# Development of Alginate-based Antibacterial Edible Films by Incoporating Green Betel Leaf Extract

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Keywords: Antibacterial, Edible Film, Green Betel Leaf Extract, Staphylococcus aureus, Escherichia coli.

Abstract: The aim of this research was to develop an environmentally friendly packaging of alginate-based antibacterial edible films. The antibacterial sources used was green betel leaf extract. Concentration levels of green betel leaf extract which was studied were 0,5%, 1,0% and 1,5%. The edible film obtained was evaluated physically, mechanically, chemically and microbiologically including parameters of Water Vapor Transmission Rate (WVTR), water solubility, tensile strength, elongation, brightness, thickness, moisture content, and antibacterial clear zone test. Experiment was conducted two replicates. The results showed that concentration level of green betel leaf extract affected on the quality and antibacterial properties of alginate-based edible films. Overall, the best alginate-based antibacterial edible film is produced by incorporating green betel leaf extract at concentration level of 1,5%.

# **1 INTRODUCTION**

Edible films are types of biodegradable plastic that attract scientists around the world to explore. Edible films have been employed to pack a variety of food products, such as meat, sausages, fresh fruits and vegetables. Edible films can maintain the quality of packaged food because edible film can withstand carbon dioxide, oxygen diffusion, water evaporation, and flavor contamination with other products. Another benefit of edible films is that it can extend the shelf life and is environmentally friendly. Edible films can be consumed together with the product. (Junianto et al. 2012). The main components of edible film can be grouped into three categories, namely hydrocolloids, lipids, and composites. Composites are a combination of hydrocolloids and lipids. The hydrocolloids that can be used to make edible films are proteins and carbohydrates. Meanwhile, the lipids used are wax and fatty acids (Donhowe and Fema, 2994). Alginate is one of hydrocolloids polysaccharides which are available in nature derived from seaweed and can be applied as bio polymeric film. Film forming and binding ability of alginate makes it suitable for the use of packaging material (Parveen et al, 2019).

Production of edible films from alginate have been explored by Murdinah *et al.* (2007), Khairunnisa *et al.* (2018), Koushki *et al.* (2015) and others. Alginate based edible films are found strong, but poor water resistance due to hydrophilic nature. One way to increase the performance of alginate based edible film is by developing it as an active packaging. The film can be used as a carrier of antioxidants, flavoring and/or coloring agents and antimicrobials to improve food safety and quality (Siracusa *et al*, 2018). Incorporating antimicrobial into alginate film can increase the resistance of the film to bacteria so that the film has good mechanical quality and can inhibit bacteria that potentially contaminate food.

One of the natural ingredients that contain antimicrobials is green betel leaf. The active antibacterial components found in green betel are tannins, flavonoids and essential oils (Baskaran *et al.*, 2011). These compounds are able to inhibit the growth of *Escherichia coli* and *Staphylococcus aureus*. These bacteria produce enterotoxins which can cause diarrhea. The antimicrobial activity of betel leaf active compounds is by denaturing proteins and damaging bacterial cell walls, as well as destructing lipids in cell membranes by reducing the surface tension of the cell membranes.

This study was aimed to develop an environmentally friendly packaging of alginate-based

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Giyatmi, G., Irianto, H., Sabariman, M. and Anggoro, B.

DOI: 10.5220/0010514000003108 In Proceedings of the 6th Food Ingredient Asia Conference (6th FiAC 2020) - Food Science, Nutrition and Health, pages 52-58 ISBN: 978-989-758-540-1

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antibacterial edible films by using green betel leaf extract as antimicrobial agent. added during the process. Meanwhile, physical, mechanical, chemical and antibacterial properties of the edible film were also observed.

### 2 MATERIALS AND METHODS

# 2.1 Material

Alginate flour used was extracted from *Sargassum* sp. by employing an extraction method developed by Yunizal (2000). The modified cassava (mocaf) flour used was the "Ladang Lima" brand, while sorbitol and beeswax were purchased from PT Geochem Globalindo, Jakarta. Green betel leaves were obtained from the Ciracas market, East Jakarta.

#### 2.2 Method

### 2.2.1 Extraction of Antibacterial Compounds from Green Betel Leaves

Antibacterial from green betel leaves were extracted through modifying a method previously employed by Sitorus (2018) for extracting antibacterial from betel leaves as well. After washing, the betel leaves were chopped so that they were exposed to more ethanol. The mixing was carried out in a large and closed container so that the maceration process of the material run perfectly. During mixing, the mixture was stirred periodically. After that, the soaking process was conducted for 3 days with stirring every 24 hours. Then the mixture was filtered with a 100 mesh filter cloth and the obtained filtrate was evaporated using a rotary vacuum evaporator until a thick extract was obtained (Sitorus, 2018).

#### 2.2.2 Preparation of Edible Film

Edible film was prepared according to the experiment conducted by Murdinah *et al* (2007) with modification. About 97 mL of distilled water in a beaker glass was heated to 50°C, then 1,5 g of alginate powder was gradually added while stirring with a magnetic stirrer. After that, 0,3 g of mocaf starch was added with stirring for 30 minutes. After mixing completely, the antibacterial green betel leaf extract was added into the solution according to the treatment (0,5%; 1,0%; 1,5%) and stirred for 30 minutes until evenly mixed at 75°C. Furthermore, the solution is casted by pouring into a square acrylic plate, where one edible film solution recipe can be casted on two acrylic plates. After that the acrylic plate was left to stand at room temperature for approximately 24 hours and then the dried edible film was removed.

#### 2.2.3 Experiment

Experiment was executed in two stages, i.e. preliminary and main experiments. Preliminary experiment was conducted to investigate antibacterial activity of green betel leaf extract to *S. aureus* and *E. coli*. The results of this preliminary experiment will be used in the main experiment. The main research was performed to determine the optimum addition level of green betel leaf extract as antibacterial agent on alginate-based edible film. The concentration level of betel leaf extract tested consisted of 3 levels (0,5%, 1,0%, 1,5%) with 2 replications.

#### 2.2.4 Analyses

Alginate based antibacterial edible films obtained were assessed their performance in terms of physical, mechanical, chemical and antibacterial properties including thickness using digital micrometer (Mitutoyo, Japan), tensile strength and elongation using Texture Analyzer (TAXT Plus, Stable Micro System, UK) (Balqis et al., 2017), water vapor transmission rate (WVTR) based on ASTM E96 using PERME® W3 / 031 Water Vapor Transmission Rate Tester (Labthink, China), brightness using Hunter Lab, water solubility (Murni et al, 2013), moisture content (Farhan and Hani, 2017), as well as antibacterial activity for S. aureus and E. coli (da Silva et al, 2013) and surface morphological analysis using SEM (Scanning Electron Microscopy) JEOL JSM-6360LA, Japan (Setiani et al, 2013).

# **3** RESULT AND DISCUSSION

### 3.1 Antibacterial Activity of Green Betel Leaf Extract

Antibacterial compounds extracted from green betel leaves using alcohol as the extracting solvent showed antibacterial activity on *S. aureus* and *E. coli* as shown in Table 1. The highest antibacterial activity of green betel extract against *S. aureus* based on the largest inhibition zone shown at the extract concentration of 1% was 14 mm, and the lowest activity exhibited by the extract concentration of 0,5% was 12,50 mm. While, the highest inhibition zone of green betel leaf extract against *E.* coli occurred at a concentration of 1,0% extract was 14 mm, and the lowest zone of inhibition shown at a concentration of 0,5% extract was 12,50 mm.

Table 1: Antibacterial activity of green betel leaf extract (Clear Zone, mm).

Microorganism	Concentration of Green Betel Leaf Extract		
Ū.	0,5%	1,0%	1,5%
Staphylococcus aureus	14,00	13,50	14,00
Escherichia coli	12,50	14,00	13,00

The antibacterial activity of betel leaf extract is suspected to be produced by the chemical compounds of the betel leaves, namely saponins, flavonoids, polyphenols, and essential oils. Saponin compounds can work as antimicrobials by damaging the cytoplasmic membrane and killing cells. Flavonoid compounds are thought to denaturate bacterial cell proteins and irreparably damage cell membranes. Natural phenols contained in essential oils have antiseptic properties 5 times stronger than ordinary phenols (bactericides and fungicides) but are not sporacid. The mechanism of phenol as an antibacterial agent acts as a toxin in the protoplasm, damages and penetrates the walls and precipitates bacterial cell proteins (Carolia and Noventi, 2016).

The main components of essential oil are betel phenol and some of its derivatives including eugenol allypyrocatechin 26,8 – 42,5%, cineol 2,4 – 4,8%, mehyl eugenol 4,2 – 15,8%, caryophyllen 3,0 -9,8%, kavikol hydroxy, kavikol 7,2-16,7%, kabivetol 2,7 6,2%, estragol, ilypryrocatekol 9,6%, carvacol 2,2 – 5,6%, alkaloids, flavonoids, triterpenoids or steroids, saponins, terpenes, phenylpropane, terpinen, diastase from 0,8 - 1,8%, and tannins 1,0 – 1,3% (Damayanti and Mulyono, 2003). Phenol at a concentration of 0,1 – 1,0% is bacteriostatic, while it is as bactericidal at a concentration of 1 - 2% (Fuadi, 2014),

### 3.2 Characteristics of Alginate-based Antibacterial Edible Film

The moisture content of the edible film increased with the increase in the concentration of antibacterial extract of green betel leaves in the film (Table 2). Suryaningrum *et al* (2005) noted that edible films being biodegradable with high moisture content will be easily grown by microorganisms, due to the presence of nutritional components in the film such as protein. On the otherhand, edible films with low moisture content will be more resistant to microbiological destruction. WVTR value of alginate edible film with different antibacterial concentration treatment resulted in different WVTR values with a tendency to decrease with increasing concentration. WVTR test results for edible film made with various concentrations of betel leaf extract yielded WVTR values ranging from 1640,65 to 1871,79 g/m<sup>2</sup>.24h. The lowest WVTR value of edible film was obtained at the addition of 1,5% antibacterial extract of betel leaves. Irianto *et al* (2006) reported that the lowest WVTR was resulted from a combination of 2,0% carrageenan, 0,7% tapioca and 0,3% beeswax, i.e. 746,2g/m<sup>2</sup>/day, which was lower than the value from this study.

The WVTR value in this study showed that the edible film with the addition of green betel leaf extract had a WVTR value which increased in line with the increase in the addition level of betel leaf extract. The higher the WVTR value, the higher the permeability of the edible film. When edible film is made into packaging, it will cause more moisture to come out and enter the package. A good edible film must not be easily passed by moisture or have a low WVTR. The compact edible film structure can inhibit water vapor diffusion through the edible film. Betel leaf extract contains oil and water. Oil components have high protective properties against water vapor which will reduce the hydrophilic properties of the film. Guilbert & Biquet (1996) informed that the fatty components such as wax, emulsifier and fatty acids in composite edible films have an effect on reducing WVTR because fat has low polarity and a dense crystal structure.

Table 2: Physical and Chemical properties of alginate based antibacterial edible film.

Parameter	Concentration of Green Betel Leaf Extract			
	0,5%	1,0%	1,5%	
Moisture Content (%)	11,09±0,40	11,53±0,04	12,42±0,10	
WVTR (g/m <sup>2</sup> .24h)	1871,79±68,19	1790,28±4,15	1640,65±57,05	
Water Solubility (%)	57,98±10,22	70,59±1,68	84,95±11,90	
Brightness (%)	14,2±1,27	12,1±0,27	11,5±1,70	
Thickness (mm)	0,114±0,014	0,104±0,013	0,108±0,008	

The water solubility value of alginate edible film with the addition of antibacterial extract of betel leaves at different concentrations resulted in film solubility which increased with increasing concentration of betel leaf extract. The more betel leaf extract is added, the more easily the edible film will dissolve. The lowest solubility value indicates that the edible film is the best because it plays a very important role when the film is used for edible products. This is also in accordance with the opinion of Stuchell and Krochta (1994) that if the application of a film is expected as an edible packaging, high solubility is desired. Likewise, if the application of edible film on foods with high water content, a film that is not water-soluble is used. Imeson (1999) adds that water resistance is an important property for films applied as food protection.

The addition level of green betel leaf extract affected the brightness of alginate edible film, in which increasing addition level of betel leaf extract resulted in edible film with lower brightness percentage values. Ningsih (2015) mentioned that the basic material used will affect the color of the edible film. Therefore, the edible film using different antibacterial concentrations will bring about different brightness of edible films.

The use of betel leaf extract with different concentrations in the production of alginate edible film did not significantly affect the thickness of the edible film, in which the thickness of edible film obtained in this study was in the range of 0,104 -0,114 mm. Those values are thicker compared to thickness values of edible film made of composite consisting of carrageenan, tapioca flour and beeswax, i.e. 0,050 - 0,074 mm (Irianto et al, 2006). Meanwhile, Saputro et al (2017) produced edible films with a thickness of 0.03 - 0.08 mm which was carrageenan, tapioca flour and glycerol used as raw materials. According to Wahyu (2009) that a good edible film has a thickness of 0,15 - 0,20 mm which can withstand external influences and reduce WVTR. Thickness is an important parameter that needs to be considered in the use of edible film as a packaging. The thickness of the film will affect the gas permeability. The thicker the edible film, the lower the gas permeability, which can protect the product.

The tensile strength values of alginate edible film were insignificantly influenced by the addition levels of green betel leaf extract. The higher addition of the betel leaf extract did not result in a pronounced difference in tensile strength of the edible film. The tensile strength of the edible film in this study was in the range of  $1,31\pm0,84 - 2,67\pm0,51$  MPa (Table 3). Those tensile strength values are not in the range of ASTM polypropylene standard values, i.e. 24,7-302 MPa. In addition, the best tensile strength of edible films is between 10 - 100 MPa while edible films with a tensile strength range of 1 - 10 MPa are classified as marginal (Han and Gennadios, 2005). The increase in the forces of attraction between the molecules making up the edible film induces an increase in the strength of its structure. The greater the tensile value, the stiffness of a material will increase or be inelastic. Sara (2015) noted that tensile strength is a mechanical property related to the strength of the edible film to withstand physical damage when applied as food packaging. Edible film with high tensile strength value is highly expected in order to minimize packaged product damage due to mechanical interferences.

Table 3: Mechanical properties of alginate based antibacterial edible film.

Parameter	Concentration of Green Betel Leaf Extract		
	0,5%	1,0%	1,5%
Tensile Strength (Mpa)	2,52±0,75	2,67±0,51	1,31±0,84
Elongation (%)	47,05±18,88	38,30±4,95	24,40±13,58

The elongation test results showed that the more the betel leaf extract on the alginate edible film, the lower the elongation value. The elongation percentage of the film is categorized as poor if the value is less than 10% and good if the elongation percentage exceeds 50%. Elongation percentage of alginate based edible film added with various concentrations of betel leaf extract in this study was in the range of 24,40±13,5 - 47,05±18,88 %. The elongation value of the edible film from this study was lower than the elongation value of edible film made of bligon goat skin gelatin, namely 70,97 -95,33% (Said et al., 2013). Edible films from whey and agar have comparable elongation, i.e. 26,06 -34,52% (Hakim, 2015). Elongation of edible films made from avocado seed starch and agar is smaller, i.e. 0,17-0,45% (Coniwanti et al, 2016).

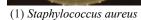
The antibacterial test showed that the alginatebased edible film incorporated with green betel leaf extract had antibacterial activity against S. aureus and E. coli (Table 4) as indicated by the formation of a clear zone on the test medium (Figure 1). The antibacterial activity was getting more significant with the higher concentration level of antibacterial betel leaf extract, both for S. aureus and E. coli which showed the same trend. Thus, if greater antibacterial activity is expected, the addition level of betel leaf extract must be increased. The development of antibacterial edible films based on various types of starch has also been carried out using several antibacterial sources from the extracts of beluntas leaves (Mulyadi et al, 2016), garlic (Anggraini et al, 2018), red galangal/Alpinia purpurata (Sholehah et al, 2016) and areca nut (Ningsih, 2018).

Parameter	Concentration of Green Betel Leaf Extract		
	0,5%	1,0%	1,5%
Staphylococcus aureus	14,25	17,85	18,15
Escherichia coli	12,40	14,00	13,40

Table 4: Antibacterial properties (Clear Zone, mm) of

alginate based edible film added with green betel leaf





extract.

(2) Escherichia coli

Figure 1: Clear zone as antibacterial effects of green betel leaf extract of alginate based edible film on *S. aureus* and *E coli*.



The molecular structure surface of the alginate edible film is dense at 500 times magnification.

The molecular structure surface of the alginate edible film is dense at 1000 times magnification.



The molecular structure surface of the alginate edible film is not dense at 2000 times magnification

Figure 2: SEM images of alginate edible film added with 1.5% betel leaf extract with three times magnification.

Surface morphological analysis using SEM was carried out on alginate based edible film added with green betel leaf extract at a concentration of 1.5% (Figure 2), generally showing the best physical, mechanical and chemical properties. Analyses revealed that the molecular structure surface of the alginate edible film at 500 and 1000 times magnification was dense. The higher the magnification used, the more obvious the surface morphology of the edible film, and it turned out that it showed the lack of molecular density in the resulting edible film. The less dense structure of these molecules brought about more water to be absorbed. The image also showed a less smooth and porous surface. The non-smooth surface indicated that the film was less homogeneous. This occurrence was probably due to a too short stirring period, so that it resulted in inhomogeneous edible film solution inducing a non-dense molecular structure surface of the alginate edible film. Therefore, further study to explore this phenomenon are highly recommended.

# **4** CONCLUSION

- 1) Green betel leaf extract has antibacterial activity against *S. aureus* and *E. coli*
- 2) Alginate-based edible film added with green betel leaf extract has relatively good physical and mechanical characteristics, especially in terms of thickness, tensile strength, elongation and WVTR.
- Edible film added with betel leaf extract as antibacterial showed antibacterial activity against *S. aureus* and *E, coli*; so it can be called as an antibacterial edible film
- Antibacterial edible film is recommended for performance testing by being used as a packaging for food products

# ACKNOWLEDGEMENT

Authors would like to thank to the Ministry of Research, Technology and Higher Education of the Republic of Indonesia, who has provided funding for executing this study through PTUPT program 2019 No: 45.3/USJ-11/H.54/2019.

# **AUTHORS' CONTRIBUTIONS**

Giyatmi Giyatmi and Hari Eko Irianto are the main contributors in conducting experiment and preparing the draft of manuscript, while Mohammad Sabariman and Bintang Anggoro are as supporting contributors. All authors read and approved the final manuscript.

### REFERENCES

- Anggraini, T.N., Agustini, T.W. and Rianingsih, L. 2018. Karakteristik Edible Film Karaginan Dengan Penambahan Ekstrak Bawang Putih (*Allium sativum*) Sebagai Antibakteri. Saintek Perikanan 14 (1) : 70-76
- Baskaran, C., Rathabai, V., & Kanimozhi, D., 2011. Screening for antimicrobial activity and phytochemical analysis of various leaf extract of *Murraya koenigi*. *Int J. Res Ayurveda Pharm*, 2, 1807-10
- Balqis, A. I., Khaizura, M. N., Russly, A. R., and Hanani, Z. N., 2017. Effects of plasticizers on the physicochemical properties of kappa-carrageenan films extracted from *Eucheuma cottonii*. International Journal of Biological Macromolecules, 103, 721-732. https://doi.org/ 10.1016/j.ijbiomac.2017.05.105
- Carolia, N. and Noventi, W., 2016. Potensi Ekstrak Daun Sirih Hijau (*Piper betle* L.) sebagai Alternatif Terapi Acne vulgaris. Majority 5 (1): 140 – 145
- Coniwanti, P., Sari, D.M., and Ferbriana, R., 2016. Pengaruh Rasio Massa Pati Biji Alpukat dan Agar -Agar Terhadap Pembuatan Edible Film. Jurnal Teknik Kimia. 22(2): 51-59
- Damayanti R. and Mulyono, 2003. Khasiat & manfaat daun sirih: obat mujarab dari masa ke masa. Agromedia Pustaka. Jakarta.
- da Silva, M., Lamanaka, B., Taniwaki, M. and Kieckbusch, T. 2013. Evaluation of the Antimicrobial Potential of Alginate and Alginate/ Chitosan Films Containing Potassium Sorbate and Natamycin. Packaging Technology and Science 26 (8): 479-492
- Donhowe, G. and Fennema, O., 1994. Edible film and coating: Characteristic, formation, definitions and testing methods. In Krochta, J.M., Baldwin, E.A. and Nisperos-Carriedo, M.O. (eds.). Edible Coating and Film to Improve Food Quality. Technomic Publ. Co. Inc. Lancaster, Pennsylvania. 378 pp
- Farhan, A. & Hani, N. M., 2017. Characterization of edible packaging films based on semi-refined kappa carrageenan plasticized with glycerol and sorbitol. Food Hydrocolloids, 64, 48-58. https://doi.org/ 10.1016/j.foodhyd.2016.10.034
- Fuadi S., 2014. Efektivitas ekstrak daun sirih hijau (*Piper betle* L.) terhadap pertumbuhan bakteri *Streptococcus pyogenes* in vitro. Skripsi. Universitas Islam Negeri Syarif Hidayatullah. Jakarta.
- Guilbert, S. and Biquet, B., 1990. Edible Films and Coatings. *In* Food Packaging Technology Vol. 1. (Eds. Bureau, G. and Multon, J.L.). VCH Publisher, Inc. New York.
- Hakim, M.Q., 2015. Karakteristik Edible Film dari Whey Dangke yang Ditambahkan Level Agar yang Berbeda. Skripsi. Fakultas Peternakan. Universitas Hasanuddin. Makassar
- Han, J.H. and Gennadios, A., 2005. Edible Film and Coatings: A Review. Innovations in Food Packaging. Academic Press.
- Irianto, H. E., Darmawan, M. and Mindarwati, E., 2006. Pembuatan Edible Film dari Komposit Karaginan, Tepung Tapioka dan Lilin Lebah (*Beeswax*). J.

Pascapanen Dan Bioteknologi Kelautan Dan Perikanan 1 (2): 93-101.

- Imeson, A., 1999. Thickening and Gelling Agents for Food. Aspen Publishers Inc. Maryland.
- Junianto., N., Kurniawati, O.S., Djunaidi, and Khan, A.M.A., 2012. Physical and Mechanical Study on Tilapia's Skin Gelatine Edible Films with Addition of Plasticizer Sorbitol. *African Journal of Food Science* 6(5):142-146.
- Khairunnisa, S., Junianto, Zahidah, and Rostini, I., 2018. The effect of glycerol concentration as a plasticizer on edible films made from alginate towards its physical characteristic. World Scientific News 112: 130-141
- Koushki, M.R., Azizi, M.H., Azizkhani, M., and Koohy-Kamaly, P., 2015. Effect of Different Formulations on Mechanical and Physical Properties of Calcium Alginate Edible Films. Journal of Food Quality and Hazards Control 2: 45-50
- Mulyadi, A.F., Maimunah Hindun Pulungan, M.H. and Qayyum, N., 2016. Pembuatan Edible Film Maizena dan Uji Aktifitas Antibakteri. Industria: Jurnal Teknologi dan Manajemen Agroindustri 5(3): 149-158
- Murdinah, Darmawan, M. and Fransiska, D., 2007. Karakteristik Edible Film dari Komposit Alginat, Gluten dan Lilin Lebah (*Beeswax*). Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan 2(1): 19-25
- Murni, S. W., Pawignyo, H., Widyawati, D., & Sari, N., 2013. Pembuatan Edible Film dari Tepung Jagung (*Zea mays* L.) dan Kitosan. Prosiding Seminar Nasional Teknik Kimia Kejuangan (p. B17 1-9).
- Ningsih, W., 2018. Formulasi Dan Uji Efektivitas Antibakteri Edible Film Ekstrak Biji Pinang (*Areca catechu* Linn). Jurnal Ilmu Farmasi dan Farmasi Klinik (JIFFK) 15 (2): 71-76
- Parveen, I., Maraz, K.M., Mahmud, M. and Khan, R.A., 2019. Seaweed Based Bio Polymeric Film and Their Application: A Review on Hydrocolloid Polysaccharides. Scientific Review 5 (5): 93-102
- Said, M.I., Triatmojo, S., Erwanto, Y., and Fudholi, A., 2013. Evaluasi Karakteristik Fisik Edible Film dari Gelatin Kulit Kambing Bligon yang Menggunakan Gliserol Sebagai Plasticizer. Jurnal Ilmu dan Teknologi Hasil Ternak. 8(2): 32
- Saputro, B.W., Eko Nurcahya Dewi, E.N. and Susanto, E., 2017. Karakteristik Edible Film Dari Campuran Tepung Semirefined Karaginan Dengan Penambahan Tepung Tapioka Dan Gliserol. J. Peng. & Biotek. Hasil Pi. 6 (2): 1-6
- Sara, N.E.M., 2015. Karakteristik edible film berbahan dasar whey dangke dan agar dengan penambahan konsentrasi sorbitol. Skripsi. Universitas Hasanuddin. Makassar
- Setiani, W., Sudiarti, T. and Rahmidar, L. 2013. Preparasi Dan Karakterisasi Edible Film Dari Poliblend Pati Sukun-Kitosan. Valensi 3 (2): 100 - 109
- Sholehah, M.M., rid Ma'ruf, W.F. and Romadhon., 2016. Karakteristik Dan Aktivitas Antibakteri Edible Film Dari Refined Carageenan Dengan Penambahan Minyak Atsiri Lengkuas Merah (*Alpinia purpurata*). J. Peng. & Biotek. Hasil Pi. 5 (3): 1-8

6th FiAC 2020 - The Food Ingredient Asia Conference (FiAC)

- Siracusa, V., Romani, S., Gigli, M., Mannozzi, C., Cecchini, J.P., Tylewicz, U. and Lotti, N., 2018. Characterization of Active Edible Films based on Citral Essential Oil, Alginate and Pectin. Materials 11(10): 1980
- Sitorus, P. 2018., Uji Efek Kombinasi Amoksisilin Dengan Ekstrak Etanol Daun Sirih (*Piper betle* L) Terhadap Pertumbuhan Bakteri Escherichia Coli Dan Staphylococcus Aureus. TM Conference Series 01: 313–319
- Stuchell, Y. M. and J. M. Krochta., 1994. Enzymatic treatments and thermal effects on edible soy protein films. J. Food Sci. 59(6): 1332-1337.
- Suryaningrum, Th. D., Basmal, J. and Nurrochmawati, 2005. Studi pembuatan Edibel Film dari Karagenan. Jurnal Penelitian Perikanan Indonesia. Edisi Pasca Panen. Badan Riset Perikanan dan Kelautan Departemen Kelautan dan Perikanan 2(4): 1 – 13.
- Wahyu, M. K., 2009. Pemanfaatan Pati Singkong sebagai Bahan Baku Edibel film. Jurusan Teknologi Industri Pangan, Fakultas Teknologi Industri Pertanian, Universitas Padjajaran. Bandung
- Yunizal, 2000. Penelitian Teknologi Ekstraksi Alginat dari Rumput Laut Coklat (*Phaeophyceae*). Balai Penelitian Perikanan Laut. Jakarta. 100 pp.