend not only on a single conformation of the peptide, but on all possible configurations. In the remaining equations, we give the relevant expressions for calculating these averages.

### 2.3.1 Helmholtz Free Energy

Since the Helmholtz free energy in canonical formalism is additive over the energies, it can be calculated using the partition function of the chain (Callen, 1985).

$$-\beta F = \ln Z \quad (5)$$

where,  $\beta = 1/kT$ .

### 2.3.2 Mean Energy

The average energy is given by

$$E = -\frac{d}{d\beta}(\ln Z) = -\frac{1}{Z} \frac{dZ}{d\beta} \quad (6)$$

The matrix multiplication formalism of the partition function leads to matrix multiplication scheme of its derivatives in the following way

$$\frac{dZ}{d\beta} = L^* \left( \prod \hat{U}_i \right) L \quad (7)$$

where  $L^* = [J^* \ 0 \dots 0]$ ,  $L = \text{column}[0 \ \dots \ 0 \ J]$  and  $\hat{U}$  is the super matrix whose elements are matrices

$$\hat{U} = \begin{bmatrix} U & U'_\beta \\ 0 & U \end{bmatrix} \quad (8)$$

$$U'_\beta = \frac{dU}{d\beta} \quad (9)$$

Therefore, the mean energy can be obtained using the following multiplication scheme

$$E = -\frac{1}{Z} L^* \left( \prod G_i \right) L \quad (10)$$

where

$$G_i = \begin{bmatrix} U & U'_\beta \\ 0 & U \end{bmatrix}_i \quad (11)$$

### 2.3.3 Entropy

The entropy of the chain can be expressed in terms of  $Z$  and its derivatives with respect to  $\beta$ . Following the equality  $S/k = \beta^2 dF/d\beta$ , is obtained.

$$\frac{S}{k} = \beta^2 \left( \frac{1}{\beta^2} \ln Z - \frac{1}{\beta} \frac{\partial \ln Z}{\partial \beta} \right) = \ln Z + \beta E \quad (12)$$

Using the matrix multiplication formalism of  $Z$  and its first derivative with respect to  $\beta$ , the entropy can be calculated as

$$\frac{S}{k} = \ln \left( J^* \left( \prod U_i \right) J \right) - \beta \frac{L^* \left( \prod G_i \right) L}{J^* \left( \left[ \prod U_i \right] J \right)} \quad (13)$$

### 2.3.4 Heat Capacity

The heat capacity is one of the most important properties of the proteins, both native and denatured.

When force acting on the chain is taken as zero, denoted below by the subscript  $f=0$ , the heat capacity can be calculated as

$$C_{f=0} = \left( \frac{\partial E}{\partial T} \right)_{f=0} = k\beta^2 \frac{\partial^2 \ln Z}{\partial \beta^2} \quad (14)$$

Similar to (7), second derivative can be obtained as

$$\frac{\partial^2 Z}{\partial \beta^2} = M^* \left( \prod \hat{U}_i \right) M \quad (15)$$

where  $M^* = [J^* \ 0 \dots 0 \ 0 \dots 0 \ 0 \dots 0]$  and

$M = \text{column}[0 \dots 0 \ 0 \dots 0 \ 0 \dots 0 \ J]$ , and

$$\hat{U} = \begin{bmatrix} U & U'_\beta & U'_\beta & U''_\beta \\ 0 & U & 0 & U'_\beta \\ 0 & 0 & U & U'_\beta \\ 0 & 0 & 0 & U \end{bmatrix} \quad (16)$$

$$U'_\beta = \frac{dU}{d\beta}, U''_\beta = \frac{\partial^2 U}{\partial \beta^2} \quad (17)$$

The second derivative of  $\ln Z$  on the right hand side of the equation is written in terms of the first and second derivatives of the partition function:

$$\frac{\partial^2 \ln Z}{\partial \beta^2} = -\frac{1}{Z^2} \left( \frac{\partial Z}{\partial \beta} \right)^2 + \frac{1}{Z} \left( \frac{\partial^2 Z}{\partial \beta^2} \right) \quad (18)$$

Hence the heat capacity to be calculated by the matrix notation

$$C_{f=0} = k\beta^2 \left\{ -\left[ \frac{L^* \left( \prod \hat{U}_i \right) L}{J^* \left( \prod U_i \right) J} \right]^2 + \left[ \frac{M^* \left( \prod \hat{U}_i \right) M}{J^* \left( \prod U_i \right) J} \right] \right\} \quad (19)$$

## 3 RESULTS

In this section, the free energy, energy, entropy, and heat capacity of peptides of different sizes ranging from 10 to 800 amino acids are calculated using the RIS model, over a temperature range of 200-700 K. Table 1 lists the protein set taken from the PDB.

The variation of the free energy, energy, entropy and heat capacity is evaluated by repeating the calculations. Results are presented in Figure 2.

The curves shown in the four panels of Figure 2 are not independent from each other, and are related by the thermodynamic relations given by (5), (6), (12), and (14). It is seen that the curves in the figures all scale with the number of residues  $N$ .

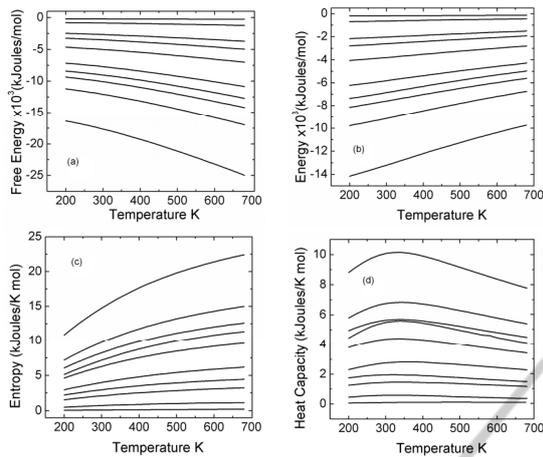


Figure 2: (a) The free energy as a function of temperature,  $T$ , for different length proteins. (b) energy as a function of  $T$ . (c) entropy as a function of  $T$ . (d) heat capacity as a function of  $T$ . The curves in parts (a),(b), and (c) are ordered from top to bottom represent proteins with the following numbers of residues: 10, 40, 120, 160, 226, 349, 408, 456, 545, and 802, respectively. In part (d) they are in reverse order.

In order to find analytical functions that will give the curves shown in Figure 2, we first chose an analytical form for the heat capacity as

$$C_{f=0}(T, N) = NT^3 \left( Ae^{BT} + Ce^{DT} \right) \quad (20)$$

keeping in mind the thermodynamic postulates. We inspired the Debye model of heat capacity in a solid that shows the dependence of  $T^3$ . Then, by integration subject to the conditions imposed by (5), (6), (12), and (14), we obtain the remaining thermodynamic functions as given in Eqs. (21)-(23). We obtain the coefficients of (20)-(23) by curve fitting as  $A = 1.5 \times 10^{-6}$  kJoules/ $K^4$  mol,  $B = -7.2 \times 10^{-3}$  1/ $K$ ,  $C = 2.6 \times 10^{-5}$  kJoules/ $K^4$  mol,  $D = -2.3 \times 10^{-2}$  1/ $K$ , and  $E = -4083$  kJoules/mol.

## 4 CONCLUSIONS

The use of the RIS model depends critically on two items: (i) the choice of the states, and (ii) the choice of the database with which the probabilities of these states are evaluated. The states are described in

$$S(T, N) = \frac{AD^3 Ne^{BT} (B^2 T^2 - 2BT + 2) + CB^3 Ne^{DT} (D^2 T^2 - 2DT + 2)}{B^3 D^3} - 2N \left( \frac{A}{B^3} + \frac{C}{D^3} \right) \quad (21)$$

$$F(T, N) = - \frac{AD^3 Ne^{BT} (B^2 T^2 - 4T + \frac{6}{B}) + CB^3 Ne^{DT} (D^2 T^2 - 4T + \frac{6}{D})}{B^3 D^3} + 2NT \left( \frac{A}{B^3} + \frac{C}{D^3} \right) + EN \quad (22)$$

terms of the populated regions on the Ramachandran map, and the possible states for the  $\phi$  and  $\psi$  angles of different amino acids are determined following the work of Karplus (Karplus, 1996). In order to apply the RIS model, however, the states available to the torsion angles  $\phi$ ,  $\psi$ , and  $\omega$  are required separately. The state space is obtained in our formulation as 13 states for  $\phi$  and 13 states for  $\psi$ , and two states for  $\omega$ . Evaluation of the probabilities follows the choice of the state space. For proof of principle, we used a coil library for the determination of the probabilities. One could alternatively construct a databank of known denatured proteins, or a subset of them depending on the nature of the investigation. Once the states are determined, the RIS model is independent of the databases used. We observed that the per residue thermodynamic properties of proteins in the random coil state scales only with the temperature. While entropy and energy increases with the temperature, free energy decreases. Heat capacity represents a decrease around 340 Kelvin that implies an energy barrier for a possible transition state. The explicit expressions that we determined for the thermodynamic functions form a thermodynamically consistent set which may be used to obtain other thermodynamic potentials by applying the known Legendre transformation techniques (Callen, 1985).

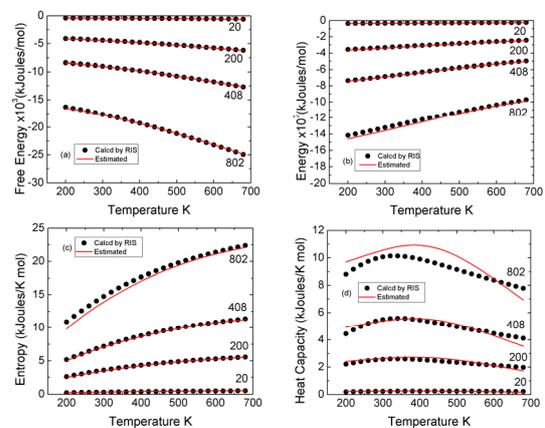


Figure 3: Comparison of (a) free energy, (b) mean energy, (c) entropy, and (d) heat capacity estimates. Exact values are calculated by matrix multiplication scheme, estimated values are calculated by fundamental relation. The lengths of chains are shown on each curve.

$$U = F + TS = \frac{AD^3 N e^{BT} (B^2 T^3 - 3BT^2 + 6T - \frac{6}{B}) + CB^3 N e^{DT} (D^2 T^3 - 3DT^2 + 6T - \frac{6}{D})}{B^3 D^3} + EN \quad (23)$$

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