

Robust Image Analysis of BeadChip Microarrays

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Abstract: Microarray images in molecular genetics are heavily contaminated by noise and outlying measurements. This paper is devoted to analysis of Illumina BeadChip microarray images, primarily to their low-level preprocessing. We point out that standard image analysis procedures, which are implemented in the beadarray package of BioConductor software, are highly sensitive to contamination by severe noise and outliers. Therefore, the habitually used methodology does not discover many of the outliers. We illustrate this on real data and show that the standard background correction method may actually amplify the noise in the image. A robust image analysis tailor-made for this type of microarray images is highly desirable. We explain principles and show preliminary results of our robust alternative to the standard approach, which aims to be robust to noise and outliers in each its step.

1 BEADCHIP MICROARRAYS

Microarrays represent a commonly used technology for measuring gene expressions. Microarray studies are typically designed to find differently expressed genes in two or more groups of samples (e.g. patients with a different form of a disease) or to perform (unsupervised) clustering or (supervised) classification analysis into given groups (Fraser et al., 2010).

Illumina BeadChip microarrays are claimed to be the currently most popular technology for measuring gene expressions (Rueda, 2014). A sample is placed in a microwell array containing about a million of silica beads corresponding to different gene transcripts (Kuhn et al., 2004). The surface of the chip is scanned to obtain a gray-scale image with a high fluorescence intensity corresponding to highly expressed genes.

We focus on the low-level preprocessing of BeadChip images, which is a part of the BeadChip image analysis with the aim to estimate expression values for each bead from the raw scanned image, i.e. after filtering out the effect of the background. Various sources call the low-level preprocessing by different names:

- Feature-intensity extraction (Smith et al., 2010)
- Low-level analysis (Dunning et al., 2008)
- Data reduction (Fraser et al., 2010)

- Quantification (Rueda, 2014)

The habitual approach to the image analysis of BeadChip microarrays is based on the algorithm of (Dunning et al., 2007), which does not contain a description of details of the methodology and practitioners have to rely on default values of its parameters. The approach is implemented in the package beadarray, which in its current version 1.16 can be considered a standard software for the preprocessing of BeadChip microarrays, before performing a more advanced analysis with the standard lumi package. Both packages are part of the open-source software Bioconductor (Rueda, 2014).

The basic intensity is observed everywhere in the image and the target intensity in a particular bead corresponds to a gene expression. A raw BeadChip image is a 16-bit gray-scale image, which contains one of 65 536 values. Performing the complex process of outlier or noise detection is computationally intensive and the difficult task of information extraction from the massive 2D images with tens of millions of pixels makes the field of microarrays image analysis to be an important hot topic in current bioinformatics (Cardona and Tomancak, 2012).

Observed gray intensities of BeadChip microarray images are heavily contaminated both in the beads as

well as in the background. We have observed a contamination by additive as well as impulsive (severely outlying) noise in isolated pixels or in smaller or larger regions in a variety of applications due to various reasons. To give only a few of them, these include measurement errors, unreliable sample preparation, or wrong identification of gene probes (Fraser et al., 2010).

Only marginal attention has been paid to the low-level preprocessing of BeadChip images. Specific problems of BeadChips do not allow to simply use sophisticated tools for image analysis and quality assessment, which are available for Affymetrix microarrays (Kuhn et al., 2004). Instead, we hold the opinion that a robust alternative tailor-made for BeadChip images would be highly desirable and we bring arguments in favor of such opinion in this paper.

This paper has the following structure. Section 2 explains that procedures habitually applied on BeadChip microarray images are highly vulnerable with respect to noise. Section 3 proposes a computational improvement for the foreground estimation. Currently, we work on robustifying standard image analysis approaches for BeadChip microarrays and our ideas of robust image analysis for this task are summarized in Section 4. Some results of the analysis of real data are shown Section 5, revealing the sensitivity of standard image analysis procedures with respect to noise and outliers in the images. Finally, Section 6 concludes the paper.

2 PROBLEMS OF STANDARD LOW-LEVEL PREPROCESSING

Standard analysis of BeadChip microarray images is sensitive to random or systematic errors in the raw data and to artifacts of different sizes and shapes. Its methods have been derived primarily with the requirement on a high speed (Kuhn et al., 2004). This section describes all steps of the standard image analysis algorithm and our critical stance will be illustrated on examples in Section 5. Thus, we bring new ideas and findings beyond those of (Smith et al., 2010), who simply recommended to ignore problematic beads in the images.

Let us now critically describe all the steps of the standard image analysis performed on each BeadChip microarray image. Commonly, the same sample is applied on two neighboring strips (Shi et al., 2009). The following are the steps implemented in the beadarray package.

- **Estimating the Local Background Effect** by the

5-th smallest value in a square neighborhood of each bead is heavily influenced by local noise.

- **Estimating the Foreground** is performed by three consecutively applied linear filters.
 1. **Image Sharpening** can yield negative intensities. Besides, it yields nonsense (also negative) values for pixels at the boundary of the image where there are no beads.
 2. **Averaging** (mean filter) over a square of size 3×3 pixels around each given bead propagates the effect of outliers to their neighbors.
 3. Another **averaging** of four neighboring pixels of a particular bead.
- **Background Correction** defined as the difference between foreground and background intensities. It may also yield negative values, which is the consequence of the sharpening.
- **Data Normalization** has been largely discussed and positively evaluated (Shi et al., 2009), but it leads to hiding some outliers.
- **Outlier Deletion** is performed after mixing the data from both strips, which prevents some outliers from being detected.

Unfortunately, these methods in the beadarray package with default values of their parameters are strongly influenced by noise and random or systematic errors in the measurements. They transfer the effect of noise from noisy pixels also to neighboring pixels (and neighboring genes), which can be amplified from one non-robust step to later ones. In this way, noise is introduced artificially to such genes which are not affected by noise in the raw image. The results of the whole process are also too much influenced by the sharpening. Besides, there is not even an attempt to correct for spatial artifacts.

Attention has been paid to the choice of a suitable background correction method (Smith et al., 2010), which has only a small influence on the result, but no doubts have been cast upon other steps of the low-level preprocessing. Nevertheless, they all can be easily shown to have a zero breakdown point, which is a statistical measure of sensitivity against outliers in the data (Davies and Gather, 2005).

The local approach to estimation of both background and foreground does not exploit information about global trends across the whole strip. Besides, the initial steps of the standard image analysis of microarrays are strongly influenced by local noise in the neighborhood of particular microbeads and the resulting biased values are passed on to the next estimation methods and transformations. Outliers become masked among the data and their consequent detection and deletion is much more complicated.

3 FOREGROUND ESTIMATION

The three linear filters of the foreground estimation within the standard image analysis of the beadarray package can be expressed by means of a single linear filter. Here, we show the filter equivalent to image sharpening together with the consecutively applied averaging. We propose its more efficient computation and discuss whether the function meets its expectations.

Let us denote the intensity in pixel with coordinates $[i, j]$ by w_{ij} . The sharpening procedure replaces w_{ij} by

$$3w_{ij} - \frac{w_{i,j+1}}{2} - \frac{w_{i,j-1}}{2} - \frac{w_{i+1,j}}{2} - \frac{w_{i-1,j}}{2}. \quad (1)$$

In the current code, sharpening is computed over the whole image and then the foreground intensities are computed as the weighted combination of four averaged values. These two filters can be namely easily described by one procedure, saving much computational complexity in terms of floating point operations.

This computational improvement based on combining the computation of sharpening and averaging will be now proposed. It can be easily derived that both filters can be joined to one replacing w_{ij} directly by linear combination of raw intensities of pixels in the neighborhood of the pixel $[i, j]$ with coefficients given by this convolution mask:

0	-1/18	-1/18	-1/18	0
-1/18	2/9	3/18	2/9	-1/18
-1/18	3/18	1/9	3/18	-1/18
-1/18	2/9	3/18	2/9	-1/18
0	-1/18	-1/18	-1/18	0

Besides, there is no justified reason for preferring this sharpening approach rather than any other one, e.g. that of (Tanaka et al., 2010), or for combining sharpening with smoothing, because sharpening increases contrast and smoothing removes it. The motivation for using sharpening, although formulated in a vague way, was formulated by (Dunning et al., 2007) as follows. A bead with a high intensity was claimed to deplete all of the target molecules from the neighborhood and therefore the measured intensity is smaller than the real intensity. A reasonable transformation should take this into account by increasing the measured intensity. Such idea is definitely not fulfilled by the sharpening of formula (1). Actually, (1) increases a local contrast, but it does not increase the intensity in a pixel with a large intensity surrounded by neighbors also with a large intensity. Besides, we can say that sharpening is accompanied by a consequent averaging, which has the function to improve the bias introduced by sharpening.

4 ROBUST IMAGE ANALYSIS

The sequence of standard low-level preprocessing in package beadarray is sensitive to heavy contamination by a serious noise, which is omnipresent in gene expression measurements. Therefore, it is highly desirable to replace standard methods by robust counterparts. Tailor-made methods for microarray image analysis should be proposed using the knowledge about the causes of noise and errors in the images. These can either detect outliers and completely ignore all of them, but other possibilities include to replace them by values estimated in a suitable model or to down-weight them.

The standard analysis of BeadChip microarrays does not exploit available image denoising methods. On the other hand, denoising itself is complicated and its numerous methods rely on certain assumptions, e.g. Gaussian distribution of noise, which may be unsuitable and in fact allows to preserve some sorts of outliers. We actually do not have a good experience if the microarray image analysis is started by a simple denoising and continues with standard methods.

It is important to robustify all steps of the approach implemented in beadarray. For example, replacing the average by a robust estimator of location with a high breakdown point turns out to be useful in estimating the foreground. However, we are not aware of any application of highly robust methods in the analysis of microarray images, which would be based on the concept of robust image analysis. This has been described as a branch of image analysis exploiting the ideas and methods of robust statistics with a high breakdown point.

We are working on implementation of a robust alternative to the standard approach in C++. Let us present some key ideas for improving the robustness:

- Robustify each individual step in the whole process of the image analysis by a method with a high breakdown point.
- Replace linear filters by robust counterparts. For example, filters based on order statistics are more suitable for impulsive noise.
- Ignore pixels with a nonsense intensity (e.g. zero intensity).
- Ignore such beads, which have outliers in their neighborhood. Further, outliers should be removed after each step of the analysis. Only this can prevent from masking of outliers.
- Treat very bright beads and their direct neighbors (Figure 1) in a specific way, e.g. delete them or model their effect and subtract it from the measured intensities.

- Perform an artifact detection and correction tailored for artifacts of various sizes and shapes.
- Use a global model for the trend across the whole strip and use an efficient algorithm for this intensive computation.

To give an example, let us consider estimating the local background effect. Here, we propose to replace the standard approach (see Section 2) by the least weighted squares estimator of location (Kalina, 2012) computed from $w_{(6)}, \dots, w_{(10)}$, where

$$w_{(1)} \leq w_{(2)} \leq \dots \leq w_{(25)} \quad (2)$$

are ordered gray intensities of a circular neighborhood of a given pixel. Better than that, considering a circular neighborhood would ensure a rotation invariance of the computation.

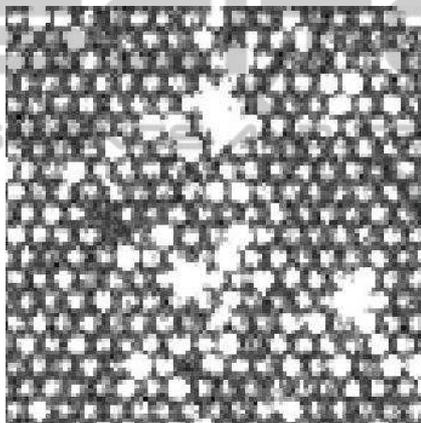


Figure 1: A cut-out of a raw BeadChip microarray image. Its standard analysis does not have a special treatment for highly expressed beads transmitting their signal to their neighborhood.

Currently, we are comparing various alternative methods, which are also accompanied by a robust statistical analysis, using e.g. following ideas.

- Use a robust regularized classification analysis method, which is suitable for high-dimensional data, or a robust dimensionality reduction, using e.g. recently proposed methods of (Filzmoser and Todorov, 2011; Kalina, 2014).
- Use a weighted classification analysis, where each gene obtains a weight according to its variability over beads.
- Classification over beads instead of over gene transcripts, at the cost of a higher computational intensity.
- Optimize parameters of image analysis procedures to improve classification results, e.g. by robust optimization (Xanthopoulos et al., 2013).

5 EXAMPLE

The harmful influence of noise on standard procedures of microarray image analysis will be illustrated on real data. We analyzed a data subset from a genetic study of the Center for Biomedical Informatics (Kalina and Zvárová, 2013). Blood samples of 24 patients with cerebrovascular stroke and of 24 control individuals were examined by HumanWG-6 Expression BeadChips according to manufacturer's protocol.

First, we investigate the effect of the sharpening and averaging within the process of foreground estimation. In practice, the user of beadarray may select to turn it off, while it is not possible to turn off the averaging. Users are advised to use sharpening (Dunning et al., 2007; Dunning et al., 2008), although its influence on the result is extremely high.

We analyzed 48 images (strips of size 2389×18309 pixels) from different microarrays by means of different approaches, including the option not to use sharpening and averaging. Figures 2 and 3 show averaged gene expressions across the 2389 rows of a microarray strip across 48 microarrays for a particular gene. The gene was selected randomly as a typical gene without a differential expression between patients and control individuals. The plots contain boxplots of four groups, corresponding to top, upper middle, bottom middle, and bottom row, respectively. In other words, the total number of 2389 rows was divided to 4 equally sized groups, while outliers were discarded from the figures.

Figure 2 shows average foreground intensities computed without performing sharpening and averaging. Figure 3 analogous values computed with sharpening and averaging. The effect of these transformations can be seen as a deformation of values in the last rows of the strip. The figures quantify our experience with heterogeneity of expressions across the strip, which we have observed in the majority of pre-processed (but not raw) microarrays.

Moreover, beadarray detects 0.9 % of beads to be outliers in the outlier deletion step of the algorithm. Compared to this strikingly small percentage, our preliminary version of robust image processing considers as much as 15 % of beads to be too influenced by spatial artifacts or noise and treats them in a specific way.

To quantify the adverse effect of outliers in the images, we evaluated a simple robust version of the image preprocessing. Its parameters were tuned to detect a given percentage of outliers, namely 10 % and 20 % of measurements (gene expressions) are discarded from the consequent computations.

Further, highly robust principal component analysis LWS-PCA of (Kalina, 2012) was computed from

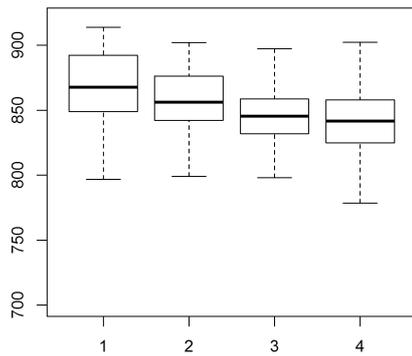


Figure 2: Example of Section 5: Estimated foreground intensities across 48 samples for a particular gene transcript without performing sharpening and averaging. The image shows averaged values for top, upper middle, bottom middle, and bottom rows of the strips.

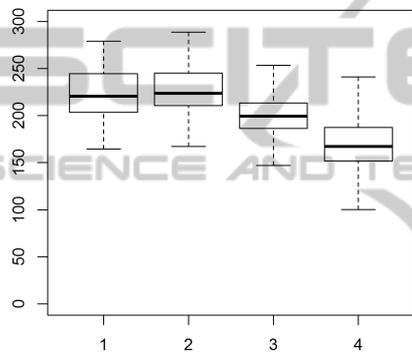


Figure 3: Estimated foreground intensities across 48 samples for a particular gene transcript with sharpening and averaging. This is exactly the result of the beadarray package with default settings of the parameters.

the data. The first row corresponds to the standard approach. The next rows included trimming off 10 % and 20 % of measurements, i.e. the fixed number equal to 10 % (or 20 % of beads are claimed to be outliers). An increase in the values in Table 1 shows that the major principal components perform better in extracting information from the data. In the standard approach, less important principal components are namely caused by a mere noise, but their influence is now reduced closer to zero and the methods can be interpreted as a denoising (Tibshirani et al., 2003).

Table 1: The contribution (in %) of the major 5 robust principal components to explaining the variability of the gene expression data for various percentage of outliers trimmed off.

Percentage of outliers	Index				
	1	2	3	4	5
0.9 %	6.4	6.1	5.0	4.3	3.9
10 %	7.8	7.4	5.9	5.2	4.0
20 %	8.8	7.8	6.3	5.4	4.1

6 CONCLUSIONS AND FUTURE RESEARCH

Microarrays remain to be a perspective technology for research in molecular genetics (Göhlmann and Talloen, 2009) and their role is believed to be still increasing (Rueda, 2014). We analyzed the code of commonly used methods of the beadarray package in C++, because their details (e.g. the formula (1)) have not been critically described in literature. This might have contributed to the fact that sufficient attention has not been paid to robustness properties of the standard methodology, not even in monographs on image analysis of microarrays (Rueda, 2014). This paper reveals some disadvantages of the standard image analysis of BeadChip microarrays.

We promote our position that the standard software for the image analysis of BeadChip microarrays, which is implemented in the beadarray package of Bioconductor, suffers from the presence of severe noise in the scanned images. Therefore, we see a need for software based on an alternative approach for the BeadChip image analysis. Our criticism of the standard approaches illustrated on examples with real data goes far beyond the mild review of (Smith et al., 2010). The problem is that results of the standard analysis are biased even on optimally prepared microarrays. As a consequence, we give a warning that results of all microarray genetic studies analyzed by the standard methodology should be interpreted carefully, because they are influenced by the high sensitivity of standard procedures with respect to the omnipresent contamination of data by noise and outlying values.

Currently, we are implementing a robust approach and tune its parameters. Such approach allows a specific treatment of outliers, spatial artifacts, and global trend across the strip of the microarray. In this context, pixels influenced by artifacts or severe noise do not have to be deleted, but carefully modeled, allowing to suppress the effect of noise. After having described some principles of robust image analysis of BeadChip microarrays in this paper, we propose a list of ideas or topics for a future research in this area.

- Measurement errors can be estimated as the variability of measured expression in beads with no corresponding targets in the genome.
- The type of gene probes or the precise location of beads may be identified wrongly with a relatively large probability, particularly for genes with a small expression.
- Beads corresponding to highly expressed genes have a tendency to transmit a part of their signal to

their neighborhood. Thus, beads with a small intensity may be overilluminated by a strong effect from their neighborhood (Figure 1).

- Errors in the process of bead localization on the microarray. They are typical for all beads, but larger for beads with a lower expression level.
- Errors in the process of bead type identification.
- Unrealistic assumption that the expressions of each gene have the same variability.
- Poisson distribution is more adequate for modeling the noise because the measured fluorescence intensities actually correspond to counts of individual photons.
- Sensitivity to discretization of coordinates. The result is highly influenced by fractional coordinates of the bead, because the assumption of linearity of the intensities of the image between neighboring pixels is strongly violated.

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REFERENCES

- Cardona, A. and Tomancak, P. (2012). Current challenges in open-source bioimage informatics. *Nature Methods*, 9:661–665.
- Davies, P. and Gather, U. (2005). Breakdown and groups. *Annals of Statistics*, 33:997–1035.
- Dunning, M., Barbosa-Morais, N., Lynch, A., Tavaré, S., and Ritchie, M. (2008). Statistical issues in the analysis of Illumina data. *BMC Bioinformatics*, 9(85).
- Dunning, M., M.L. Smith, M. R., and Tavaré, S. (2007). Beadarray: R classes and methods for Illumina bead-based data. *Bioinformatics*, 23:2183–2184.
- Filzmoser, P. and Todorov, V. (2011). Review of robust multivariate statistical methods in high dimension. *Analytica Chimica Acta*, 705:2–14.
- Fraser, K., Wang, Z., and Liu, X. (2010). *Microarray image analysis: An algorithmic approach*. Chapman & Hall/CRC, Boca Raton.
- Göhlmann, H. and Talloen, W. (2009). *Gene expression studies using Affymetrix microarrays*. Chapman & Hall/CRC, Boca Raton.
- Kalina, J. (2012). Implicitly weighted methods in robust image analysis. *Journal of Mathematical Imaging and Vision*, 44:449–462.
- Kalina, J. (2014). Classification analysis methods for high-dimensional genetic data. *Biocybernetics and Biomedical Engineering*, 34:10–18.
- Kalina, J. and Zvárová, J. (2013). Decision support systems in the process of improving patient safety. In *E-health Technologies and Improving Patient Safety: Exploring Organizational Factors*. IGI Global, Hershey, 71–83.
- Kuhn, K., Baker, S., Chudin, E., Lieu, M.-H., Oeser, S., Bennett, H., Rigault, P., Barker, D., McDaniel, T., and Chee, M. (2004). A novel, high-performance random array platform for quantitative gene expression profiling. *Genome Research*, 14:2347–2356.
- Rueda, L. (2014). *Microarray image and data analysis: Theory and practice*. CRC Press, Boca Raton.
- Shi, W., Banerjee, A., Ritchie, M., Gerondakis, S., and Smyth, G. (2009). Illumina WG-6 beadchip strips should be normalized separately. *BMC Bioinformatics*, 10(372).
- Smith, M., Dunning, M., Tavaré, S., and Lynch, A. (2010). Identification and correction of previously reported spatial phenomena using raw Illumina BeadArray data. *BMC Bioinformatics*, 11(208).
- Tanaka, G., Suetake, N., and Uchino, E. (2010). Image enhancement based on nonlinear smoothing and sharpening for noisy images. *Journal of Advanced Computational Intelligence and Intelligent Informatics*, 14:200–207.
- Tibshirani, R., Hastie, T., and Narasimhan, B. (2003). Class prediction by nearest shrunken centroids, with applications to DNA microarrays. *Statistical Science*, 18:104–117.
- Xanthopoulos, P., Pardalos, P., and Trafalis, T. (2013). *Robust data mining*. Springer, New York.