

A COMPUTATIONAL STRATEGY TO INVESTIGATE RELEVANT SIMILARITIES BETWEEN VIRUS AND HUMAN PROTEINS

Local High Similarities between Herpes and Human Proteins

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Abstract: Investigating primary sequence and structural features of viral proteins/genes has revealed molecular mimicry and evolutionary relationship linking viruses to eukaryotes. The continuous improvement in sequencing-techniques makes available almost daily the whole genome/proteome of several microorganisms, making now possible systematic analyses of evolutionary correlations and accurate phylogeny investigations. In the present study we set up a methodology to identify significant and relevant similarities between viral and human proteomes. To this aim, the following steps were applied: i) identification of local similarity corresponding to continuous identity over at least 8-residues long fragments; ii) filtering results for statistical significance of the identified similarities, according to BLAST parameters for short sequences; iii) additional filters applied to the BLAST outputs, to select specific viruses. The present study indicates a novel accurate methodology to find relevant similarities among virus and human proteomes, useful to further investigate pathogenic mechanisms underlying infectious and non-infectious diseases.

1 INTRODUCTION

Several studies investigate genetic predisposition toward human disorders (Baranzini, 2009; Rubstov,

2010). Mechanisms involving a genetic basis may require events occurring at the germinal level (giving hereditary disorders) or at somatic levels. Environmental stimuli and life style, such as smoke

habit, professional chemical exposure, diet style or exposure to pollution or to infectious agents, may interfere at the genetic level on the system homeostasis within the human body. In several cases infective agents, besides the direct infection disease, may trigger additional mechanisms responsible of the onset of additional diseases. Virus infections are often responsible of additional serious pathologies, and in most cases the mechanisms underlying the onset of these “secondary” pathologies are not fully elucidated. For instance, Human Papilloma virus (HPV) infection is known to strongly relate to the occurrence of cervix cancer in humans (Leggatt, 2007); helicobacter pylori infection is strongly related to gastric ulcer and gastric cancer (Cao, 2007); hepatitis B virus (HBV) infection has been proposed to have a role in the development of hepatocellular carcinoma (Neuveut, 2010); diabetes has been related with rotavirus infection (Maklea, 2006) or coxsackie virus infection (Peng, 2006); bacterial infections have been shown to be related to heart diseases (Ott, 2006); myocarditis has been proposed to have an autoimmune basis related to auto-antibodies against cardiac myosin and based on a mimicry process between cardiac myosin and the beta-adrenergic receptor (Li, 2006); Herpes Virus type 7 has been shown to be involved in initiation and maintenance of Graves’ disease autoimmune process (Leite, 2010); Herpes Virus type 1 infection has been strongly related to autoimmune reaction based on the auto-reactive T lymphocytes recognizing shared epitopes (Zhao, 1998); herpes antigens have been suggested to play a molecular mimicry role for autoimmune-based diseases such as psoriasis (Kirby, 2000; Mehraein, 2004) and antibodies against Herpes viruses have been found in immuno-deficient or auto-immune patients (Thomas, 2008). A recent study (Fumagalli, 2010) pointed out that up to 8% of the human genome is of viral source, representing the “fossil remnants of past infections” via integration at several sites. Consistent with these findings, a strong rational supports the investigation of human-to-viral structural-functional relationships based, at least in some cases, onto a common-antigen sharing process. We have previously suggested unexpected evolutionary relations between human-lymphocytes CD4 receptor and his counterpart HIV-capsid-constituent GP120 (Facchiano, 1995; Facchiano, 1996); allele variability has been related to viral infection and susceptibility, with a significant association of HIV progression with patients HLA status (Limou, 2009; Fellay, 2007). Based on significant proteins similarities we have also suggested the occurrence of

molecular mimicry between pathogens and human proteins possibly underlying different diseases (Benvenga, 1995; Benvenga, 1999; Benvenga, 2003). According to such large body of evidence, it is now accepted that eukaryotes have been, and still are, under a strong selective pressure, driven, at least in part, by viral/prokaryotes infections, with human gene variants associated to pathogens-related infections. The strong improvement of the sequencing techniques is producing a continuous novel identification of the entire genome/proteome of several micro-organisms, including viral pathogens. Such sequences, or at least those available freely on the net, can be analyzed with a number of different approaches and software to investigate structural features as well as local or general evolutionary correlations. In the current study we present a novel accurate methodology which allows to identify relevant high local similarities between viral entire proteome and human proteome.

2 METHODS

Protein sequences from viral and human source were obtained by the RefSeq database to constitute our viral and human proteome databases, which included 82918 and 39037 protein sequences, respectively. The two databases were initially compared by using self-developed proprietary PERL scripts. The comparison was performed in two steps: in the first step all 8 amino acids-long fragments were extracted from the viral database; each virus fragment was then searched in the human database. Any time a full identity was found (8 identities out of 8), extensions at the N- or C- terminus were investigated to verify the occurrence of identity over a longer fragment.

The second step of the analysis was then carried out with BLASTplus (available at the NCBI web site: <http://www.ncbi.nlm.nih.gov/>) to verify the statistical significance of the identities found. All viral fragments selected within the first step were compared to the human proteome database in order to obtain scores and significance evaluation. The “blastp-short” settings were applied, i.e. parameters were optimized for query sequences shorter than 30 residues. Namely, the following settings were applied: word_size=2, gapopen=9, gapextend=1, matrix=PAM30, threshold=16, comp_based_stats=2.

Finally, a final filter based on the virus name was applied to extract the similarities identified by BLAST, concerning protein sequences from specific viruses.

3 RESULTS AND DISCUSSION

During the first step of our search strategy, we extracted all fragments of 8 amino acids from the viral proteins. This minimum length was chosen as a threshold to select fragments with a putative activity as antigenic peptides, as discussed below. These fragments were searched in the human proteome database, and extended when possible to the maximum length of identical segment. The search identified a very high number of non redundant fragments, i.e. 42993 having identical sequence in viral and human proteins, with length of 8 or more amino acids. In many cases, the sequences are characterized by low sequence complexity, i.e., composed by only one or two different residues. The statistical relevance of the similarity involving very short sequences or low-complexity fragments is often considered very low or of difficult evaluation. However, highly repetitive sequences are known to play several key functions: for example, collagen and sericin (the protein composing the silk) are characterized by low-complexity sequences; further, proline-rich peptides have low-complexity but represent key sequences recognized by SH3 domain (Yu, 1994); a 9 residues long poly-arginine has been demonstrated to play a key role in cellular uptake (Wender, 2000); proline-enrichment within peptide-sequence may increase binding to mitochondrial targets (Serasinghe, 2010); leucine-rich or lysine-rich fragments have shown adhesive-, heparin-binding and DNA-binding features. In order to follow a statistics-driven approach, we decided to proceed the analysis using only BLAST-defined statistically significant identities; therefore, in the second step of the present procedure we compared any selected virus-fragment to the human proteome by BLASTplus, using statistical significance settings specifically chosen for short sequences. This second step selected 1076 viral fragments which share statistically significant similarity with human protein sequences and constituted the VISHUM database (Viral Similarities to HUMAN fragments). This is one of the products of the present study; it is a growing database, planned to be updated periodically for the presence of new available viral sequences, as well as extended to sequences extracted from different databases such as GenBank and Uniprot. It presently contains the sequences and the cross-references to the original viral and human databases. Further investigation is now being carried out on the VISHUM database; as an example, we selected from the VISHUM database the herpesvirus fragments and sorted the results by sequence

identity. The herpesvirus selected fragments showing at least 8 continuous identities and at least 90% of sequence identity with human proteins are listed in Table 1. In all cases, with just one exception, the extension of the peptide is longer or much longer than the minimum requested threshold.

While a simpler search strategy could be applied, nevertheless we aimed at finding results significant from both structural and functional point of view, to select fragments with a full identity for a minimum length of 8 amino acids (as minimum requirement for a putative antigenic activity), further extended to the maximum length of identity or similarity; the BLAST evaluation of scores and significance was then applied to each fragment to fix the best level of extension.

To our knowledge this study reports for the first time a systematic approach to investigate structural-functional correlations of viral proteins with human proteins. Choosing 8 consecutive residues, as the minimum requested identity, represented the way to wipe out all short identities difficult to evaluate from the statistical and functional point of view, and allowed to select those with high statistical significance and with an immune-related biological role. In fact, antigen presenting cells (APCs) carry 8-residues long peptides to lymphocytes to elicit the immune response. Additional post-filtering may further improve the fragment selection procedure and the further ongoing investigation of the VISHUM database. For instance only human-specific viral fragments may be selected to be compared to human sequences or both human-specific and human-non-specific fragments can be selected to implement evolutionary-based analyses. Moreover, our study can also be compared to data collected from public databases related to human – virus interactions. We present here data obtained by a post-filtering procedure based on virus name (see Table 1); other filters may be implemented within this strategy, such as, for an example, a 3D-filter to select fragments with a known 3D structure.

The procedure presented in the current study allows to identify viral fragments sharing full identity with human proteins, with a strong statistical significance. The occurrence of highly significant local identities may indicate specific regions with common ancestry or regions where a local evolutionary pressure or a molecular mimicry process may have occurred. A very recent report underlines the role of viral infections as possible trigger of autoimmune disorders in genetically predisposed individuals (Ji, 2010); hence developing VISHUM database and effective procedures to

Table 1: Results generated by the search strategy, filtered for human herpesviruses. The best hits are shown, in terms of percentage of identity (at least 90%).

| Peptide Code | Length | Virus name | BLAST results | | | |
|--------------|--------|----------------------------|---------------|----------|-------------------|------------------------|
| | | | Score | E-value | Number of matches | Percentage of identity |
| 10932 | 11 | Human herpesvirus 5 | 26.2 | 3.8 | 11 | 100 |
| 18349 | 42 | Human herpesvirus 4 type 2 | 87.8 | 2.00E-18 | 42 | 100 |
| 18349 | 42 | Human herpesvirus 4 | 87.8 | 2.00E-18 | 42 | 100 |
| 22175 | 13 | Human herpesvirus 4 type 2 | 30.4 | 0.19 | 12 | 100 |
| 22175 | 13 | Human herpesvirus 4 | 30.4 | 0.19 | 12 | 100 |
| 22175 | 13 | Human herpesvirus 8 | 28.5 | 0.8 | 11 | 100 |
| 28804 | 10 | Human herpesvirus 8 | 25 | 6.4 | 10 | 100 |
| 34315 | 11 | Human herpesvirus 8 | 26.6 | 2.5 | 11 | 100 |
| 34752 | 14 | Human herpesvirus 6 | 35.8 | 0.006 | 14 | 100 |
| 34752 | 14 | Human herpesvirus 6 | 35.8 | 0.006 | 14 | 100 |
| 34752 | 14 | Human herpesvirus 7 | 35 | 0.01 | 14 | 100 |
| 35947 | 12 | Human herpesvirus 8 | 28.5 | 0.66 | 11 | 100 |
| 4113 | 12 | Human herpesvirus 8 | 28.5 | 0.72 | 12 | 100 |
| 41176 | 20 | Human herpesvirus 8 | 44.7 | 1.00E-05 | 20 | 100 |
| 43174 | 14 | Human herpesvirus 8 | 32.3 | 0.055 | 14 | 100 |
| 44922 | 9 | Human herpesvirus 2 | 24.6 | 7.3 | 9 | 100 |
| 46915 | 11 | Human herpesvirus 8 | 31.6 | 0.089 | 11 | 100 |
| 53339 | 31 | Human herpesvirus 4 type 2 | 63.9 | 3.00E-11 | 31 | 100 |
| 53339 | 31 | Human herpesvirus 4 | 63.9 | 3.00E-11 | 31 | 100 |
| 5707 | 9 | Human herpesvirus 5 | 24.3 | 10 | 9 | 100 |
| 57304 | 13 | Human herpesvirus 6 | 28.9 | 0.55 | 12 | 100 |
| 6005 | 8 | Human herpesvirus 2 | 24.3 | 8.3 | 8 | 100 |
| 62235 | 24 | Human herpesvirus 8 | 55.8 | 9.00E-09 | 24 | 100 |
| 55656 | 39 | Human herpesvirus 4 type 2 | 45.8 | 9.00E-06 | 38 | 97 |
| 55656 | 39 | Human herpesvirus 4 | 45.8 | 9.00E-06 | 38 | 97 |
| 58282 | 20 | Human herpesvirus 8 | 43.5 | 4.00E-05 | 18 | 94 |
| 40584 | 15 | Human herpesvirus 3 | 34.7 | 0.014 | 14 | 93 |
| 40584 | 15 | Human herpesvirus 8 | 33.1 | 0.041 | 14 | 93 |
| 34752 | 14 | Human herpesvirus 5 | 34.7 | 0.013 | 13 | 92 |
| 35527 | 15 | Human herpesvirus 8 | 33.5 | 0.028 | 13 | 92 |
| 41386 | 14 | Human herpesvirus 4 | 33.5 | 0.026 | 13 | 92 |
| 41386 | 14 | Human herpesvirus 4 type 2 | 33.5 | 0.026 | 13 | 92 |
| 57304 | 13 | Human herpesvirus 8 | 32.3 | 0.05 | 12 | 92 |
| 57304 | 13 | Human herpesvirus 4 type 2 | 32.3 | 0.05 | 12 | 92 |
| 57304 | 13 | Human herpesvirus 4 | 32.3 | 0.05 | 12 | 92 |
| 35158 | 23 | Human herpesvirus 4 type 2 | 45.4 | 1.00E-05 | 21 | 91 |
| 35158 | 23 | Human herpesvirus 4 | 45.4 | 1.00E-05 | 21 | 91 |
| 57304 | 13 | Human herpesvirus 6 | 27.3 | 1.8 | 11 | 91 |
| 62235 | 24 | Human herpesvirus 3 | 53.5 | 4.00E-08 | 22 | 91 |
| 16190 | 22 | Human herpesvirus 8 | 48.5 | 1.00E-06 | 20 | 90 |
| 40505 | 21 | Human herpesvirus 8 | 45.4 | 1.00E-05 | 19 | 90 |
| 41737 | 10 | Human herpesvirus 4 type 2 | 26.6 | 2.1 | 9 | 90 |
| 41737 | 10 | Human herpesvirus 4 | 26.6 | 2.1 | 9 | 90 |

investigate local protein regions may represent a step forward the improved understating of mechanisms underlying virus-related disorders.

4 CONCLUSIONS

The present study reports a novel procedure to identify relevant similarities among virus and human

proteomes, useful to investigate structural/functional features underlying infectious and non-infectious diseases.

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