

SHANNON ENTROPY AND FRACTAL ANALYSIS FOR THE 16S RIBOSOMAL RNA AND COX2 MT-DNA SEQUENCES IN PRIMATES INCLUDING NEANDERTHAL

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Keywords: Mono-nucleotide entropy, Di-nucleotide entropy, Fractal dimension, Neanderthal mt-DNA COX2, Neanderthal mt-DNA 16S rRNA.

Abstract: The primate mt-DNA 16S rRNA and COX2 sequences, including Neanderthal sequences, were studied using nucleotide frequency, mono- and di-nucleotide entropy, and fractal dimension. The fractal dimension was computed with the Higuchi method when a nucleotide sequence is expressed as a numerical sequence where each nucleotide is assigned its proton number. The results shows that the C+G percent correlates with the fractal dimension with R-square value of around 0.88 ($N = 8$) for both gene sequences. The Di- and mono-nucleotide entropy is also well correlated with similar R-square values. For the COX2 gene, the human and Neanderthal cluster at high entropy suggests that chimp, gorilla, and orangutan were subjected to a higher selection pressure for this gene. The human COX2 has less entropy than the Neanderthal COX2 consistent with the presence of some selection pressure.

1 INTRODUCTION

Closely related species can be classified by a collection of phylogenetic markers with numbers determined from mathematical operations on DNA sequence. In cases where such markers are significantly different from what would be expected for random mutations, we can get an idea of how much natural selection has influenced the evolution of a particular gene. Here, we analyze primate mt-DNA 16S ribosomal RNA (rRNA) and COX2 sequences, including Neanderthal sequences, using nucleotide frequency, mono- and di-nucleotide entropy, and fractal dimension. For protein encoded by the mt-DNA genome, COX2 have experienced four amino acid substitutions on the modern human mtDNA lineage (Green et al, 2008). COX2 encodes a protein that maintains a proton gradient across the mitochondrial inner membrane, which drives the phosphorylation of ADP to ATP. The conclusion that the evolution of human COX2 from Neanderthal has been for minor adaptive advantages without significant functional consequences for mitochondrial function were put forward using the fact that the substitutions are on non-functional site

in the crystal structure. The relationship of the above mentioned phylogenetic markers would shed light whether there is selection pressure corresponding to some yet to be discovered significant function. The 16S rRNA is an important gene in classification and is included in this study.

The nucleotide base pair changes over a gene sequence can be viewed as a fluctuation and, consequently, can be investigated with standard tools that include correlation and fractal dimension analysis. For this study, the numerical sequence representing the fluctuation of nucleotides in a gene sequence was generated using the proton number of each nucleotide. This numerical series can then be processed further using numerical methods such as a moving average, which is often used in stock market time series analysis. The fractal dimension of such a random series or random series derived from the original atomic number based sequence can be computed. A recent comparison of human and chimpanzee genomes revealed that it is possible to measure the acceleration rate of the accelerated regions of the human genome (Pollard, et al, 2006a). The most accelerated region, HAR1, was shown by a gene expression experiment in the human embryo to be transcription active and co-expressed with

reelin, which is an essential protein involved in the development of the six-layer cortex of the human brain. Fractal analysis was applied to the HAR1 nucleotide sequence and the homologous sequence in the chimpanzee genome. Analysis shows that the differences in fractal dimension can be used as a marker of evolution. The 118-bp in HAR1 contains 18 point substitutions over an evolutionary span of 5 million years when comparing the human to the chimpanzee. However, the same 118-bp region only contains two point substitutions over a span of 300 million years when comparing the chicken to the chimpanzee. The implications of evolution and positive selection have been discussed in recent literature (Pollard, et al, 2006b).

2 MATERIALS & METHODS

The nucleotide sequences were downloaded from Genbank. The accession numbers of the mtDNA database are listed in the Appendix. The studied primates are human, Neanderthal, chimp (chimp and pygmy chimp), gorilla (western and western lowland), and orangutan (Bornean and Sumatran).

The ATCG sequence was converted to a numerical sequence by assigning the atomic number, the number of protons, to each of the nucleotides: A(70), T(66), C(58), G(78). The assigned number is roughly proportional to the nucleotide mass. This assignment was consistent with the recently reported mass fractal analysis of a ribosome sequence (Lee 2006). The A-T and C-G pairs in a double strand DNA would have the same value of 136.

Fractal dimension analysis can be used in the study of correlated randomness. Among the various fractal dimension methods, the Higuchi fractal method is well suited for studying signal fluctuation (Higuchi, 1998). The signal from the sequence represents a random spatial intensity series. The spatial intensity (Int) random series with equal intervals could be used to generate a difference series ($\text{Int}(j)-\text{Int}(i)$) for different lags in the spatial variable. The non-normalized apparent length of the spatial series curve is simply $L(k) = \sum \square \text{absolute}(\text{Int}(j)-\text{Int}(i))$ for all $(j-i)$ pairs from 1 to k . The number of terms in a k -series varies and normalization must be used to get the series length. If the $\text{Int}(i)$ is a fractal function, then the $\log(L(k))$ versus $\log(1/k)$ should be a straight line with the slope equal to the fractal dimension. Higuchi incorporated a calibration division step (divide by k) such that the maximum theoretical value is calibrated to the topological value of 2. The detailed

calculation is given in the literature (Higuchi, 1998). When comparing the dimension of two fractal forms, the popular method of taking the difference of the two Higuchi fractal dimension values is valid to within a constant regardless of the calibration division step. The Higuchi fractal algorithm used in this project was calibrated with the Weierstrass function. This function has the form $W(x) = \sum a^{-nh} \cos(2\pi a^n x)$ for all the n values 0, 1, 2, 3... The fractal dimension of the Weierstrass function was given by $(2 - h)$ where h takes on an arbitrary value between zero and one.

The Shannon entropy of a sequence can be used to monitor the level of functional constraints acting on the gene (Parkhomchuk, 2006). A sequence with relatively low nucleotide variety would have a low Shannon entropy (more constraint) in terms of the set of 16 possible di-nucleotide pairs. A sequence's entropy can be computed as the sum of $(p_i) \log(p_i)$ over all states i and the probability p_i can be obtained from the empirical histogram of the 16 di-nucleotide-pairs. The maximum entropy is 4 binary bits per pair for 16 possibilities (2^4). The maximum entropy is 2 bits per mono-nucleotide with 4 possibilities (2^2).

3 RESULTS & DISCUSSION

For the 16S rRNA sequences, the C+G percent correlates with fractal dimension, FD, with R-square value of 0.88, $N = 8$, in Figure 1. Dropping human and Neanderthal data would increase the R-square value to ~ 0.91 because the data are in the middle as small outliers.

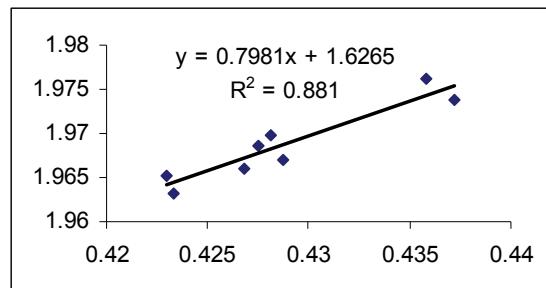


Figure 1: The C+G percent (x-axis) versus FD (y-axis) for the studied 16S rRNA sequences.

The mono-nucleotide entropy correlates with di-nucleotide entropy in the 16S rRNA sequence with R-square value of ~ 0.88 , $N = 8$ (Figure 2). Dropping human and Neanderthal increases R-

square value of ~ 0.99 because they are in the middle as moderate outliers

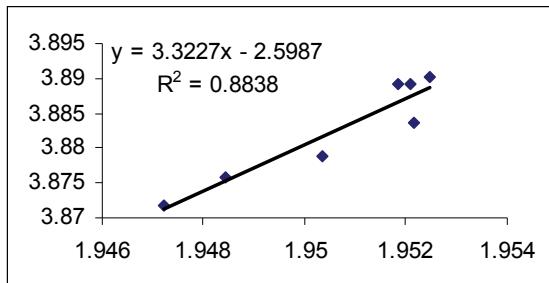


Figure 2: The mono-nucleotide entropy (x-axis) versus the di-nucleotide entropy (y-axis) for the studied 16S rRNA sequences.

Similar correlation of the C+G percent with FD is observed for COX2 sequences R-square value of ~ 0.8756, N = 8 (Figure 3). Dropping the human and Neanderthal data would increase the R-square value to ~ 0.93 because they are in the middle as moderate outliers.

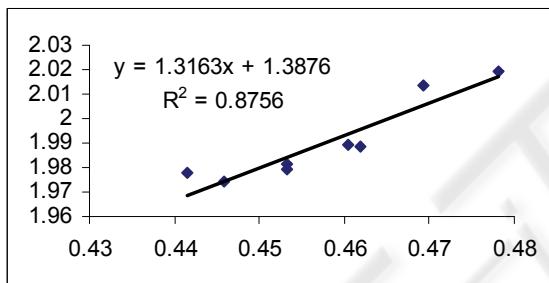


Figure 3: The C+G percent (x-axis) versus FD (y-axis) for the studied COX2 sequences.

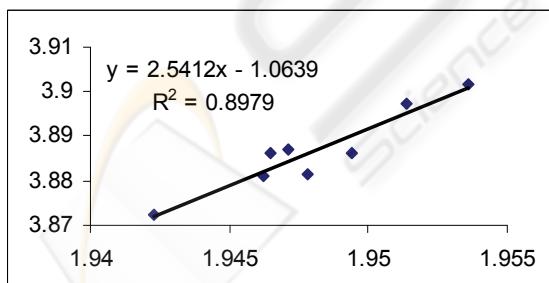


Figure 4: The mono-nucleotide entropy (x-axis) versus the di-nucleotide entropy (y-axis) for the studied COX2 sequences.

Similar correlation of the single entropy with pair entropy is observed for COX2 sequences R-sq ~ 0.8979 (Figure 4). The interesting point is that if human and Neanderthal are deleted then the correlation drops to 0.7601 because they are the two

end points at large values (mono-nucleotide entropy for Neanderthal ~ 1.953 bits, for human 1.951 bits).

Correlation would suggest similar selection pressure mechanism. Conserved regions would imply less random nucleotide fluctuation across species and the fractal dimension would differ from 2 and the di-nucleotide Shannon entropy would be smaller than 4 bits. Our previous results have associated fractal dimension with functionality (Tremberger, Jr., et al., 2009). The observation of fractal dimension correlation to the C+G percent (which is correlates to the A+T percent) shows that human and Neanderthal occupy the mid range region and thus would indicate a mild selection pressure on the functionality of the 16S ribosomal RNA and COX2 sequences as compared to the other primates. Our previous results have associated entropy with constraints (Holden et al, 2008). The high-value positions of the COX2 sequences in the single-entropy and pair entropy correlation suggests a weaker constraint in human and Neanderthal as compared to other primates. Di-nucleotide frequency distributions confirm the closeness of relations among the various primates. For the COX2 genes, lowered entropy is largely a manifestation of high Cytosine (~30 %) and low Guanine (~15 %) content. Di- and mono-nucleotide entropy is well correlated. That human and Neanderthal cluster at high entropy indicates that chimp, gorilla, and orangutan were subjected to a higher selection pressure for this gene. The human COX2 has less entropy than the Neanderthal COX2 consistent with the presence of some selection pressure.

Whether the mild selection pressure and the weak constraint observed in the mtDNA 16S rRNA and COX2 sequences have activated other selection response such as those observed in the brain function related HAR1 sequence is an interesting consideration. The HAR1 hardly evolved in 300 million years from chicken to chimp and then experienced accelerated selection pressure. The fractal analysis shows that the HAR1 has a fractal of 2.02 while the chimp is at 1.97, the di-nucleotide entropy is 3.86 bits for human and 3.64 bits for chimp; and single entropy is 1.97bits for human, 1.86 bits for chimp. If the HAR1-like activation exists such that the selection pressure would be mild on the mt-DNA, it would suggests that the Neanderthal would have similar HAR1-like response as their 16S rRNA and COX2 sequences are very similar to the human's in terms of the above studied parameters. The activation would be an indication of cooperation in multi-cellular organism. In this regard the CNV (copy number variants) strategy

should also be considered. It is possible that less demand on the Cs and Gs with multiple copies could be more advantageous than higher demand on Cs and Gs bias with fewer copies for responding to selection pressure. The correlation analysis method can be a good supplemental tool to the relative comparison method inherent in the BLAST method.

4 CONCLUSIONS

The nucleotide fluctuation of the mtDNA 16S rRNA and COX2 gene sequences in primates including Neanderthal were studied. The tools are fractal dimension, Shannon mono- and di-nucleotide entropy, and C+ G content. We found that C+G content correlates with the fractal dimension. The correlation of the mono- and di-nucleotide entropy shows that the human COX2 gene has experienced some selection pressure. Future studies include the extension to other species.

ACKNOWLEDGEMENTS

The project was partially supported by several CUNY PSC and Collaborative grants. N.G. received partial support from CUNY Collaborative CCIRG and Perkins Grant. E.C. thanks the hospitality of QCC. We thank the research groups for posting their gene sequence data in Genbank.

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APPENDIX

The studied sequences were downloaded from the Genbank mtDNA database. The accession numbers are:

NC_001644 Pan paniscus mitochondrion, (pygmy chimpanzee).

NC_001643 Pan troglodytes mitochondrion, (chimpanzee).

NC_011120 Gorilla gorilla gorilla mitochondrion (Western lowland gorilla).

NC_001645 Gorilla gorilla mitochondrion (Western Gorilla).

AC_000021 Homo sapiens mitochondrion, (human).

NC_011137 Homo sapiens neanderthalensis mitochondrion, (Neanderthal).

NC_001646 Pongo pygmaeus mitochondrion (Bornean orang-utan).

NC_002083 Pongo abelii mitochondrion, (Sumatran orangutan).