Towards More Robust Transcription Factor Binding Site Classifiers Using Out-of-Distribution Data

István Megyeri[1][a] and Gergely Pap[1,2,3][b]

[1]*University of Szeged, Dugonics Square 13, Szeged, 6720, Hungary*
[2]*HUN-REN-SZTE Research Group on Artificial Intelligence, Szeged, Hungary*
[3]*Princess Margaret Cancer Centre, 610 University Ave. Toronto, ON M5G 2M9, Canada*

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Abstract: The use of deep learning methods for solving tasks in computational biology has increased in recent years. Many challenging problems are now addressed with novel architectures, training strategies and techniques involved in deep learning such as gene expression prediction, identifying splicing patterns, and DNA-protein binding site classification. Moreover, interpretability has become a key component of those methods used to solve computational biology tasks. Gaining a novel insight by analyzing the learners is a key factor. However, most deep learning models are hard to interpret, and they are prone to learn features which generalize poorly. In this study, we examine the robustness of high performing neural networks using in-distribution (ID) and out-of-distribution (OOD) examples. We demonstrate our findings in two different tasks taken from the domain of DNA-protein binding site classification and show that the overconfident and incorrect predictions are a result of the training data that has been built exclusively from ID samples. Adding OOD data to the training process enhances the reliability of the networks and it improves the performance on the ID tasks.

1 INTRODUCTION

Transcription factors (TFs) are proteins that regulate gene expression. They exert their influence through binding to the strands of DNA. Disruptions in the regulatory mechanism of TFs are associated with several diseases like some types of cancer (Vishnoi et al., 2020; Lambert et al., 2018). Therefore, understanding the way TFs work in healthy and unhealthy circumstances is an important research area. Sequencing techniques such as ChIP-seq and CUT&RUN provide information about the segments of DNA where TF binding occurs. However, these experiments are costly and not completely accurate. Consequently, exploring other methods to predict TF binding might prove advantageous.

Recently, machine learning and deep learning methods have advanced the state of the field regarding TFs by being able to detect their binding sites. These detections rely on using the large amount of data produced by Next-Generation-Sequencing (NGS) techniques, then a trained machine learning model is used to examine the TFs. Extracting such regular patterns from neural networks can describe nucleotide sequences where the binding occurs, this being analogous to Position Weight Matrices (Alipanahi et al., 2015).

Altering neural network architectures to enhance prediction performance has received significant attention in the field. Early methods relied on convolutional neural networks with nucleotide sequences as input (Alipanahi et al., 2015; Qin and Feng, 2017). Then recurrent neural networks and attention mechanisms were adopted for Transcription Binding Site (TFBS) classification (Hassanzadeh and Wang, 2016; Park et al., 2020). Often interpretability and reliability of the models are expected to improve owing to these improvements. Here, we claim that novel architectures are not sufficient and better training strategies need to be developed to enhance reliability.

Including other modalities in the training process (Chiu et al., 2023; Wang et al., 2023), and different ways of representing the input sequence can help models learn patterns that are not feasibly recognizable from just using the base pairs. Methods that rely on physico-chemical or conformational, or epigenomics features are likely to outperform nucleotide

[a] https://orcid.org/0000-0002-7918-6295
[b] https://orcid.org/0000-0002-6641-5845

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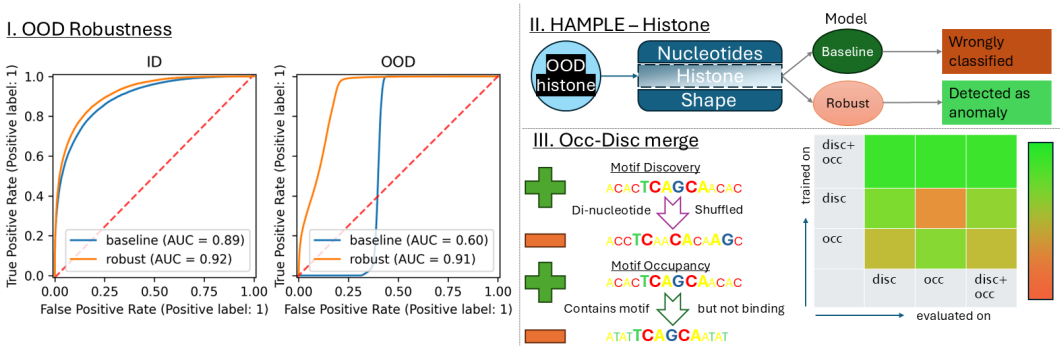

Figure 1: Networks trained and evaluated on in-distribution samples show high performance. However, when evaluated on OOD examples their performance suffers irrespective their training methods. More robust models can be built by augmenting training with out-of-distribution batches for uni-modal as well as multi-modal learners. For multi-modal, the OOD samples are generated by modifying a single feature of the multi-modal input. Training a uni-modal network on one (occupancy) classification subtask and evaluating on another (discovery) yields poor results - even though the underlying classification problem is the same. In contrast, when the model is trained with both subtasks in parallel, the accuracy increases.

based learners. While physico-chemical properties can be calculated from the nucleotide sequences on a di- or trimer basis, many useful epigenomic features rely on data sources and experiments that are hard to come by. For example adding a histone modification signal to a machine learning model will most likely increase the performance. But the source of histone marks is also a ChIP-seq experiment, thus limiting the real-world impact of the computational method, since both the output's and the input's data relies on wet-lab techniques that require considerable resources to carry them out.

In this study, we design methods for generating out-of-distribution (OOD) samples that can be combined with novel data sources i.e. for multi-modal network training and also improving uni-modal nucleotide sequence learners. We mainly rely on domain knowledge to generate an inconsistent input which does not exist in the real world so the TFs shall not bind to them. The generation is suitable for multi-modal networks where we mix binding and non-binding inputs by keeping one or more modalities untouched while the other is replaced from a different class sample. We challenge also uni-modal models by shuffling their binding sequences and present them as non-binding sequences. We investigate multiple approaches of mixing samples for multi-modal as well as uni-modal networks. The presented mixing and shuffling mechanism destroy the underlying structure of binding site so it can be a reliable source for model evaluations.

Interestingly, we find that despite additional data sources or novel architectures, generated OOD samples are a significant challenge even for state-of-the-art networks (Han et al., 2021; Wang et al., 2023). We note that the algorithms' performance drops when evaluated on out-of-distribution samples. Therefore, these models do not adequately learn the mechanisms of TFBSs and need to be improved by training on OOD data.

To address this problem, we propose novel training strategies to enhance the reliability of the models on OOD samples. We leverage the generated data while ensuring that we evaluate on OOD samples the model was not trained on. Moreover, we present a multi-task training method that can be used to enhance model performance via examples where the ground truth is only partially available. The proposed methods enhance the neural networks representation on out-of-distribution samples without any extra cost, and in certain cases the in-distribution performance is significantly improved.

Our main findings and the overall pipeline for increasing robustness by training with additional OOD entities is shown in Fig. 1. The source code is available at https://github.com/szegedai/tf_ood_robustness.

2 RELATED WORK

2.1 Connection with Interpretability and Robustness

Most TFBS prediction models prioritize interpretability through methods like saliency maps, gradient visualizations, or motif extraction. Robustness is essential in approaches like these, which are often applied for personal genome diagnostics or epigenetic analysis where models must handle OOD inputs and explain their predictions effectively. However, these approaches often neglect robustness, leading to overconfident predictions based on statistical noise or irrelevant features.

Adversarial examples highlight this issue by exposing how easily models can be misled, undermining interpretability. Training with adversarial or modified examples has been shown to improve robustness and align learned features with human understanding (Geirhos et al., 2019). Our prior work (Pap. and Megyeri., 2022) demonstrated that TFBS prediction models perform poorly with modified sequences, in which binding sites remain intact but nucleotides are either cut from or appended to the sides. Augmenting training data with shifted or cropped sequences improved both robustness and performance. We note that domain generalization techniques and transfer learning can also improve robustness.

Here, we focus on TFBS classifiers' sensitivity to OOD samples which can also characterize the learner's internal features. We incorporated OOD data into regular training approaches designed specifically for the two TFBS tasks defined in Section 3. These approaches are applicable to different tasks too, and help increase performance and robustness through using OOD examples in training.

2.2 Generalization to Other Cell Lines

A key goal in DNA-protein binding detection is generalizing well to new cell types, as experimental data for many TF/cell type pairs are unavailable. Successful predictions across cell-types, as demonstrated in Virtual ChIP-seq (Karimzadeh and Hoffman, 2022), increase confidence in the meaningful biological characteristics captured. However, performance varies significantly between TFs and cell types. Models could fail to recognize critical statistical signals (Schreiber et al., 2020). Improvements can be made to nucleotide-based learners by augmenting training with point mutations (Lee et al., 2024). Integrating additional data modalities like physico-chemical descriptors and epigenomic features is advantageous. (Chiu et al., 2023; Wu et al., 2024).

In this study, we show that additional features of multi-modal inputs alone are not sufficient to detect OOD samples, and incorporating OOD samples into the training process provides an orthogonal improvement. Moreover, we demonstrate that the OOD data can be generated from ID data so it has no extra costs.

2.3 Classification with Convolutional Networks

Many TFBS detection models rely on convolutional neural networks (CNNs) with 1–3 layers, designed to learn binding motifs from one-hot encoded sequences as a binary classification task. Shallow architectures enable effective motif extraction, as deeper networks tend to fragment binding information over layers (Koo and Eddy, 2019). DeepSEA (Zhou and Troyanskaya, 2015) tackles the prediction of multiple TFs, which is more challenging. More recently, MAResNet (Han et al., 2021) addresses the two issues detailed above. By applying the well-known architectural choices developed for computer vision tasks, a ResNet-like architecture is employed for binding site prediction. In addition, pre-training is used, resulting in the learners having information about multiple TFs and this gives better performance when evaluated on downstream tasks.

In our study, we evaluate both ResNet-like networks and CNNs with attention mechanisms. Both types show a performance drop when evaluated on OOD data, and are successfully improved with the proposed training methods.

3 EXPERIMENTAL SETUP

Let us first introduce our notations and metrics. We will train neural networks for binary classification problems to recognize TFBSs. An example is given as (x,y), where $x \in \mathbb{R}^d$ and $y \in \{0,1\}$. d denotes the dimension of the input. A binary classifier network is denoted by $f_\theta : \mathbb{R}^d \to \mathbb{R}^1$, using the following cost function:

$$\min_\theta \mathcal{L}(f) = y \cdot \log f(x) + (1-y) \cdot (\log(1-f(x))) \quad (1)$$

To measure the performance of a trained network, we will use accuracy, AUC, and AUPR. Accuracy is defined as the ratio of the correctly classified inputs in a given set. Area Under the ROC Curve(AUC) measures the area under the true positive rate vs. false positive rate curve, while AUPR measures the area under the precision vs. recall curve. Accuracy will only change when the prediction exceeds the 0.5 threshold. In contrast, AUC and AUPRC can reflect smaller improvements with a more appropriate threshold.

3.1 Datasets

Here, we first describe the uni-modal dataset and then the multi-modal dataset. In each, we define the OOD data generation methods and their notations.

For uni-modal experiments, we use the ENCODE DREAM5 Transcription Factor Binding Site Challenge data. In (Zeng et al., 2016) the authors defined two tasks, namely discovery and occupancy (denoted as Occ and Disc, respectively) for 422 and 690

binding prediction tasks. Stated briefly, in the discovery task, the positive entities are sequences containing binding sites based on ChIP-seq peaks, whereas the negative ones are di-nucleotide shuffled versions of the positive ones. Occupancy defines the positive class similarly, but the negative sequences are chosen based on them containing the binding motif pattern but having no experimentally verified peaks. Generally speaking, occupancy proves to be a more difficult task for the typical convnets with nucleotide sequence inputs. Here, we define two scenarios for training and evaluation. First, we simulate unseen OOD data via training on occupancy and testing on discovery or vice-versa. We will refer to the in-distribution data as $\mathcal{D}_{in}$ and use $\mathcal{D}_{out}$ for the unseen data distribution. In addition to this, we also take the union of the occupancy and discovery data denoted as $\mathcal{D}_{all}$ and train models over the joint distribution. During evaluation, we will focus on the observed performance for the following three TFs: MAFK, SP1 and ZNF147. These three were selected in order to represent a different level of difficulty in terms of the classification task. We refer to this as $\mathcal{D}_3$. Note that we also train models over a larger set with 422 TFs using both the occupancy and the discovery data which is referred as $\mathcal{D}_{422}$.

For multi-modal experiments, we use the dataset employed to train HAMPLE (Wang et al., 2023). The goal of the classification task in this case is to predict the correct cell type for which a given TF binds the input sequence. The model relies on three different modalities, namely nucleotide sequence, physico-chemical and 3D DNA properties (in brief: 'shape'), and histone modifications. The nucleotide sequence belonging to a given bin does not vary regardless of cell type (or class label). The physico-chemical properties are calculated from the nucleotide sequence, so the same is true. Histone marks, however is different.

Unlike in uni-modal data, there are no different variants of the same problem. Therefore, we define different strategies to generate OOD samples. Sample generation is done by manipulating the histone modifications of the entities. That is, without disrupting nucleotide or shape information, the signal values belonging to the histone marks were rendered independently of the correct class label for the OOD entities. In this way, the model could not rely on the most informative modality for cell type classification. Several methods were developed to create the out-of-distribution entities' histone values. As mentioned above, in each case the histone features belonging to an augmented example were misleading: they contain no information about the correct label. First, the histone values were substituted with randomly generated numbers from a normal distribution $\mathbb{N}$ characterized by the mean and the standard deviation of the original histone values – denoted as $\mathcal{D}_{\mathbb{N}}$. Second, the histone values were substituted using all of the other possible histone values which do not belong to the given entity's class. In other words, given a sample, its histone values were changed to a random one belonging to a different class – denoted as $\mathcal{D}_{bs}$ - batch-switch. Third, a subset of the original in-distribution batch (e.g., 25%) was duplicated and then its histone values were shuffled only using the values present in that current batch. During shuffling the only criteria was that the new, out-of-distribution histone properties must belong to a class different from the original one – denoted as $\mathcal{D}_{mixin}$. Creating OOD entities using a normal distribution allows for an almost infinite number of combinations. However, $\mathcal{D}_{\mathbb{N}}$ might be the least similar to the ID examples. Using $\mathcal{D}_{bs}$ makes use of the whole dataset, by switching the histone modification in the currently generated batch, ensuring many new OOD example creations. For $\mathcal{D}_{mixin}$ the number of possible combinations is limited by the present batch's size and the class distribution in it. Still, we hypothesize that the mixin strategy forces the learner to exploit nucleotide and shape information quite severely.

Table 1: Settings for the used ResNet configurations.

Depth	Width	Number of parameters
10	1	0.028M
	2	0.061M
16	1	0.093M
	2	0.109M
	10	6.152M
22	1	0.126M
	2	0.159M
28	1	0.192M
	2	0.239M
	10	12.612M
34	1	0.369M
	2	0.499M
40	1	0.629M
	2	0.759M
	10	19.073M

3.2 Uni-Modal Experiments

We define two training strategies over different sets of data: single-task and multi-task training. For multi-task training, we define two subtypes based on the amount of data that is used. Next, these are introduced in more detail.

In (Shen et al., 2021; Han et al., 2021), it is demonstrated that ResNet-based models can efficiently learn binding motifs for multiple transcription factors (TFs) simultaneously. In our setup, we will

use wide residual architecture (Zagoruyko and Komodakis, 2016) with 1D convolutions. Since we will train models on different dataset sizes, thus we need to adjust the network sizes for the given dataset. Therefore, we explore network depth from 10 layers up to 40 and width parameters of $\{1,2,10\}$. This will allow us to examine the performance of different training strategies as a function of network capacity. The list of all network configurations and their parameter values are presented in Table 1.

All residual networks are trained for 10^5 iterations using the SGD optimizer with a learning rate of 10^{-1} and weight decay of 0.0005. For networks with a width of $\{1,2\}$ the learning rate is reduced according to the cyclic learning rate schedule (Smith, 2017) with a cycle length of 10^4 iterations. In the case of networks with a width of 10, we observed poor convergence so we used cosine schedule (Loshchilov and Hutter, 2017) with a single cycle during their training. The batch size was set to 192 and 3376 for $\mathcal{D}_3$ and $\mathcal{D}_{422}$ datasets, respectively. During training, the weights are evaluated at every 1000th step using a validation set and the checkpoint which provides the best validation accuracy is saved.

Single-Task Training. As baselines, we train residual networks for single transcription factor binding site prediction using one type of negative example, derived either from occupancy or discovery. The training objective is binary cross-entropy. During the evaluation phase, we might make predictions on occupancy or discovery regardless of what the model was trained on. We define two evaluation scenarios: **s-in** and **s-out** for single-task models. In s-in, all models are evaluated on the corresponding test set a.k.a. $\mathcal{D}_{in}$ (i.e., occupancy models are only used for occupancy data and discovery models are exclusively used for discovery test sets). This evaluation will show the performance of the models under ideal circumstances, which is unrealistic. In s-out - a more realistic evaluation - the models are challenged via cross-task evaluations using $\mathcal{D}_{out}$ (i.e., occupancy models are used to predict discovery and vice versa).

Multi-Task Training. To improve OOD detection performance, we train multi-task networks using three unions of the datasets. MD means the union of all TF datasets but only using negative samples from the discovery task. Similarly, MO means the network is trained on all of the available TFs using the occupancy datasets only. The union of MO and MD is referred to as ALL. For model training, multi-task models used the binary cross-entropy loss function. However, we excluded the loss of those output neurons where the ground truth is undefined (i.e., when predicting on MAFK input, Sp1 output labels are not present). Similar to single-TF model evaluations, we also define **m-out** and **m-in**. In m-in, a combination of the models is taken so that they are only used on in-distribution data. The m-out case is the reverse, and will reflect the results with cross-evaluations (i.e., the models are adversely used). For the networks which are trained on the ALL set we can only evaluate in-distribution performance that is referred to as **all**. This can represent an upper bound of what we might expect from an outlier detection method.

3.3 Multi-Modal Experiments

f_{ID} is defined as the original, baseline HAMPLE model, as proposed in (Wang et al., 2023). The robust learners are introduced next. First, $f_{\mathbb{N}}$ relies on a Gaussian distribution to augment histone properties. (Similarly to how $\mathcal{D}_{\mathbb{N}}$ is generated.) Second, an out-of-distribution batch was created in addition to the original in-distribution batch – denoted as f_{bs} - batch-switch. In the former the histone values were substituted using all of the other possible histone values which do not belong to the given entity's class. In other words, given a sample, its histone values were changed to a randomly selected one belonging to a different class, for all classes. This OOD batch has four times more examples then the ID one. During loss calculation the ID and OOD losses were multiplied by 0.8 and 0.2 before summation, respectively. (The multiplication values were established empirically.) Third, a subset of the original in-distribution batch (e.g., 25%) was duplicated and then its histone values were shuffled only using the values present in that current batch. During shuffling the only criteria was that the new, out-of-distribution histone properties must belong to a class different from the original one – denoted as f_{mixin}. Creating OOD entities using a normal distribution allows for an almost infinite number of combinations. However, $f_{\mathbb{N}}$ might be the least similar to the ID examples. Using $\mathcal{D}_{bs}$ makes use of the whole dataset, by switching the histone modification in the currently generated batch, ensuring many new OOD example creations. For $\mathcal{D}_{mixin}$ the number of possible combinations is limited by the present batch's size and the class distribution in it. On the other hand, we hypothesize that the mixin strategy forces the learner to exploit nucleotide and shape information most stringently. For evaluation three datasets are used. $\mathcal{D}_{in}$ is the original test set and is used for measuring the models' performance for solving the base TFBS detection task. For evaluating robustness the original test set is duplicated (so that there are twice the number of examples when compared with $\mathcal{D}_{in}$) and the histone properties are mod-

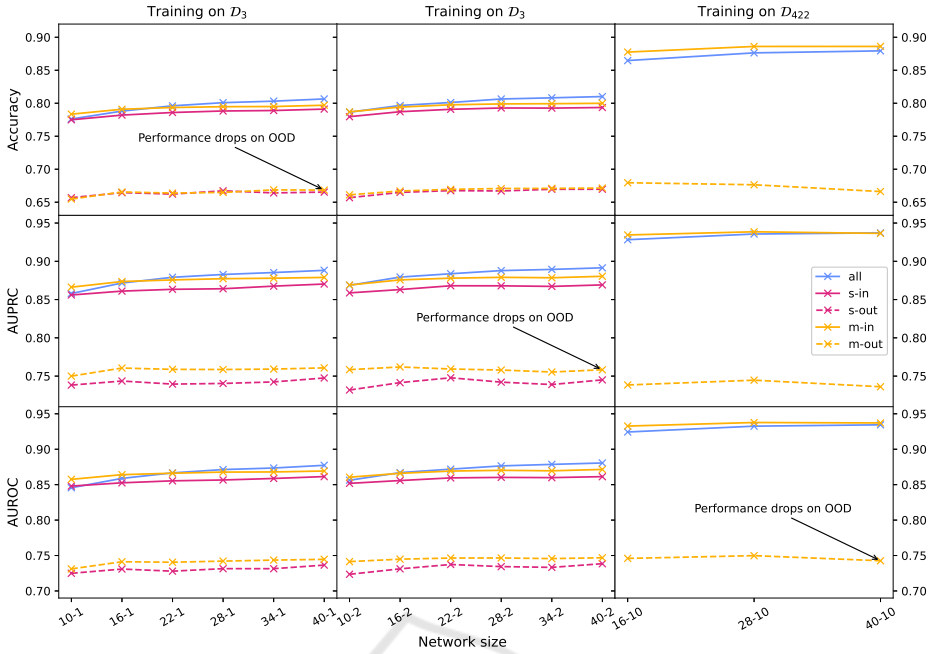

Figure 2: Accuracy, AUPRC, and AUROC scores are displayed as the function of network size and single(**s-in/out**) vs multi-task training(**m-in/out**,**all**) for in and out distribution when applicable. Performance drops when models are evaluated on OOD data for both multi-task as well as single-task models irrespective of the network sizes. A small but consistent improvement is visible for **m-out** in the case of AUPRC and AUROC metrics. In addition to the improved OOD detection capability, multi-task networks(**m-in** and **all**) outperform single-task networks for the in-distribution task, especially for larger networks.

ified in the newly created part. $\mathcal{D}_{\mathbb{N}}$ includes entities where the values of the histone marks are replaced from a Normal distribution while preserving the inherent statistical properties. For $\mathcal{D}_{sub}$ the duplicated part's histone features are substituted with values belonging to a different class, selected randomly. During the robustness test, the unmodified part is labelled as 1 and the modified as 0. Then using an Area Under the Receiver Operating Characteristic Curve the classifiers' ability to separate the two parts is measured.

10% of the training set was split for validation. Early stopping with a criteria of 5 epochs was employed, the maximum number of epochs was 25. For training the binary cross-entropy loss with the ADAM optimizer was used with a learning rate of 5×10^{-4}. The learning rate was reduced by 0.5 is no improvements were observed in terms of the validation loss for 3 epochs during training. The batch size was 64. For the robust training runs the number of convolutional neurons was multiplied by four. This resulted in the increase of the number of trainable parameters from 167,049 to 377,121. When using the increased size with f_{ID}, overfitting became an issue. Although in a few cases the larger ID networks performed better than the original ones - both on ID and robustness tasks. We use the higher measurement in our comparisons for each case.

4 RESULTS

Single- and Multi-Task ResNets. Accuracy, AUPRC, and AUROC scores are displayed as the function of network size and single vs multi-task training in Fig. 2. We also show the in and out distribution performance of the models when applicable.

Our first observation is that the models' performance drops by 0.1 in all the metrics when they are evaluated on OOD data for both multi-task as well as single-task models irrespective of the network sizes. This highlights the risk of the current practice of training and testing on a single distribution, one might not see the model learned specific features that only apply to the training distribution and does not allow general inspection of the TF's behavior.

The **all** curves in Fig. 2 show that when the same architecture is trained in conjunction with a merged dataset (e.g., both tasks' entities are included during fitting), the network can solve the task reliably, and the combined training also provides a small boost in performance. A small but consistent improvement is visible for **m-out** in the case of AUPRC and AUROC metrics. However, training on all the data results in a significantly better performance which shows the need for more appropriate training algorithms and OOD data generation methods.

In addition to the improved OOD detection capability, multi-task networks (m-in and all) outperform single-task networks for the in-distribution task, especially for larger networks. Further increasing the number of tasks and training a single model that can recognize 422 TFs on $\mathcal{D}_{422}$ boosts the performance by a large margin with respect to all the metrics. This highlights that multi-task training provides better representations. We note that even though we had to scale up the networks to millions of parameters to provide competitive performance relative to single-task models, it is still more compute- and parameter-efficient than training 422 individual networks. Based on Table 1, a single-task network on $\mathcal{D}_{422}$ with an architecture of WRN-40-1 would have $2 \times 422 \times 0.629\text{M} = 530.876\text{M}$ parameters while a WRN-40-10 multi-task network with $19.073M$ provides better results.

For Histone Modifications with HAMPLE. In the following the results for ID and OOD performance of the models based on the HAMPLE architecture are shown. The first two columns of 2 show the TF and training method. f_{ID} or baseline means the original normal/unmodified training setting as described in (Wang et al., 2023). The $\mathcal{D}_{in}$ column shows the AUROC performance on the test set for the classification task. The third and fourth columns show the robustness of the networks measured in AUROC when the task is the separation of the unmodified and the modified examples based on the test sets. $\mathcal{D}_{\mathbb{N}}$ or random normal means that the histone values were changed to random samples from a Gaussian distribution, for all classes. The $\mathcal{D}_{sub}$ substitute randomly method considered the original class labels during substitution, so that the new replacement values are from a different class in every case - similarly to f_{bs}.

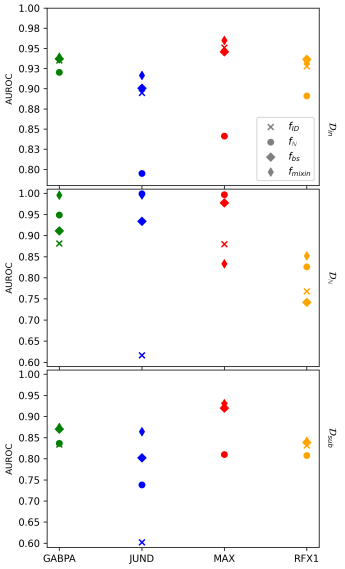

Figure 3: HAMPLE evaluated on ID and OOD examples.

Table 2: Test performance and robustness when evaluating on ID and OOD sets for multi-modal networks.

TF	Network	Evaluation data		
		$\mathcal{D}_{in}$	$\mathcal{D}_{\mathbb{N}}$	$\mathcal{D}_{sub}$
GABPA	f_{ID}	0.9352	0.8811	0.8337
	$f_{\mathbb{N}}$	0.9204	0.9487	0.8366
	f_{bs}	0.9372	0.9109	0.8704
	f_{mixin}	**0.9391**	**0.9957**	**0.8740**
JUND	f_{ID}	0.8949	0.6163	0.6022
	$f_{\mathbb{N}}$	0.7951	**0.9988**	0.7382
	f_{bs}	0.9006	0.9336	0.8023
	f_{mixin}	**0.9167**	0.9962	**0.8639**
MAX	f_{ID}	0.9510	0.8796	0.9265
	$f_{\mathbb{N}}$	0.8414	**0.9970**	0.8099
	f_{bs}	0.9458	0.9777	0.9200
	f_{mixin}	**0.9601**	0.8333	**0.9306**
RFX5	f_{ID}	0.9278	0.7679	0.8313
	$f_{\mathbb{N}}$	0.8910	0.8258	0.8079
	f_{bs}	**0.9365**	0.7418	0.8385
	f_{mixin}	0.9336	**0.8522**	**0.8417**

For the TF GABPA in Table 2 the original network's test AUROC is 0.9352. Although the training method $f_{\mathbb{N}}$ fails to outperform this, both the f_{mixin} 'mixin' and f_{bs} 'batchswitch' networks achieve more (0.9391 and 0.9372, respectively). In terms of robustness all modified learners manage to handle out-of-distribution entities better (AUROC for f_{mixin} is 0.9957 compared to the 0.8811 observed for the f_{ID} unmodified net - which results in a difference of 0.1146). Regarding $\mathcal{D}_{sub}$ evaluation, f_{mixin} and f_{bs} was better by about 0.04 AUROC score compared to f_{ID} and $f_{\mathbb{N}}$ learners. In summary f_{mixin} training proves to be the best in all categories. For MAX and JUND f_{mixin} shows better performance in $\mathcal{D}_{in}$ and $\mathcal{D}_{sub}$ evaluation, meanwhile for random histone marks the $f_{\mathbb{N}}$ training augmentation produced almost 100% separation. For RFX1 the f_{bs} network outperform in terms of ID, but f_{mixin} is observed to be more robust.

Fig. 3 shows evaluations using the different sets (top: $\mathcal{D}_{in}$: in-distribution test AUROC, middle: $\mathcal{D}_{sub}$: ID-OOD separation AUROC for histone values generated from a Gaussian distribution, bottom: $\mathcal{D}_{sub}$: ID-OOD separation AUROC for histone values sampled from a class different from the original). In terms of ID performance, the addition of extra OOD entities while increasing the network size provides better AUROC values. Moreover when measuring robustness, in almost every case the ID learner is outperformed by the augmented models.

5 CONCLUSIONS

The performance of DL for predicting transcription factor binding sites has improved in recent years.

However, the issue of robustness and generalization ability to handle OOD entities deserves further investigation. Here we provide two examples for two classification tasks, where the models fail to generalize well to OOD samples. We propose robust training techniques where we introduce OOD entities during fitting, and train multi-task models. We find that the performance with OOD examples increases. Future directions include examining other OOD scenarios and making comparisons with different adversarial training settings.

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REFERENCES

Alipanahi, B., Delong, A., Weirauch, M. T., and Frey, B. J. (2015). Predicting the sequence specificities of dna- and rna-binding proteins by deep learning. *Nature Biotechnology*, 33(8):831–838.

Chiu, T.-P., Rao, S., and Rohs, R. (2023). Physicochemical models of protein–dna binding with standard and modified base pairs. *Proceedings of the National Academy of Sciences*, 120(4):e2205796120.

Geirhos, R., Rubisch, P., Michaelis, C., Bethge, M., Wichmann, F. A., and Brendel, W. (2019). Imagenet-trained CNNs are biased towards texture; increasing shape bias improves accuracy and robustness. In *International Conference on Learning Representations*.

Han, K., Shen, L.-C., Zhu, Y.-H., Xu, J., Song, J., and Yu, D.-J. (2021). MAResNet: predicting transcription factor binding sites by combining multi-scale bottom-up and top-down attention and residual network. *Briefings in Bioinformatics*, 23(1):bbab445.

Hassanzadeh, H. and Wang, M. D. (2016). Deeperbind: Enhancing prediction of sequence specificities of dna binding proteins. In *2016 IEEE International Conference on Bioinformatics and Biomedicine (BIBM)*, pages 178–183, Los Alamitos, CA, USA. IEEE Computer Society.

Karimzadeh, M. and Hoffman, M. M. (2022). Virtual chip-seq: predicting transcription factor binding by learning from the transcriptome. *Genome Biology*, 23(1):126.

Koo, P. K. and Eddy, S. R. (2019). Representation learning of genomic sequence motifs with convolutional neural networks. *PLOS Computational Biology*, 15(12):1–17.

Lambert, M., Jambon, S., Depauw, S., and David-Cordonnier, M.-H. (2018). Targeting transcription factors for cancer treatment. *Molecules*, 23(6).

Lee, H., Ozbulak, U., Park, H., Depuydt, S., De Neve, W., and Vankerschaver, J. (2024). Assessing the reliability of point mutation as data augmentation for deep learning with genomic data. *BMC Bioinformatics*, 25(1):170.

Loshchilov, I. and Hutter, F. (2017). SGDR: Stochastic gradient descent with warm restarts. In *International Conference on Learning Representations*.

Pap., G. and Megyeri., I. (2022). Translational robustness of neural networks trained for transcription factor binding site classification. In *Proceedings of the 14th International Conference on Agents and Artificial Intelligence - Volume 3: ICAART*, pages 39–45. INSTICC, SciTePress.

Park, S., Koh, Y., Jeon, H., Kim, H., Yeo, Y., and Kang, J. (2020). Enhancing the interpretability of transcription factor binding site prediction using attention mechanism. *Scientific Reports*, 10(1):13413.

Qin, Q. and Feng, J. (2017). Imputation for transcription factor binding predictions based on deep learning. *PLOS Computational Biology*, 13(2):1–20.

Schreiber, J., Singh, R., Bilmes, J., and Noble, W. S. (2020). A pitfall for machine learning methods aiming to predict across cell types. *Genome Biology*, 21(1):282.

Shen, L.-C., Liu, Y., Song, J., and Yu, D.-J. (2021). SAResNet: self-attention residual network for predicting DNA-protein binding. *Briefings in Bioinformatics*, 22(5):bbab101.

Smith, L. N. (2017). Cyclical learning rates for training neural networks. In *2017 IEEE Winter Conference on Applications of Computer Vision (WACV)*, pages 464–472.

Vishnoi, K., Viswakarma, N., Rana, A., and Rana, B. (2020). Transcription factors in cancer development and therapy. *Cancers (Basel)*, 12(8).

Wang, Z., Xiong, S., Yu, Y., Zhou, J., and Zhang, Y. (2023). HAMPLE: deciphering TF-DNA binding mechanism in different cellular environments by characterizing higher-order nucleotide dependency. *Bioinformatics*, 39(5):btad299.

Wu, X., Hou, W., Zhao, Z., Huang, L., Sheng, N., Yang, Q., Zhang, S., and Wang, Y. (2024). Mmgat: a graph attention network framework for atac-seq motifs finding. *BMC Bioinformatics*, 25(1):158.

Zagoruyko, S. and Komodakis, N. (2016). Wide residual networks. In Wilson, R. C., Hancock, E. R., and Smith, W. A. P., editors, *Proceedings of the British Machine Vision Conference 2016, BMVC 2016, York, UK, September 19-22, 2016*. BMVA Press.

Zeng, H., Edwards, M., Liu, G., and Gifford, D. (2016). Convolutional neural network architectures for predicting dna–protein binding. *Bioinformatics*, 32:i121–i127.

Zhou, J. and Troyanskaya, O. G. (2015). Predicting effects of noncoding variants with deep learning–based sequence model. *Nature Methods*, 12(10):931–934.