Isolation and Characterization of an Antioxidant Compound from Kayu Hitam Leaves (*Diospyros celebica* Bakh.F.)

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Keywords: Kayu Hitam (Diospyros celebica Bakh.F.), Isolation, Characterization, Methyl Gallate, Antioxidant.

Abstract: Isolation and characterization of an antioxidant compound from kayu hitam leaves (*Diospyros celebica* Bakh.F.) had been done by extraction and column chromatography method. Kayu hitam leaves powder was extracted with methanol and methanol extract reextracted with aquadest. Aqudest extract was partitioned with ethyl acetate and ethyl acetate extract repartitioned with n-hexane. The residues which are phenolic compounds were separated by column chromatography (SiO2, chloroform: methanol 90:10, 80:20, 70: 30,60: 40). The isolate obtained was purified with a preparative thin layer chromatography and obtained 9.5 mg of pure isolate in the form of yellow solid. characterization of pure isolate was determined by UV-Vis, FT-IR and 1H-NMR spectroscopic analysis. Based on the analysis carried out it can be characterized that the pure isolate obtained is methyl gallate. The antioxidant activity of methyl gallate was determined based on the DPPH free radical scavenging method. The activity of the methyl gallate was classified as strong with IC50 value of 4.41 μg / mL.

1 INTRODUCTION

Kayu hitam (*Diospyros celebica* Bakh.F.), classified as luxury wood species. Other names of kayu hitam in Indonesia including eboni, toetandu, sora, kayu lotong, kayu maitong, etc. (Prajadinata et al, 2011). Kayu hitam is endemic to Indonesia that distributes from Northern Sulawesi and Central Sulawesi to Southern (Larekeng, 2016). It is durable and strong wood, the heartwood with black and reddish brown stripes makes the texture very beautiful and widely used for luxury furniture, sculpture, carving, fan, statues, decorative tools, fancy veneer, musical instruments and ornaments (Prajadinata et al, 2011).

Sawdust from the processing of kayu hitam can function as a fungicide. At a concentration of 5% sawdust ethanol extract can cause a clear zone of 11 mm against the growth of Phytophthora palmivora Butler (Alwi et al., 2010) and Minimum Bactericidal Concentration (MBC) value of *S. aureus* and *E. coli* were 12% and 13% respectively (Wahyuni et al., 2018). This is due to, the sawdust extract contain chemical compounds such as tannins, saponins and terpenoids (Wahyuni et al., 2018). Ethanol extract of kayu hitam also had acute toxicity with LD50 value of 5.168 mg / kg against male mice (Mus musculus) (Syam, 2016. The toxicity of a plant depends on various factors, including quanti-consumed, time of exposure, different parts of the plant, individual chemistry, climate and soil, and genetic, species differences and strength of secondary metabolites (Mounanga et al, 2015).

Secondary metabolites are products of metabolism found in plants. Secondary metabolite compounds are divided into several parts, including phenolic compounds (Cheynier et al, 2013). Phenolics are characterized by having at least one aromatic ring with one or more hydroxyl groups attached (Crozier, et al., 2006). Phenolics are important components of the human diet due to their potential antioxidant activity and their capacity to diminish oxidative stress induced tissue damage resulted from chronic diseases (Khadem and Marles, 2010).

Antioxidants are compounds that neutralize chemically active products of metabolism, such as free radicals which damage the body. Sources of natural antioxidants are primarily phenolics that may occur in all products and parts of a plant such as fruits, vegetables, nuts, seeds, leaves, roots, and bark (Hajaji et al., 2010) and also in woody plants such as Toona sureni (Ekaprasada, et al., 2009 and in the Archidendron jiringa plants (Lubis, et al., 2018).

234

Br. Sembiring, H. and Romasni Purba, Y.

Isolation and Characterization of an Antioxidant Compound from Kayu Hitam Leaves (Diospyros celebica Bakh.F.). DOI: 10.5220/0008919802340238

In Proceedings of the 1st International Conference on Chemical Science and Technology Innovation (ICOCSTI 2019), pages 234-238 ISBN: 978-989-758-415-2

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Herein we report the isolation characterization of an antioxidant compound obtained from kayu hitam leaves and its antioxidant activity. Chemical structure was determined based on spectroscophy data interpretation and antioxidant activity based on scavenging activity of DPPH(1,1-diphenyl-2picrylhydrazil) radical method and ascorbic acid was used as positive control. Isolation and characterization of an antioxidant compound obtained from kayu hitam leaves never been reported.

2 MATERIALS AND METHODS

2.1 Materials

Kayu hitam leaves were collected from the front yard of the Universitas Sumatera Utara, Medan, Sumatera Utara, Indonesia. Identification of plant was done at Herbarium Medanense (MEDA) Universitas Sumatera Utara. Silica (70 - 230 mesh, E-merck) for column chromatography, FeCl₃ 5%, chloroform (p.a E Merck), silica 60 F254 (E.Merck) for thin layer chromatography, TLC Preparative 60 F254, Benzene (p.a E Merck), Acetone (p.a Merck) methanol (p.a E Merck) and DPPH (Sigma Aldrich). Methanol as solvent was distilled before used.

2.2 Instrument

The ¹H-NMR spectrum was recorded on a Agilent 2NMR 500MHz spectrometer instrument with CD3OD as a solvent and TMS as an internal standard and chemical shifts are given in δ (ppm). IR spectrum were recorded on FT-IR (Shimadzu), UV spectrum were recorded on Spectrophotometer UV-Vis (Hewlett Packard Agilent), solvent evaporation with rotary evaporator (Heidolph), monitoring sample spots with UV lights (254nm / 356nm, UVGL 58) and measuring antioxidant activity with a UV-Vis spectrophotometer (SP-300).

2.3 Procedure

2.3.1 Extraction and Isolation

This extraction and isolation were done based on Megawati, et al (2015) with a slight modification. The leaves powder of kayu hitam (1800 g) was macerated with 8L methanol for 2 x 24 hours. The macerate is collected, concentrated with a rotary evaporator and dried on a water bath. Methanol extract (209.63g) was dissolved with aquadest, the filtrate obtained were reextracted using ethyl acetate. The solvent in the ethyl acetate fraction is evaporated to obtained Ethyl acetate extract. Ethyl acetate extract (32.25 g) was dissolved with methanol and reextracted by using nhexane. The methanol layer was dried using a rotary evaporator so that the dry methanol extract (12 g) was obtained. The phenolic compounds in the methanol extract were separated by using column chromatography using chloroform: methanol (100:0; 90:10, 80:20, 70:30, 60:40 (%v/%v). Isolates were collected in the vial every 10 mL and analyzed by TLC using chloroform: methanol 90:30. Each fraction with the same Rf value was c combined and evaporated. Fraction 38-92 (100 mg) at Rf 0.29 was purified by preparative TLC (Hostettmann et al., 1995) with chloroform: ethyl acetate 50:50 (% v /% v) and produced one band spot at the Rf 0.45. The band spot was crushed, eluted and tested with 5% FeCl₃, evaporated to obtain pure isolates 9.5 g in the form of yellow solid. The pure isolate was identification by UV-Vis, FT-IR and ¹H-NMR analysis and antioxidant activity test.

2.3.2 Antioxidant Activity Test

Use Antioxidant activity test for pure isolate from kayu hitam leaves was done based on free radical scavenging method using DPPH (1,1-diphenyl-2-pikrylhydrazil) developed by Molyneux (2004) and Saranya et al., (2017). Samples and ascorbic acid were dissolved in methanol (p.a E Merck) with concentrations of 0.5, 10, 15 and 20 μ g/mL. The inhibition percentage can be determined using equation formula (1) as follow:

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inhibition percentage = \frac{blank \ absorbance-sample \ absorbance}{blank \ absorbance} x100\%
(1)
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3 RESULTS AND DISCUSSION

3.1 Isolation and Characterization

Kayu hitam (Figure 1A) used in this study was the Ebenaceae family, a species of *Diospyros celebica* Bakh. F. with the local name kayu hitam. Pure isolate was isolated from kayu hitam leaves (Figure 1B) was phenolic compound, this was evidenced by the formation of black colloid on the addition of FeCl3 5%. The pure isolate is a yellow solid (Figure 1C). Identification of phenolic compounds was determined by UV-Vis, FT-IR and ¹H-NMR spectroscopic analysis.

The UV-Visible (CH3OH) spectrum λ_{max} 290 nm which is the length of the gallic acid group (Sujata, 2005) is shown in Figure 2.



Figure 1: A Kayu hitam plant, 1B Kayu hitam leaf, 1C Pure isolate.

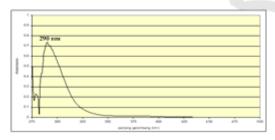


Figure 2: Spectrum UV-Visible of pure isolate.

This can be supported by the calculation of the wavelength for UV-Visibel in theory. Main Chromophore (246 nm), m-OH (2 X 7 nm = 14 nm), p-OH (25 nm), so that it is obtained λ_{max} 285 nm. Based on the calculation results λ_{max} pure isolate corresponds to λ_{max} comparative compound that is gallic acid (Pavia, 2001).

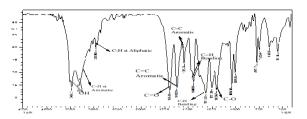


Figure 3: Spectrum FT-IR of pure isolate.

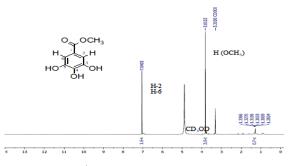


Figure 4: ¹H NMR spectrum of pure isolate.

FT-IR spectrum of pure isolated was shown in Figure 3. The FT-IR spectrum for pure isolates showed (KBr, v max, cm⁻¹) 3468.01 (O-H), 3311.78 (C-H), 2955.02 (C=H), 1618.29 (C = O), 1313.52 (C-H), 1251.80 (C-O), indicated that the pure isolate has a group commonly found in phenolic compound (Andersen and Markham, 2006). ¹H NMR spectrum of pure isolate shown in Figure 4. Based on ¹H NMR spectrum (Methanol-D6, 500 MHz, (ppm)) δ 7.04 $(2H, s, H-2, H-6), \delta 3.81 (3H, s, OCH_3)$, indicated that pure isolate had two aromatic protons and three methyl protons. The data in FT-IR and ¹H NMR spectrum are similar to FT-IR and ¹H NMR data reported by Ekaprasada, et al. (2009). Based on data analysis and interpretation carried out on the UV-Visible, FT-IR and ¹H-NMR spectrum and comparative

Spectrum reported by Hisham, et al. (2011) it was stated that pure isolates obtained from the leaves of kayu hitam plant was simple phenolic compound, methyl gallate with the structure shown in Figure 5.

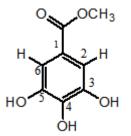


Figure 5: Structure of Methyl Gallate.

3.2 Antioxidant Activity of Pure Isolate

Table 1 showed the percentage of inhibition and IC50 values of methyl gallate and ascorbic acid as positive control.

Table 1: IC50 of ascorbic acid and methyl gallate.

Sample	Concentration	Inhibition (%)	IC ₅₀ (ug/mL)
Methyl gallate	0	0	
	5	84.77	
	10	92.38	4.41
	15	95.43	
	20	97.71	
Ascorbic acid	0	0	4.09
	5	85.53	
	10	92.38	
	15	96.19	
	20	97.72	

IC50 value of methyl gallate had no significantly different with ascorbic acid. It showed that methyl gallate has proton donating ability and could scavenge the free radical of DPPH.

4 CONCLUSIONS

Pure compound had been isolated from kayu hitam leaves. Based on the data spectrum UV Vis, FT-IR and 1H NMR the pure compound was methyl gallate. Methyl gallate is an antioxidant compound with IC50 value $4.41 \mu g/mL$.

ACKNOLEDGEMENTS

Each We would like to thank to Herbarium medananse (MEDA), Laboratory of Natural Sciences Chemistry Faculty of mathematical and Science and Laboratory of Research, Faculty of Pharmacy University of Sumatera Utara for identification of sample, isolation and absorbance measurements in determining antioxidant activity of pure isolate. We would also like to thank to Lanang solakhudin and Elvira Hermawati for the analysis of Spectrophotometer UV Visible, FT IR and ¹H-NMR, Laboratory of Organic Chemistry, ITB Bandung.

REFERENCES

- Alwi, M., Ramadanil dan Puspa, D.N., 2010. Ekstrak Serbuk Gergaji Kayu Eboni (Diospyros celebica Bakh.) Sebagai Fungisida Terhadap Phytophthora palmivora Butler. Jurnal Biocelebes. 4 (2),89-97. (Indonesian).
- Andersen, Q.M. and Markham, K.M., 2006. Flavonoids Chemistry, Biochemistry and Applications. Taylor and Francis Group, LLC. CRC Press.
- Chatterjee, S., Zareena Niaz, S. Gautam, Soumyakanti Adhikari, Prased S. Variyar and Arun Sharma, 2007. Antioxidant Activity of Some Phenolic Constituents from Green Pepper (Piper ningrum L.) and Fresh Nutmeg Mace (Myristica fragrans), Food Chemistry. 101, 515-523.
- Crozier, A., Clifford, M.N. and Ashihara, H., 2006. Plant Secondary Metabolites: Occurrence, Structure and Role in the Human Diet. Blackwell Publishing Ltd. First Published. Oxford. USA.
- Ekaprasada, M.T., Nurdin, H., Ibrahim, S. and Dachriyanus., 2009. Antioxidant Activity of Methyl Gallate Isolated from The leaves of Toona sureni. Indo. J. Chem. 9 (3), 457 - 460 457.
- Lubis, M.Y., Siburian, R., Marpaung,L. Simanjuntak, P. and Nasution, M.P., 2018. Methyl Gallate From Jiringa (Archidendron jiringa) and Antioxidant Activity. Asian J Pharm Clin Res. 11 (1), 346-350.
- Khadem, S. and Marles, R.J., 2010. Review: Monocyclic Phenolic Acids; Hydroxy- and Polyhydroxybenzoic
- Acids: Occurrence and Recent Bioactivity Studies. Molecules. 15, 7985-8005.
- Megawati, Saepudin, E., Hanafi, M., Darmawan, A. Lotulung, P.D.N., 2015. Identification and Bioactivity Studies of Flavonoid Compounds from Macaranga hispida (Blume) Mull. Arg. Makara J. Sci. 19(3), 96-100
- Molyneux, P., 2004. The Use of the Stable Free Radical Diphenylpicrylhydrazyl (DPPH) for Estimating Antioxidant Activity, Songklanakarin J. Sci. Technol, 2004, 26 (2), 211-219.
- Prajadinata, S., Effendi, R. and Murniati., 2011. Review of Management and Conservation Status of Ulin (Eusideroxylon zwageri Teijsm & Binn.), Ebony (Diospyros celebica Bakh.) and Cempaka (Michelia champaca Linn.) in Indonesia. Itto Project Pd 539/09 Rev.1 (F) in Cooperation with Center for Conservation and Rehabilitation Research and Development, Forestry Research and Development Agency Ministry of forestry Bogor – Indonesia
- Hajaji, H.E., Lachkar, N., Alaoui, K., Cherrah, Y., Farah, A., Ennabili, A. Bali, B.E and Lachkar, M., 2010. Antioxidant Properties and Total Phenolic Content of Three Varieties of Carob Tree Leaves from Morocco. Records of Natural Products. 4(4), 193-204.
- Larekeng, S.H., 2016. Polymorphism of Simple Sequence Repeat Regions of Sulawesi Ebony (*Diosphyros* celebica Bakh.) in Experimental Forest of Hasanuddin University Provenance. Agrotech Journal. 1 (1), 37-44
- Mounanga, M.B., Mewono, L. and Angone, S.A., 2015. Toxicity studies of medicinal plants used in sub-

Saharan Africa. Journal of Ethnopharmacology. 174, 618–627.

- Saranya D, Sekar J, Adaikala RG. Assessment of antioxidant activities, phenol and flavonoid contents of different extracts of leaves, bark and root from the *Abutilon indicum* (L.) sweet. *Asian J Pharm Clin Res* 2017; 10:88-94.
- Sembiring, H.B., Barus, T., Marpaung, L. Simanjuntak, P., 2015. Antioxidant and Antibacterial Activity of Some Leaves Extracts (Methanol, Ethyl Acetate and N-Hexane) of *Scurrula fusca* G. Don. International Journal of Pharm Tech Research.8(9), 24-30.
- Wahyuni, Ibrahim, N. and Nugrahani, A.W., 2018. Uji Aktivitas Antibakteri Ekstrak Serbuk Gergaji Kayu Eboni (Diospyros celebica Bakh.) Terhadap Bakteri Staphylococcus aureus dan Escherichia coli. Biocelebes. 12 (1), 54-64.
- Hisham, MN., Lip MJ., Noh MJ., Normah A., Nabila MF., 2011. Identification and Isolation Of Methyl Galate As A Polar Chemical Marker For Lobisia Pumila Benth. J. Trop. Agric. and Fd. Sc. 39(2), 279-284.
- Hostettmann, K., Hostettmann, M., Marston, A., 1995. Cara Kromatografi Preparatif, Penggunaan Pada Senyawa Bahan Alam. Penerbit ITB. Bandung
- Pavia, D.L., Lampman, G.M., Kriz, G.S., 2009. Introduction to Spectroscopy: A Guide for Students of Organic Chemistry. Saunders College. Philadelphi
- Syam,A,K, (2016). Uji Toksititas Akut Ekstak Etanol Daun Kayu Hitam (Diospyros celebica b.) Terhadap mencit (mus musculus). Fakultas Kedokteran dan Ilmu Kesehatan Universitas Islam Negeri Alauddin Makassar Samata-Gowa (Skripsi).