## Generation of H&E-Stained Histopathological Images Conditioned on Ki67 Index Using StyleGAN Model

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Abstract: The analysis of tissue staining is a crucial aspect of cancer diagnosis. Hematoxylin and Eosin (H&E) staining captures fundamental morphological structures, while analysing Ki67-stained images provides deeper information about the tissue. However, this method is more expensive and time-consuming. Integrating machine learning techniques into pathologists' workflow can save time and resources and provide reproducible results without intra- and inter-observer variability. However, the model must be explainable to be applicable in clinical practice. A generative model can add supplementary information that serves as an explanation for model predictions. This paper demonstrates the preliminary results of the conditional StyleGAN model trained on H&E-stained images conditioned on the corresponding Ki67 indexes. In our future research, StyleGAN will be part of a model for the estimation of Ki67 index from H&E staining and will generate explanations for the model's predictions.

## **1 INTRODUCTION**

Pathologists diagnose cancer by scrutinising tissue sections under a microscope or images acquired by digital scanners. Tissue sections are stained to visually distinguish various tissue components. The analysis of stained tissue is a crucial factor in diagnosing different cancer types, influencing tumour classification and treatment recommendation.

Hematoxylin and Eosin (H&E) staining is the gold standard commonly utilised in cancer diagnosis (Gurcan et al., 2009). The H&E-stained sections depict essential morphological structures (Wittekind, 2003). To obtain more information from a tissue, pathologists often use Immunohistochemistry (IHC) staining. IHC analysis evaluates the expression of a specific protein in a tissue. Protein Ki67 indicates cell proliferation, and the Ki67 index reckons its expression as the percentage of Ki67-positive cells. While Ki67 staining provides more profound information about a tissue than H&E staining, it comes with higher time and cost requirements.

Deep learning models hold promise for improving medical diagnoses by delivering prompt, costeffective, and consistent decisions. However, a model must be explainable before it can be used in clinical practice. Particularly in medical contexts, mistakes can endanger human lives. Furthermore, the General Data Protection Regulation (GDPR) (Regulation, 2020) in the European Union requires algorithm transparency as a prerequisite for its application in patient care.

Combining a deep learning model with a generative model can provide additional information serving as explanations for model predictions, e.g., it can generate counterfactual examples. Counterfactual examples search for a minimal modification in the original data, leading the model to reverse its prediction, e.g., shifting the label from healthy to unhealthy in medical image analysis.

Generative Adversarial Networks (GAN) introduced by (Goodfellow et al., 2014) can generate new, realistic samples from the high-dimensional training data distribution. GAN is composed of a generator that creates new images and a discriminator that decides whether the generated image is similar to the ones in the training set. Conditional GAN (Mirza and Osindero, 2014) utilises an additional label of the training samples, making both the generator and discriminator conditioned on this information. StyleGAN is an improved GAN model proposed by NVIDIA researchers (Karras et al., 2019). It has en-

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hanced generator and discriminator architecture based on the principle of progressive growing, which leads to high-resolution image generation. It automatically separates high-level image features, thus building the latent space with higher interpretability.

The goal of our research is to create an explainable model for the prediction of Ki67 index from an H&E-stained image. In our research, we apply generative models to the histopathology domain in order to add explainability to our predictions and study the hidden relationship between H&E and Ki67 staining. Currently, we explore conditional StyleGAN model to generate synthetic H&E-stained images conditioned on Ki67 indexes. We utilise a histopathological image dataset consisting of pairs of adjacent H&E and Ki67-stained tissue sections. Additionally, we have the corresponding Ki67 indexes for the H&E-stained images. The paper demonstrates our preliminary results.

The paper is organised as follows. Section 2 discusses related works which apply StyleGAN model to histopathology image generation. In Section 3, we introduce our dataset and explain the model. Section 4 reports our preliminary results, and section 5 concludes with future work.

## 2 RELATED WORK

Some works have applied StyleGAN model to histopathology. The study of (Quiros et al., 2019) introduces PathologyGAN, designed for generating  $224 \times 224$  H&E-stained histopathological image patches from interpretable latent space. Pathology-GAN utilises the BigGAN architecture (Brock et al., 2018), combined with few StyleGAN features. Researchers' experiments with linear interpolation between two latent vectors illustrate a realistic transition from benign to malignant tissue with a growing number of cancer cells.

The paper (Schutte et al., 2021) utilises StyleGAN model with Convolutional Neural Network as encoder and logistic regression for generating a series of synthetic images depicting the evolution of pathology. The proposed approach generates images from the shortest path in latent space between two vectors with opposite model predictions. One of the applications analysed in the paper is changing the probability of tumour in H&E-stained image patches. However, the model cannot appropriately reconstruct histopathological images.

The work of (Daroach et al., 2021) employs Style-GAN for the generation of H&E-stained prostatic histology images. Specifically, the authors use StyleGAN2 (Karras et al., 2020), which is the improved version of the original StyleGAN (Karras et al., 2019). Their model generates new realistic 1024  $\times$ 1024 patches proved by expert pathologists. Furthermore, researchers do various experiments, e.g., with latent space representations at different StyleGAN generator levels, in order to explore their influence on different histologic morphologies. In the next experiment, scientists group generated samples into classes and recognise that the class's latent space mean represents particular morphological information. In another experiment, researchers explore the interpolation between two latent vectors of images with different histologic labels. Authors conclude that generated images look realistic but do not imitate legitimate physical transitions. Nevertheless, the work of (Daroach et al., 2021) proves that StyleGAN model can learn and subsequently generate high-resolution histopathological images.

Our research adopts a similar approach as (Daroach et al., 2021). However, we employ conditional StyleGAN3 with additional Ki67 index information about H&E-stained image patches.

# **3 METHODS**

This study demonstrates the use of conditional Style-GAN model to generate H&E-stained tissue images for given Ki67 indexes. Specifically, we aim the model to generate H&E staining corresponding to a specific Ki67 index.

#### 3.1 Dataset

The dataset was constructed from H&E and Ki67stained whole slide images (WSI) of seminoma, testicular tumor. Images were provided by the Department of Pathology, Jessenius Medical Faculty of Comenius University and University Hospital. Altogether, 84 pairs of H&E-stained tissue scans and corresponding Ki67-stained tissue scans were available. H&E and Ki67 staining were applied to adjacent sections to ensure the tissues were as similar as possible, although they did not match at the cellular level. Nevertheless, we assume the tissues in both images from the same region have similar characteristics.

Apart from the images, the dataset did not contain any labels, so it was necessary to annotate the data first. To do this, we used an improved semi-automated approach based on (Petríková et al., 2023), which contains three main steps: tissue registration, clustering into primary colours and quantification of Ki67 index. Due to the huge size of the scans and limited



Figure 1: Example of corresponding H&E and Ki67 patches from the dataset.

computational capacity, we processed images from the first level, i.e., with the second-highest resolution. Ki67 indexes were estimated from Ki67 patches using image analysis methods. Each calculated Ki67 index was then assigned as the label to the corresponding H&E patch cut in the same position. An example of a H&E patch and the corresponding original Ki67 patch is shown in Figure 1. The third clustered Ki67 patch was generated as a result of clustering. Clustered Ki67 patches were used to estimate the Ki67 indexes as a ratio of the area with the brown and blue pixels.

The training dataset for StyleGAN consists of  $256 \times 256$  square patches of H&E images. Each H&E patch is labelled with the Ki67 index calculated from the corresponding clustered Ki67 patch. An example of H&E patches with Ki67 labels from our dataset is illustrated in Figure 3. The training dataset contains 49 tissue scans cut into 189602 patches.

#### 3.2 Model AND

StyleGAN allows high-resolution image generation, which is essential for microscopic histopathology images. We utilise StyleGAN3 (Karras et al., 2021), which is equivariant to translation and rotation. Specifically, we adopt conditional StyleGAN model to H&E image patch generation conditioned on the Ki67 index. StyleGAN generates synthetic H&Estained image patches from an input latent vector and given Ki67 index. The model is illustrated in Figure 2. For the training, we use the pairs of H&E patches with corresponding Ki67 indexes. Our goal is that pathologists cannot distinguish between real and synthetically generated H&E images.

	Ki67 index	
latent vector	StyleGAN	→ H&E patch

Figure 2: StyleGAN model.



Figure 3: Example of H&E patches with Ki67 index labels from the dataset.

#### 4 **RESULTS**

We trained StyleGAN3, specifically the translation and rotation equivariant StyleGAN3-R model, from the official GitHub NVlabs repository (NVlabs, 2023). The regularisation parameter gamma was set to 2, and adaptive discriminator augmentation (ADA) was enabled. The final model viewed 5343 000 image patches (5343 kimgs) during the training, meaning it iterated through the whole dataset about 28 times. The training ran for 3 days and 8 hours on two GPUs, namely NVIDIA GeForce RTX 3090 and NVIDIA GeForce RTX 3080 Ti.

The sample of images generated by our StyleGAN model is presented in Figure 4. Images are arranged according to the increasing value of the input Ki67 index. The Ki67 index value is written above each generated image.

We evaluated the quality of generated images and latent space using two metrics. Fréchet Inception Distance (FID) (Heusel et al., 2017) assesses the quality and diversity of generated images by comparing them to real data distribution. It was calculated for the whole training dataset and 50000 generated images. Perceptual Path Length (PPL) was introduced together with StyleGAN model (Karras et al., 2019). It quantifies the disentanglement of latent space by analysing interpolation between latent vectors. It was calculated as mean for 50000 samples. FID and PPL must be minimised for more realistic generated images and higher quality latent space.

The final model was evaluated by FID and PPL metrics. The value of the FID is 16.62, while the PPL metric is 118.74. Additionally, we compared the values of these two metrics with models from the work of (Daroach et al., 2021). Firstly, their best FID model achieved FID value of 2.86 and PPL of 139.34. Secondly, their best PPL model achieved FID of 3.69 and PPL of 33.25. Researchers obtained mostly better results; however, these values are not fully comparable because models were trained on different datasets, and correspondingly the metrics were calculated on the different data.

The progress of intermediate models is illustrated in Figure 5a by FID metric and in Figure 5b by PPL metric. Metrics were calculated after every 200

Ki67 =0.0	Ki67 =0.01	Ki67 =0.01	Ki67 =0.02	Ki67 =0.03	Ki67 =0.04	Ki67 =0.04	Ki67 =0.05	Ki67 =0.06	Ki67 =0.06
Ki67 =0.07	Ki67 =0.08	Ki67 =0.09	Ki67 =0.09	Ki67 =0.1	Ki67 =0.11	Ki67 =0.12	Ki67 =0.12	Ki67 =0.13	Ki67 =0.14
Ki67 =0.14 Ki67 =0.22	Ki67 =0.15 Ki67 =0.22	Ki67 =0.16 Ki67 =0.23	Ki67 =0.17 Ki67 =0.24	Ki67 =0.17 Ki67 =0.24	Ki67 =0.18 Ki67 =0.25	Ki67 =0.19 Ki67 =0.26	Ki67 =0.19 Ki67 =0.27	Ki67 =0.2	Ki67 =0.21 Ki67 =0.28
Ki67 =0.29	Ki67 = 0.29	Ki67 = 0.3	Ki67 = 0.31	Ki67 =0.32	Ki67 =0.32	Ki67 =0.33	Ki67 =0.34	Ki67 =0.35	Ki67 = 0.35
Ki67 =0.36	Ki67 =0.37	Ki67 =0.37	Ki67 =0.38	Ki67 =0.39	Ki67 =0.4	Ki67 =0.4	Ki67 =0.41	Ki67 =0.42	Ki67 =0.42
Ki67 =0.43	Ki67 = 0.44	Ki67 =0.45	Ki67 =0.45	Ki67 =0.46	Ki67 =0.47	Ki67 =0.47	Ki67 =0.48	Ki67 =0.49	Ki67 =0.5
Ki67 =0.5	Ki67 =0.51	Ki67 =0.52	Ki67 =0.53	Ki67 =0.53	Ki67 =0.54	Ki67 =0.55	Ki67 =0.55	Ki67 =0.56	Ki67 =0.57
Ki67 =0.58	Ki67 =0.58	Ki67 =0.59	Ki67 = 0.6	Ki67 = 0.6	Ki67 =0.61	Ki67 =0.62	Ki67 =0.63	Ki67 =0.63	Ki67 =0.64
Ki67 = 0.58	Ki67 =0.65	Ki67 =0.66	Ki67 =0.67	Ki67 =0.68	Ki67 =0.68	Ki67 =0.69	Ki67 =0.7	Ki67 =0.71	Ki67 = 0.04
Ki67 =0.72	Ki67 =0.73	Ki67 =0.73			Ki67 = 0.76		Ki67 =0.77	Ki67 =0.71	Ki67 = 0.78
Ki67 =0.72	Ki67 =0.73	Ki67 =0.73	Ki67 =0.74 Ki67 =0.81	Ki67 =0.75	Ki67 =0.78	Ki67 =0.76			Ki67 =0.78
	1			Ki67 =0.82		Ki67 = 0.83	Ki67 =0.84	Ki67 =0.85	
Ki67 =0.86 Ki67 =0.94	Ki67 =0.87	Ki67 = 0.88	Ki67 = 0.88	Ki67 =0.89	Ki67 =0.9	Ki67 =0.91	Ki67 = 0.91	Ki67 =0.92	Ki67 =0.93
KI07 =0.94	Ki67 =0.94	Ki67 =0.95	Ki67 =0.96	Ki67 =0.96	KI67 =0.97	Ki67 =0.98	Ki67 =0.99	Ki67 =0.99	Ki67 = 1.0

Figure 4: Sample of generated images.



Figure 6: Example of similar pattern occurring in generated images.

kimgs. As depicted in the figures, the FID metric decreased during the training. However, the PPL metric started with low values because the model generated low-quality blurred images which were not diverse at the beginning of the training. Therefore, PPL values increased after 2000 kimgs when the image generation quality and diversity raised. At the end of the training, the PPL value started decreasing. It will probably decrease more if the training is longer.

We can observe several structures and patterns in generated images when we study them in more detail. An example of such structures is illustrated in Figure 7, where synthetic images are placed in the first column, and selected similar real images are in the second column. Both synthetic and real images have one or two zoomed parts next to the image for mutual comparison. In the first row, we can observe that the structure of synthetic cells contains similar patterns



Figure 7: Comparison of synthetic and real images.

and is considerably more regular compared to real cells. The second row shows the case when cells are arranged in regular structures, such as regular circles or arcs, which do not appear in real H&E images. Another undesirable phenomenon appears in the layout of images resembling each other. Consequently, several generated images follow a similar pattern in their layout. An example of generated images with comparable layouts demonstrating this problem is shown in Figure 6. Furthermore, we also notice typical Style-GAN artifacts which resemble water droplets in generated images.

Additionaly, the quality of generated images was evaluated by an expert pathologist from the Department of Pathology, Jessenius Medical Faculty of Comenius University and University Hospital. For this purpose, we created a sample of 30 synthetic and 30 real images arranged in random order. The evaluation was divided into two parts, namely fast and slow estimation. In fast estimation, the pathologist quickly looked at an image and estimated its realism at first glance. In slow estimation, the pathologist was allowed to analyse images for an unlimited time. For each image, the pathologist marked one of five categories, which were certainly real, rather real, certainly synthetic, rather synthetic, or they could not decide.

Table 1 shows fast and slow estimation results. In fast estimation, the pathologist could not decide about eight images, three of which were synthetic. Then was convinced that six synthetic images were real and another six synthetic images were rather real. These 12 best-ranking generated images are shown in Fig-



Figure 8: Best-ranking synthetic images.

Table 1: Pathologist e	estimation results.
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	Fast Pathologist Estimation							
	Certainly Real	Rather Real	Undecided	Rather Synthetic	Certainly Synthetic			
Real	11	10	5 4		0			
Synthetic	6	6	3	7	8			
	Slow Pathologist Estimation							
	Certainly Real	Rather Real	Undecided	Rather Synthetic	Certainly Synthetic			
Real	16	9	5	0	0			
Synthetic	0	2	0	2	26			

Table 2: Pathologist estimation Confusion Matrix.

	Fast Pathologist Estimation				Slow Pathologist Estimation		
	Estimated Real	Estimated Synthetic			Estimated Real	Estimated Synthetic	
Actual Real	NC21 AN			Actual Real		CATONS	
Actual Synthetic	12	15		Actual Synthetic	2	28	

ure 8. In summary, 15 synthetic images deceived the pathologist that they were real or the pathologist was not sure about, and 15 images were correctly labelled as synthetic. In slow estimation, the pathologist could correctly determine almost all synthetic images, except for two, which were labelled as rather real. The pathologist was certain about 26 synthetic images. Considering real images, the pathologist was correct for almost all of the examples in both fast and slow estimations. To conclude, generated images could persuade pathologists when estimating at first glance; however, the pathologist could detect synthetic examples after a more detailed analysis.

Comprehensive results from Table 1 are processed into confusion matrices in Table 2, omitting the Undecided category and merging Certain and Rather categories. In fast estimation, the accuracy of the pathologist's estimations was 69.23%, meaning the pathologist was unsure about choices. On the contrary, in slow estimation, the pathologist was almost confident with an accuracy of 96.36%.

## 5 CONCLUSION

Preliminary results demonstrate that conditional StyleGAN model is capable of generating highquality H&E-stained histopathological images. Pathologists could distinguish real and synthetic images at first glance with an accuracy of 69.23%. However, after a more detailed analysis of the images, their accuracy increased to 96.36%. Therefore, the generated image quality still needs to be enhanced.

To improve the quality of image generation, Style-GAN hyperparameters can be tuned, e.g., gamma regularisation weight or augmentation settings. In the next iteration, we will use a bigger dataset with more slides cut into patches and run the training for more kimgs. We also plan to experiment with different image patch resolutions and WSI zoom levels.

The following research will analyse the Ki67 index captured in StyleGAN latent space and the relationship between H&E and Ki67 staining. We expect that the Ki67 information improves the structure of latent space.

In our future research, we will train a model for reverse mapping to StyleGAN latent space. The model will estimate a latent vector and Ki67 index of real H&E image. Mapping real H&E images to latent space will enable different experiments with Style-GAN model, e.g., to generate counterfactual examples as an explanation of Ki67 index prediction. We can modify an input image's Ki67 index or latent vector and investigate StyleGAN's outputs. Another experiment can analyse linear interpolation in latent space, similarly to (Daroach et al., 2021). These experiments can potentially unveil concealed associations between H&E and Ki67 tissue staining, expanding scientific knowledge.

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