PodNet: Ensemble-based Classification of Podocytopathy on Kidney Glomerular Images

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Abstract: Podocyte lesions in renal glomeruli are identified by pathologists using visual analyses of kidney tissue sections (histological images). By applying automatic visual diagnosis systems, one may reduce the subjectivity of analyses, accelerate the diagnosis process, and improve medical decision accuracy. Towards this direction, we present here a new data set of renal glomeruli histological images for podocytopathy classification and a deep neural network model. The data set consists of 835 digital images (374 with podocytopathy and 430 without podocytopathy), annotated by a group of pathologists. Our proposed method (called here PodNet) is a classification method based on deep neural networks (pre-trained VGG19) used as features extractor from images in different color spaces. We compared PodNet with other six state-of-the-art models in two data set versions (RGB and gray level) and two different training contexts: pre-trained models (transfer learning from Imagenet) and from-scratch, both with hyperparameters tuning. The proposed method achieved classification results to 90.9% of f1-score, 88.9% precision, and 93.2% of recall in the final validation sets.

1 INTRODUCTION

Computational pathology is a research area that associates biological tissue analysis with digital image processing and computer vision techniques (Srinidhi et al., 2021). As a result of advances in image analysis algorithms, the main approaches adopted are currently based on deep learning architectures (Deng et al., 2020).

The difficulty of finding fully annotated medical image data sets and various cases associated with the complexity of the anatomy of the images’ biological structures makes the development of histological image analysis systems a challenging task.

Currently, there are many proposals in the literature for automatic histological image analysis systems applied to different organs and diseases (Yari et al., 2020; Candelero et al., 2020; Thomas et al., 2021). However, there are little explored diseases, as podocytopathy.

Podocytes are cells of the kidneys’ visceral epithelium, and they are present in the internal structure of renal glomeruli (Chen et al., 2006). The primary function of podocytes is to restrict the passage of proteins from the blood through the urine (Nagata, 2016). Podocyte lesions can compromise the ability of a glomerulus to filter proteins, causing damage to the glomerular structure, and are biomarkers of progressive glomerulosclerosis (Saga et al., 2021) (See in Figure. 1 image with (a) and without (b) podocytopathy).

The diagnostic of the lesions in renal glomeruli may substantially vary according to the experience of the pathologist and a system that could automatically classify such lesions could be of great help for pathologists. On one hand, these systems could reduce the subjectivity of analyses, accelerate the diagnosis process, and improve medical decision accuracy; on the other hand, they could be used as a teaching tool for training new pathologist (Jayapandian et al., 2021).
The evolution of artificial intelligence systems allow them to achieve better results in many others areas, as described in (Salas et al., 2019; Silva et al., 2020; Abade et al., 2021; Abade et al., 2019).

As described in (Salas et al., 2019; Silva et al., 2020; Abade et al., 2021; Abade et al., 2019), visual recognition that a glomerulus has podocytopathy, a pathologist needs to identify the podocytes and then the lesions. However, podocytes are easily confused with other intraglomerular cells (endothelial and mesangial for example). Therefore, a system that automatically classifies aglomerulus image with podocytopathy can be a very useful aid tool in the practice of nephropathologists. (Maraszeck et al., 2020; Zimmermann et al., 2021; Zeng et al., 2020; Govind et al., 2021b). We found few proposals for automatic podocyte analysis systems and none of the studies found classify glomerulus images concerning the presence of podocytopathy, but segment podocytes to associate them with other diseases.

In this work, we propose a method, an artificial intelligence system based on a convolutional neural network (CNN) for the classification of histological images of renal glomeruli with podocytopathy. We also present a novel labeled public data set of renal glomeruli images with podocytopathy.

2 RELATED WORKS

The literature review that we carried out for this work focused on seeking two groups of researches: (i) classification of lesions in renal glomeruli and (ii) classification of glomerular podocytopathies.

Among the works on the classification of lesions in glomeruli, the classification task is performed in two ways: (i) in a single step, using images of isolated glomeruli, or (ii) in two steps, performing the segmentation of glomeruli in the Whole Slide Images (WSI) before executing the classification. We present both works below, focusing on the results of the classification task.

A recent work (Yang et al., 2021), classified the glomeruli into five classes of lesion (including sclerosis) using the Densenet network (Huang et al., 2018) and an LSTM (Long Short Term Memory) (Ullah et al., 2018). The data set consisted of 1379 WSI (41886 glomeruli), stained in HE (15298), PAM (5649), PAM(5641), and trichrome (5679). The best result achieved for the task was 94.0% accuracy for the different lesions in HE stained images. A previous work (Kannan et al., 2019), used 1706 images of glomeruli, stained in trichrome to classify them into four classes: (i) non-glomerulus, (ii) normal (iii) partially sclerosed and (iv) globally sclerosed. The network used was Inception v3 (Szegedy et al., 2015) and the result obtained was a classifier capable of discriminating non-glomerular images and images with lesions with an accuracy of 92.67% ± 2.02%. In the work of (Gallego et al., 2021), after segmenting glomeruli with the U-Net network, they perform the classification of glomeruli as normal or sclerotic. The data set used had 51 tissue slides, stained in PAS (37) and HE (14). The obtained results were of F1 of 94.0% (normal) and 76.0% (sclerosed).

Following the presented work in (Bukowy et al., 2018), has proposed a method for identifying healthy or injured glomeruli (without specifying the type of lesion). The data set used had 87 WSI slides, all stained with trichrome. The network used was Alexnet and the average precision and recall were 96.94% and 96.79%, respectively. (Jiang et al., 2021) classify glomeruli into 3 classes: normal, with sclerosis, or other lesions. The network was trained with 1123 snapshots, which are smaller portions of the blade with one or more glomeruli. The results obtained in the work were f1 scores of 91.4%, 089.6%, 68.1%, and 75.6%, for normal glomeruli, with sclerosis, global sclerosis, and other lesions, respectively.

In the work of (Uchino et al., 2020) 15888 images of renal glomeruli were used, distributed among 7 classes of histological lesion types: global sclerosis, segmental sclerosis, endocapillary proliferation, mesangial matrix accumulation, mesangial cell proliferation, crescent and structural membrane changes basal. The network used was Inception v3 and the best result obtained was an area under the curve (auc) of 0.986 for classification of stained images in PAS and 0.983 in PAMS. (Mathur et al., 2019) performs two tasks: (i) classifies glomeruli as normal or abnormal and (ii) classifies regions of tissue without glomerulus into three classes of fibrosis: mild, moderate or severe. The data sets were composed of patches of images extracted from tissue slides, totaling 935 images of glomeruli in data set 1 and 923 images of regions without glomerulus in data set 2. The method used was a new proposed model, the Multi-Gaze Attention Network (MGANet). The result ob-
obtained was 87.25% and 81.47% accuracy for classification of glomeruli and fibrosis, respectively. As described in (Barros et al., 2017) a classifier of renal glomeruli images regarding hypercellularity was proposed. The result obtained was 88.3% accuracy. The data set had 811 images stained in PAS and HE. Based on this work (Chagas et al., 2020) performed the same task and data set, however, using convolutional neural networks as features extractor and classification with the SVM algorithm. In addition to the binary classification task, the authors performed the classification of sub-lesions of hypercellularity: mesangial, endocapillary, and both, reaching an average accuracy of 82.0%. Both in the binary task and the multiclass classification, the proposed method surpassed the Xception, ResNet50, and Inception v3 networks, as well as the (Barros et al., 2017) method.

Among the works focusing on automatic podocyte analysis, we found works on podocyte detection and segmentation, however, none focused on podocytopathy, but to associate podocytes with other diseases (Zeng et al., 2020; Govind et al., 2021b; Govind et al., 2021a; Zimmermann et al., 2021; Maraszek et al., 2020).

The work of the (Zeng et al., 2020) aimed to locate glomeruli, classify glomerular lesions, and identify and quantify different intrinsic glomerular cells. For the task of classification of glomeruli, 1438 images of glomeruli were used. The method used in the classification was the DenseNet-121 and LSTM-SENet networks. In the internal cell segmentation task (including podocytes), 460 images of glomeruli were used, containing approximately 70 thousand annotated cells. The method used in the segmentation of the internal cells was the 2D V-Net network. The results obtained in the study were 95.0% for classification of the glomerular lesion, 88.2% for average precision, and 87.9% for average recall for detection of glomerular internal cells.

In (Govind et al., 2021a), the authors use PAS stained WSI slides to detect and quantify podocytes and correlate podocyte loss in diseased glomeruli. The data set consisted of 122 slides stained in PAS, with images originating from rat, mouse, and human tissue. The results obtained were a sensitivity and specificity of 0.80/0.80, 0.81/0.86, and 0.80/0.91 in mice, rats, and humans, respectively. (Govind et al., 2021b), perform the segmentation and quantification of podocytes for recognition of Wilms’ tumor. The data set used was composed of PAS stained images, originating from mice. The proposed method uses a GAN (Generative Adversarial Network) to convert images of glomeruli to its immunofluorescence version, where the segmentation of podocytes occurs by traditional image processing methods. The result obtained was 0.87 sensitivity and 0.93 specificity for detecting podocytes.

In (Maraszek et al., 2020) the objective was also to detect and quantify renal podocytes, however, to associate them with the presence of diabetes mellitus. Like the work of (Govind et al., 2021b), the proposed method used immunofluorescence images with PAS versions of the same glomeruli. Immunofluorescence imaging was processed with classical methods of digital image processing. The data set consisted of 883 images of rat glomeruli. At the end of the work, the authors calculated the damage to the glomeruli through morphological analysis of the podocytes and their intraglomerular distribution. The results obtained had a sensitivity of 72.7%, specificity of 99.9%, and accuracy of 95.9% in the location of podocytes.

Finally, (Zimmermann et al., 2021) used a data set with 1095 immunofluorescence images, containing a total of 27696 labeled podocytes. The aim of the study was also to detect podocytes but to associate them with the disease antineutrophil cytoplasmic antibody-associated glomerulonephritis (ANCA-GN). The network used to segment glomeruli and podocytes was the U-net. The Dice coefficient obtained in the segmentation tasks of both glomeruli and podocytes (0.92) was greater than 0.90.

3 MATERIAL AND METHODS
3.1 Data Set Preparation
The data set used in this work has 835 images of renal glomeruli (340 with podocytopathy and 430 without podocytopathy). All images were labeled by two pathologists and were obtained from different institutions and laboratories. The data set was available by Instituto Gonçalo Moniz - FIOCRUZ and comprises images stained in trichrome (173), Periodic Acid-Schiff (PAS)(409), PAM (169), and Hematoxylin and Eosin (H&E) (74). The images were captured using whether cameras were attached to microscopy or by digital scanners and came in different formats (JPG, PNG, and TIF) and resolutions (from 238 × 201 to 1920 × 1440 pixels). The images labeled “with podocytopathy” have different podocyte lesion types: hypertrophy, hyperplasia, and degeneration. Some samples are depicted in Figure. 2. Additionally, both in the group of images with podocytopathy and in the group without podocytopathy there are other types of associated lesions (hypercellularity, sclerosis, membranous).
3.2 Proposed Method (PodNet)

We built our solution over the hypothesis that when converting the images, originally in the RGB color space, to another color spaces that could isolate information from the stain used in the acquisition of the images, associated with the extraction of features through pre-trained convolutional neuron networks (a practice adopted by (Chagas et al., 2020) and (Mathur et al., 2019)), could extract complementary features, which does not occur using an end-to-end network with images in a single color space.

Traditional convolutional neural network architectures receive images in only a certain color space. However, given the possibility of isolating the information of the stains with which the images were obtained in channels of a color space, our hypothesis that, associating features extracted from the images in different color spaces could result in a classification method more robust, is plausible.

The method is organized into three steps: Preprocessing, feature extraction, and classification. The figure. 3 illustrates the architecture of the network and its three steps.

In the first step, preprocessing, the images were normalized (values between 0 and 1). Then, the conversion from the RGB version to the HED and HDX versions takes place. The HED (used in (Barros et al., 2017)) convert into color space information relating to Hematoxylin, Eosin, and DAB stains into its channels. The HDX color space converts into color spaces channels of Hematoxylin and PAS information.

The second step is the extraction of features, which is performed by processing the 3 images resulting from the conversion to the VGG19 network pre-trained with the Imagenet data set. The network has been modified so that the output is the result of max-pooling of the last convolutional layer. After that, a flatten operation is performed, which generates a vector of features for each image. Then, each of the three feature vectors is scaled using the PCA algorithm (Tipping and Bishop, 2006). The objective of this operation, in addition to reducing the network’s hyperparameters, is to speed up the training process and eliminate unimportant features. Finally, the three vectors are concatenated, resulting in a single vector.

In the classification step, there is a dense artificial neural network formed by an input layer of 350 neurons (features resulting from the PCA algorithm) and 4 hidden layers (256, 128, 64 and 64 neurons, respectively), with dropout regularization (0.1) between each hidden layer. The hyperparameters of this network were tuned with the Keras (Chollet et al., 2015) grid search tool strategy. The network output is a neuron with a sigmoid activation function.

3.3 Experiments Protocol

All the experiments reported in this work were made using Python 3.6.8 programming language, Tensorflow 2.41 deep learning library for GPU, Docker environment (Merkel, 2014), and Keras Tuner (O’Malley et al., 2019) (for hyperparameters tuning), running on NVidia GeForce 2080 TI (11 GB) GPU.

The classification results obtained with the proposed method were compared with six other models based on deep learning architectures: Resnet101 v2, VGG19, Densenet201, Inception Resnet v2, Inception v3, and Xception. These architectures were chosen, especially, because they represent different models in terms of depth and learning strategy, which allowed a broad analysis of the performance of conventional networks for the execution of the proposed task.

The training and validation of the networks was carried out in two ways: (i) Generalization test and (ii) Final validation. In the final validation the entire data set was divided into 70% for training and 30% for testing. In the generalization test, a 5-fold cross validation was performed on the training set used in the final validation (on the same set of 70% of the data). In the training sets of the cross-validation and the final validation set, the same data augmentation was performed. The operations performed were: horizontal and vertical flip, rotation (30, 90, 270 degrees),

1More info.: https://www.tensorflow.org/.
Figure 3: Proposal architecture. The proposed method (PodNet) extracts feature from the image in different color spaces with the pre-trained VGG19 network.

Before training the models, a hyperparameters tuning was performed. The adjusted hyperparameters were: Batch size (16, 32 and 62); number of neurons from top dense layers (2048, 1024, 512 and 256), learning rate (to 0.1 from 0.000001), top dense layers activation functions (softmax, relu and tanh), optimizer (RMSprop, Adam, SGD and Adamax), momentum (0.3, 0.6 and 0.9) and loss function (binary crossentropy and hinge).

Baseline networks were trained in two scenarios: (1) Trained from scratch (with initialization of random weights) and (2) trained with transfer learning (with weights initialization of Imagenet data set (Rusakovsky et al., 2015)). In the scenario trained from scratch, the baseline networks were also trained with the data set in the gray level version. This was done to observe the effects of the absence of color information over the performance of the networks.

In the transfer learning scenario, the weights of networks trained on the Imagenet data set were loaded into baseline networks (pre-trained networks). The top layers of the baseline networks (with 1000 neurons - originals classes number of the Imagenet data set) were replaced by new layers: two dense layers and one neuron (with sigmoid activation) for binary classification (with and without injury). The last step was to perform a tuning training of the added top layers by five epochs with all other layers frozen, and finally, unfreezes and trains all layers of the networks.

We use the early stopping (monitor = validation loss) as a strategy to stop training networks with patience 5 for all models (to avoid over fitting). The final state of the baseline models was obtained by loading the best weights obtained during net training.

4 RESULTS AND DISCUSSION

The metrics used to calculate the performance of the networks were: precision, recall, and f1-score (James et al., 2013). We also calculated the area under the ROC curve of all evaluated models on final validation set (see Baselines ROC curves in Figures: 5, 6 and 7. See Top 4 ROC curves models in Figure: 4). The results show in Table 1 are the average of the metrics calculated for the five folds from cross-validation (generalization set) and final validation results.

The complexity and diversity of data set images and the fact that the discriminatory features are contained in some nuclei, which are microscopic structures, contributed to the general results not reaching absolute values. Additionally, even with data augmentation operations, the data set is still small. The method proposed presented satisfactory results when compared to the other networks, with the highest F1 score in the final validation (90.0%) and the second better on the generalization set (90.1%). The network with the best values in the generalization set was Resnet 101 v2 (90.2%) trained with transfer learning with RGB data set version but presented a much lower result in the f1 score final validation (84.4%), which is a more relevant test set, as it has the largest number of samples.

We also consider the results to be good, when comparing the results of PodNet with other studies on the classification of glomeruli concerning other lesions, such as (Mathur et al., 2019) (87.25% and 81.47% of f1 score for Fibrosis), (Gallego et al., 2021)
Table 1: Summary of results obtained in all models evaluated and ranked from F1-score in the final validation set.

<table>
<thead>
<tr>
<th>Classification Models</th>
<th>Generalization Set (average)</th>
<th>Validation Set</th>
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<tbody>
<tr>
<td></td>
<td>Prec(%)</td>
<td>Rec(%)</td>
</tr>
<tr>
<td>Proposed method (PodNet)</td>
<td>90.6±2.10</td>
<td>89.6±1.36</td>
</tr>
<tr>
<td>Densenet201 TL (RGB)</td>
<td>90.0±3.66</td>
<td>90.0±5.31</td>
</tr>
<tr>
<td>Inception v3 TL (RGB)</td>
<td>87.0±1.03</td>
<td>88.0±8.93</td>
</tr>
<tr>
<td>Resnet101 v2 TL (RGB)</td>
<td>94.0±2.65</td>
<td>86.0±7.44</td>
</tr>
<tr>
<td>VGG19 TL (RGB)</td>
<td>93.0±2.56</td>
<td>86.0±6.03</td>
</tr>
<tr>
<td>Xception TL (RGB)</td>
<td>89.0±2.66</td>
<td>90.0±8.26</td>
</tr>
<tr>
<td>Inception Resnet v2 FS (RGB)</td>
<td>86.0±5.9</td>
<td>75.0±12.0</td>
</tr>
<tr>
<td>Inception Resnet v2 TL (RGB)</td>
<td>92.0±3.76</td>
<td>90.0±5.75</td>
</tr>
<tr>
<td>Densenet201 FS (RGB)</td>
<td>82.0±7.9</td>
<td>83.0±9.02</td>
</tr>
<tr>
<td>Inception v3 FS (GL)</td>
<td>80.0±6.94</td>
<td>67.0±6.02</td>
</tr>
<tr>
<td>Resnet101 v2 FS (GL)</td>
<td>80.0±6.7</td>
<td>82.0±1.97</td>
</tr>
<tr>
<td>Inception Resnet v2 FS (GL)</td>
<td>79.0±7.67</td>
<td>88.0±9.60</td>
</tr>
<tr>
<td>Densenet201 FS (GL)</td>
<td>72.0±7.8</td>
<td>84.0±1.36</td>
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<tr>
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<td>86.0±1.41</td>
<td>61.0±14.4</td>
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<tr>
<td>Xception FS (GL)</td>
<td>82.0±5.02</td>
<td>69.0±5.82</td>
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<tr>
<td>Inception v3 FS (RGB)</td>
<td>83.0±4.02</td>
<td>78.0±10.0</td>
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<td>Resnet101 v2 FS (RGB)</td>
<td>72.0±10.1</td>
<td>65.0±14.0</td>
</tr>
<tr>
<td>VGG19 FS (RGB)</td>
<td>89.0±4.80</td>
<td>80.0±9.80</td>
</tr>
<tr>
<td>Xception FS (RGB)</td>
<td>76.0±5.92</td>
<td>69.0±12.0</td>
</tr>
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</table>

Figure 4: ROC curves from top 4 models. The proposed method is the best area under curve followed by three models trained with transfer learning in the RGB dataset version.

Figure 5: ROC curves from baseline models trained with from scratch in the gray level dataset version. The best model this training context was Inception v3.

Figure 6: ROC curves from baseline models trained with transfer learning in the RGB dataset version. The best model this training context was Densenet 201.

Figure 7: ROC curves from baseline models trained with transfer learning in the RGB dataset version. The best model this training context was Densenet 201.
F1 score of 94.0% (normal) and 76.0% (sclerosed) and (Kannan et al., 2019) 92.6% acc, also for sclerosis. About baselines results, when comparing the networks trained from scratch with the data set in the RGB and gray level version, we were unable to conclude whether the absence of color information offered benefits or harm to the learning of networks, given that there were architectures with better results in the RGB version and others in the gray level version.

5 CONCLUSIONS

In this work, we propose a method, called PodNet, for the classification of histological images of renal glomeruli with podocytopathy, and we present an unpublished public data set of histological images of renal glomeruli with podocyte lesions. The proposed method has better results when compared against well-known CNN networks. The experiments indicated that deep neural networks are a promising approach for supporting the development of a system to automatically classification of podocytopathy in histological images. The data set presented will continue to increase with new images, and be made available to researchers with academic interests. Additionally, the studied lesions do not have a high incidence, we intend to use new data augmentation strategies to solve the low amount of available images. An ablation study will be carried out to systematically analyze the contributions of end-to-end training networks with different color spaces, in the data set provided here and other problems with multi-stain images. Finally, we also highlight the possibility of segmentation and classification of podocyte lesions, quantifying or correlating them to diseases using models based on deep learning.

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